

## Supporting Information

### Table of Contents

**Figure S1.** Chemical structures of the C2 and C12 tags

**Figure S2.** Mass spectra of cysteine mutants of IMP-1 showing the incorporation of  $^{13}\text{C}^\zeta$  labelled deuterated indoles and ligation yields with the C2-Y $^{3+}$  tag

**Figure S3.** Superimpositions of [ $^{15}\text{N}, ^1\text{H}$ ]-HSQC spectra of IMP-1 tagged with C2-Ln $^{3+}$  tags

**Figure S4.** Superimpositions of [ $^{15}\text{N}, ^1\text{H}$ ]-HSQC spectra of IMP-1 tagged with C2-Ln $^{3+}$  tags and in the presence of captopril

**Figure S5.** Spectral region of the [ $^{15}\text{N}, ^1\text{H}$ ]-HSQC spectra of Figures S3 and S4 showing the indole N $^{\varepsilon 1}$ -H $^{\varepsilon 1}$  cross-peaks

**Figure S6.** Superimpositions of [ $^{15}\text{N}, ^1\text{H}$ ]-HSQC spectra recorded of IMP-1 N172C ligated with the C12 tag, in the absence and presence of captopril

**Figure S7.** Spectral region of the [ $^{15}\text{N}, ^1\text{H}$ ]-HSQC spectra of Figure S4 showing the tryptophan N $^{\zeta 2}$ -H $^{\zeta 2}$  cross-peaks

**Figure S8.** Pulse sequence of [ $^{13}\text{C}, ^1\text{H}$ ]-HSQC with S $^3\text{E}$  filter designed to select the low-field TROSY component in the  $^{13}\text{C}$  dimension

**Figure S9.** Pulse sequence of [ $^{13}\text{C}, ^1\text{H}$ ]-HSQC with NOE relay

**Figure S10.**  $Q$  factors of  $\Delta\gamma$ -tensor fits to  $^1\text{H}$  PCSs measured of backbone amides for different PDB structures

**Figure S11.** Correlations of  $^1\text{H}$  PCSs measured of backbone amides in the presence versus the absence of captopril

**Figure S12.** PCS isosurfaces obtained with three C2-Tb $^{3+}$  tags plotted for the Trp28 H $^{\varepsilon 1}$  atom

**Figure S13.** Localisation spaces of the Trp28 H $^{\zeta 2}$  and H $^{\varepsilon 1}$  atoms defined by PCSs obtained with Tm $^{3+}$  tags

**Figure S14.** Localisation spaces of the H $^{\zeta 2}$  and H $^{\varepsilon 1}$  atoms of Trp62, Trp88, Trp124 and Trp147

**Table S1.** Pseudocontact shifts of the backbone amide protons of IMP-1 cysteine mutants ligated with lanthanoid tags

**Table S2.** Pseudocontact shifts of the backbone amide protons of IMP-1 cysteine mutants ligated with lanthanoid tags and in the presence of captopril

**Table S3.** Pseudocontact shifts of the  $\text{H}^{\varepsilon 2}$  proton of the tryptophan residues in the IMP-1 cysteine mutants ligated with C2-Tb<sup>3+</sup> or C2-Tm<sup>3+</sup> tags

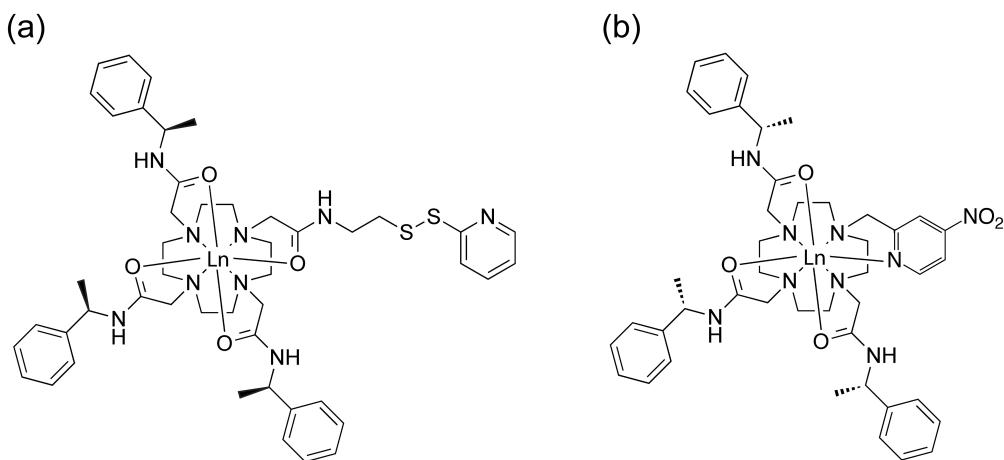
**Table S4.** Pseudocontact shifts of the tryptophan  $\text{H}^{\varepsilon 1}$  proton in the IMP-1 cysteine mutants ligated with C2 and C12 lanthanoid tags

**Table S5.** Pseudocontact shifts of the tryptophan  $\text{H}^{\varepsilon 2}$  protons in the IMP-1 cysteine mutants ligated with C2 and C12 lanthanoid tags and in the presence of captopril

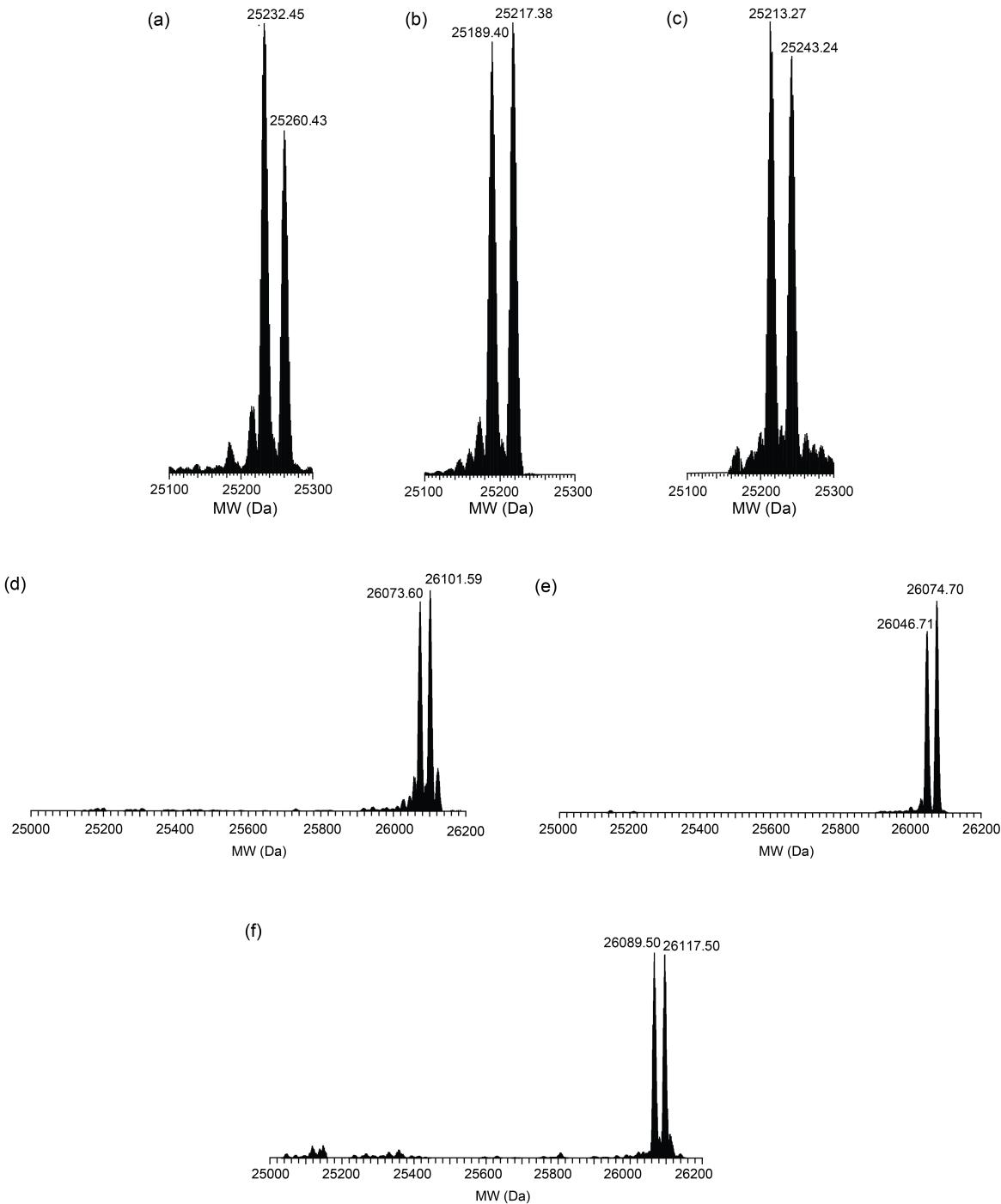
**Table S6.** Pseudocontact shifts of the tryptophan  $\text{H}^{\varepsilon 1}$  protons in the IMP-1 cysteine mutants ligated with lanthanoid tags in the presence of captopril

**Table S7.**  $\Delta\chi$  tensor parameters fitted to different IMP-1 structures using <sup>1</sup>H PCSs of backbone amides

## References

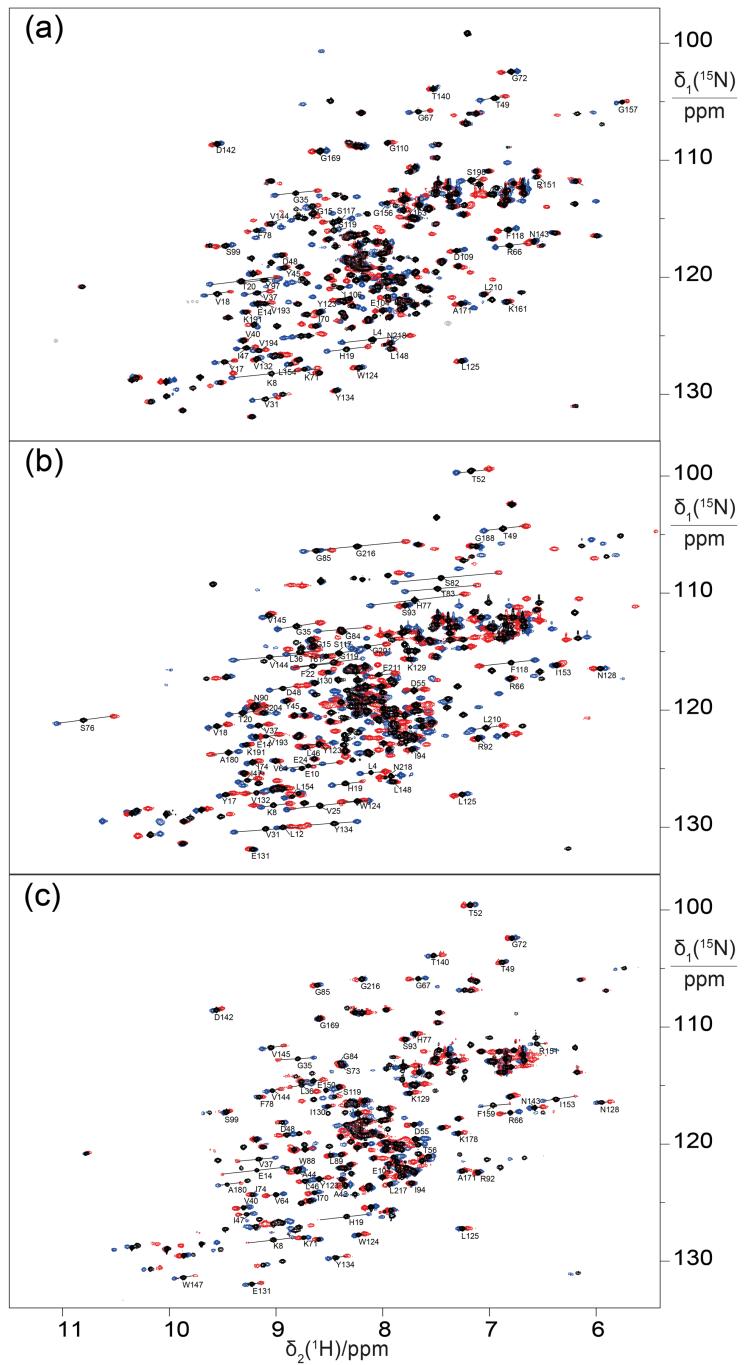


**Figure S1.** Chemical structures of the (a) C2 and (b) C12 lanthanoid tags. The activated disulfide bond in the C1 tag reacts with a cysteine thiol group with formation of a new disulfide bond. In the C12 tag, the nitro group acts as a leaving group in the reaction with a cysteine thiol, leading to the formation of a thioether.

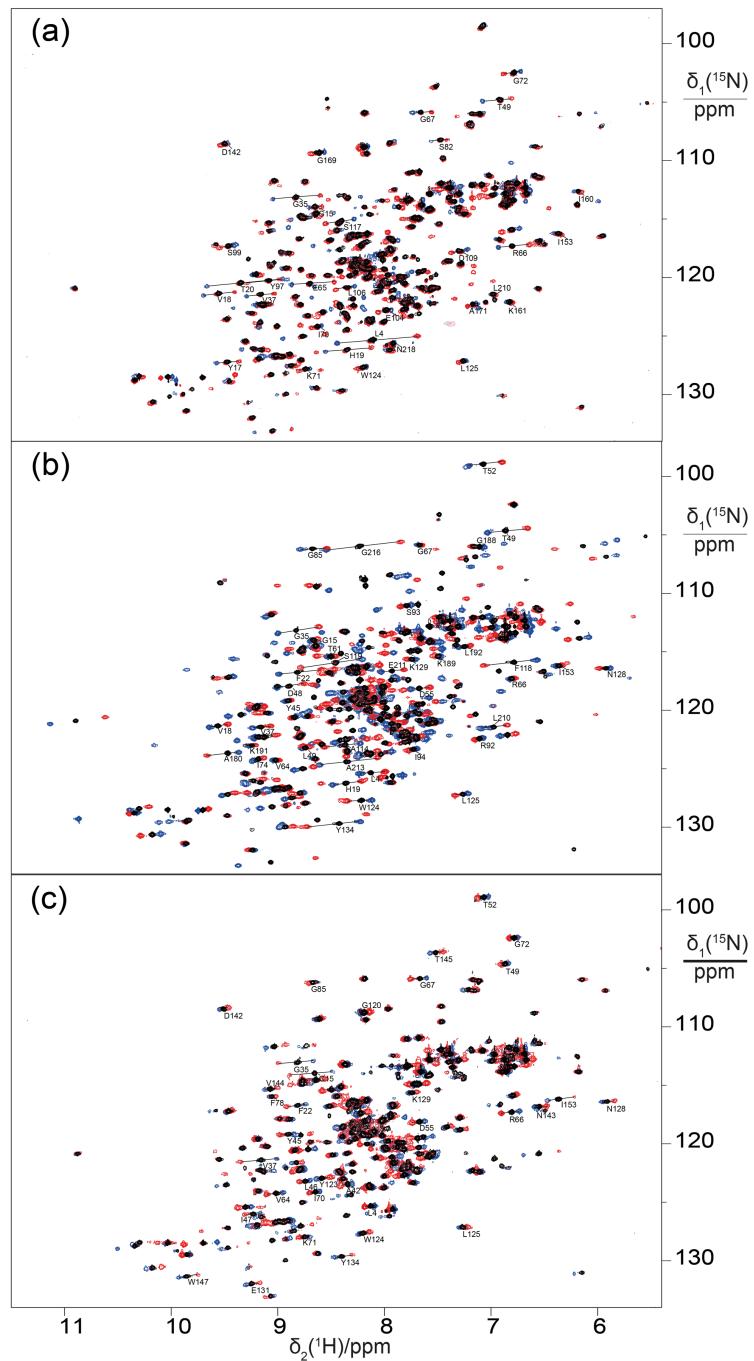


**Figure S2.** Mass spectra of cysteine mutants of IMP-1 showing the incorporation of  $^{13}\text{C}^{\text{C}2}$  labelled deuterated indoles and ligation yields with the  $\text{C}2\text{-Y}^{3+}$  tag. The second major peak in each spectrum is attributed to N-terminal formylation, which adds 28 Da. The ligation with the tag increases the mass by 832 Da. (a) A53C mutant. Calculated mass for six labelled tryptophan residues: 25240.88 Da (6 Da less if only five tryptophans are labelled). (b) N172C mutant. Calculated mass: 25197.86 Da if all six tryptophan residues are labelled and 6 Da less if only five are labelled. (c) S204C mutant. Calculated mass: 25224.88 Da if all six tryptophans are

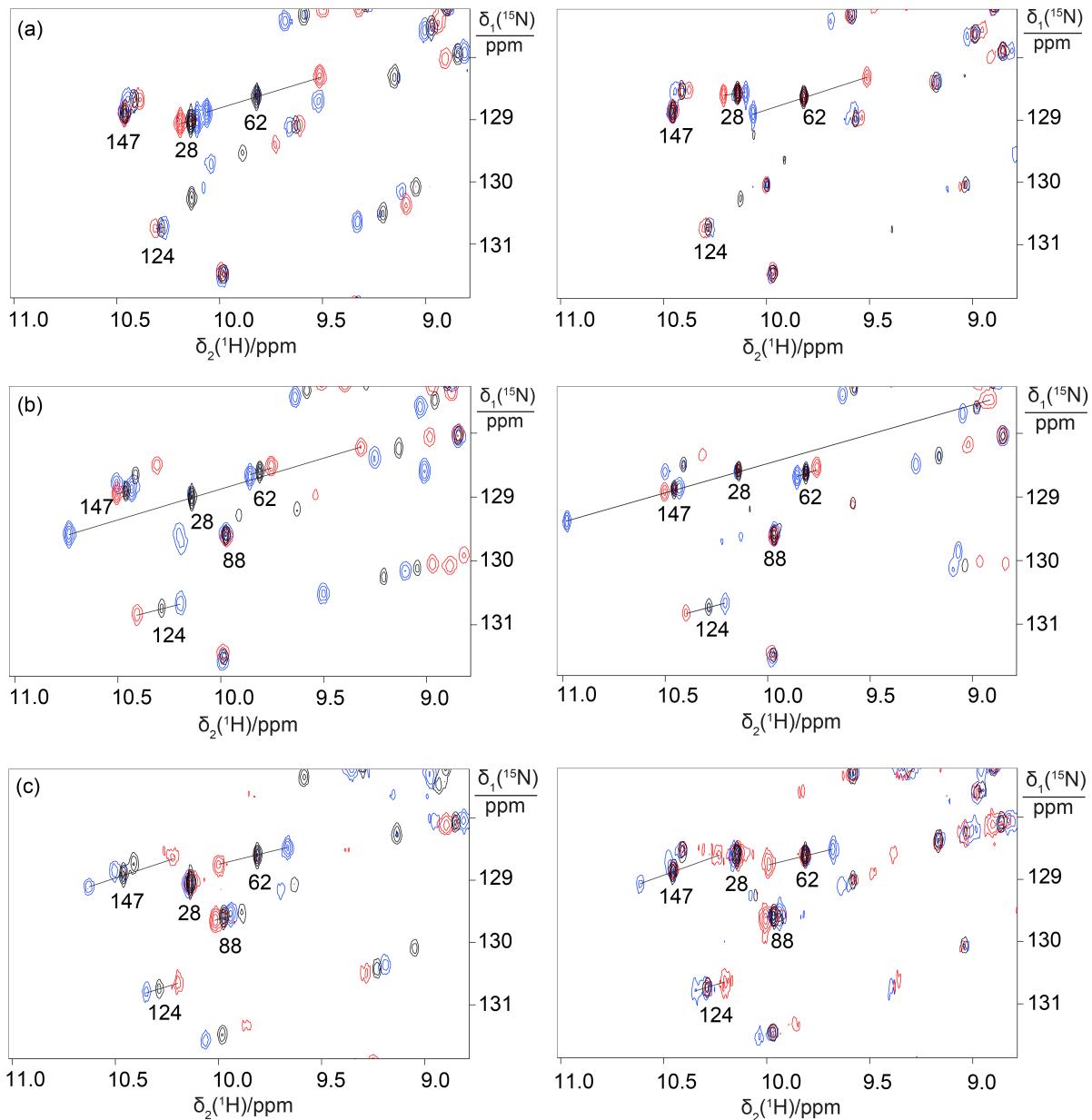
labelled (12 Da less if only four tryptophans are labelled). (d) Same as (a), but after tagging.  
(e) Same as (b) following tagging. (f) Same as (c) following tagging.



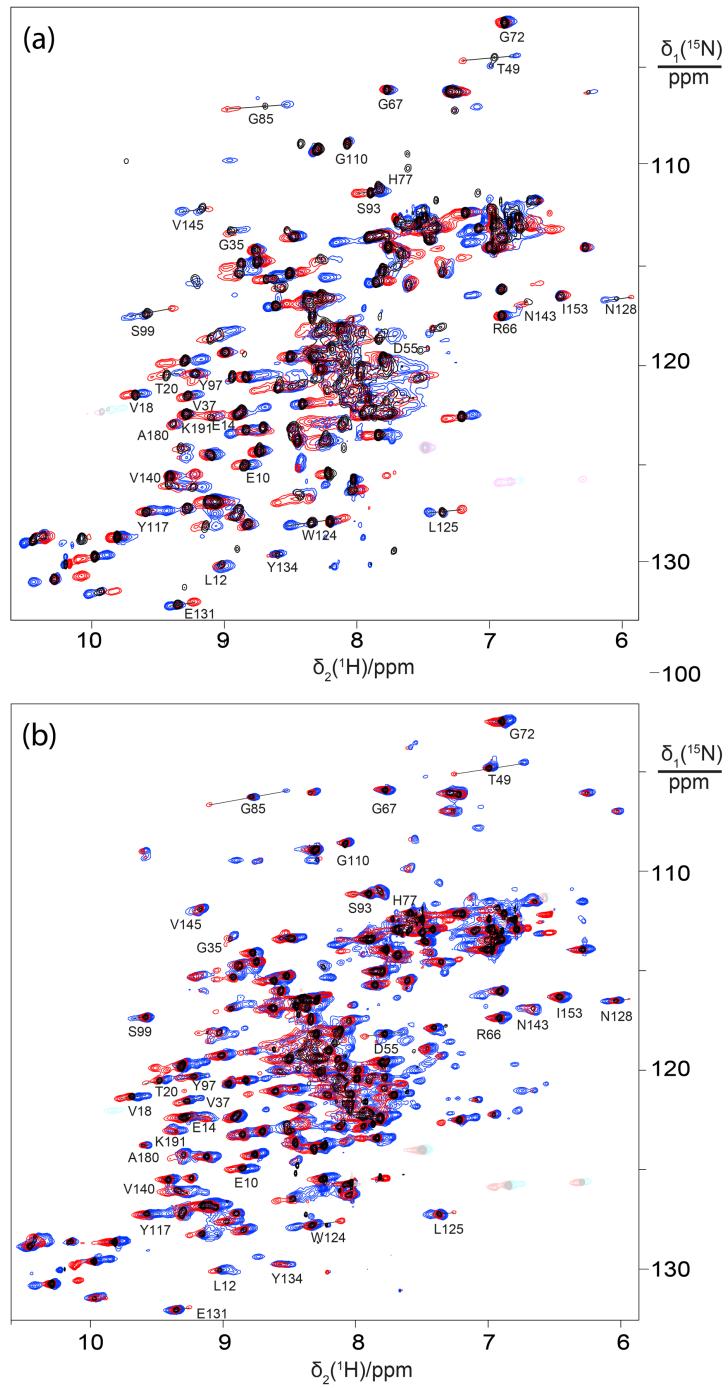
**Figure S3.** Superimpositions of  $[^{15}\text{N}, ^1\text{H}]$ -HSQC spectra recorded of 0.6 mM solutions of uniformly  $^{15}\text{N}$ -labelled IMP-1 tagged with C2-Ln<sup>3+</sup> at three different sites. Spectra with diamagnetic tag (C2-Y<sup>3+</sup>) are plotted in black and the corresponding spectra with paramagnetic tags are shown in red (C2-Tb<sup>3+</sup>) and blue (C2-Tm<sup>3+</sup>). All spectra were measured at 310 K in NMR buffer (20 mM MES, pH 6.5, 100 mM NaCl) on a Bruker 800 MHz NMR spectrometer. Some of the PCSSs are indicated by lines connecting the peaks of paramagnetic and diamagnetic samples. (a) Mutant A53C. (b) Mutant N172C. (c) Mutant S204C.



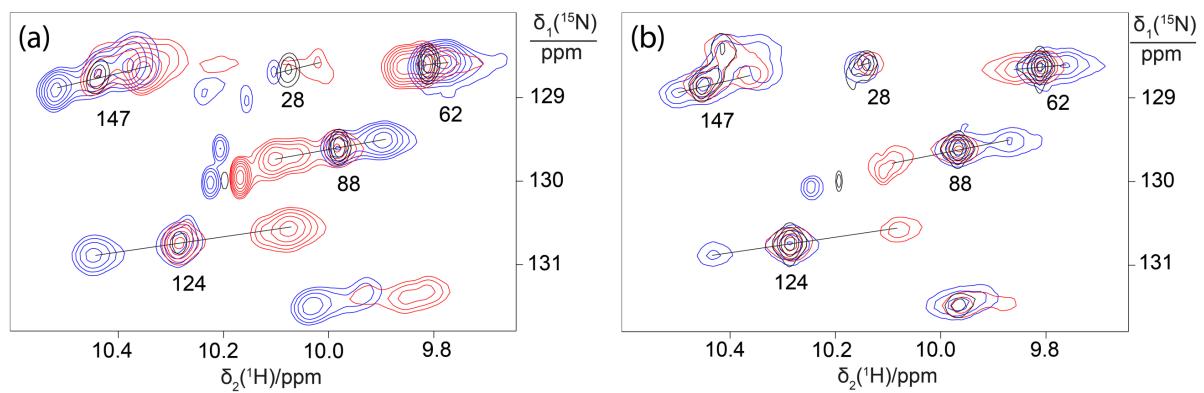
**Figure S4.** Superimpositions of  $[^{15}\text{N}, ^1\text{H}]$ -HSQC spectra recorded of 0.6 mM solutions of uniformly  $^{15}\text{N}$ -labelled IMP-1 tagged with C2-Ln $^{3+}$  at three different sites in the presence 1.5-fold excess of captopril. All other parameters were the same as in Figure 1. (a) Mutant A53C. (b) Mutant N172C. (c) Mutant S204C.



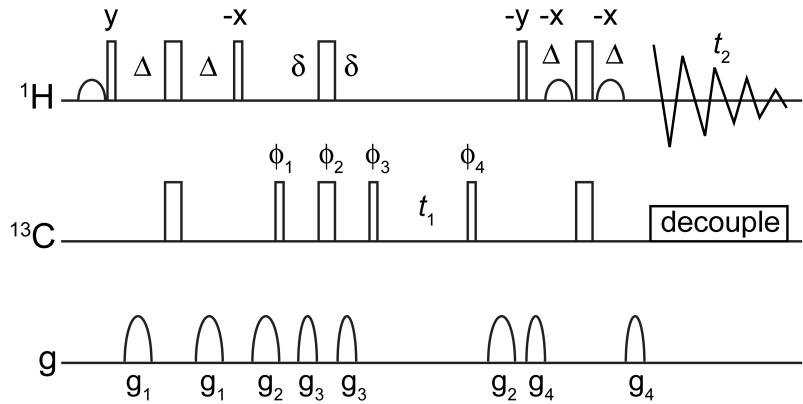
**Figure S5.** Spectral region of the  $[^{15}\text{N}, ^1\text{H}]$ -HSQC spectra of Figures S3 and S4 showing the indole  $\text{N}^{\varepsilon 1}-\text{H}^{\varepsilon 1}$  cross-peaks of the tryptophan residues without (left panels) and with (right panels) captopril. (a) Mutant A53C. (b) Mutant N172C. (c) Mutant S204C. Cross-peaks were assigned by comparison with the  $^1\text{H}$  chemical shifts and PCSs observed in the NOE-relayed  $[^{13}\text{C}, ^1\text{H}]$ -HSQC spectra of Figures 2 and 3. This comparison did not allow identification of the  $\text{N}^{\varepsilon 1}-\text{H}^{\varepsilon 1}$  cross-peak of Trp176 in any of the  $[^{15}\text{N}, ^1\text{H}]$ -HSQC spectra and the cross-peak of Trp88 could not be assigned for the mutant A53C (which is spatially close to Trp88).



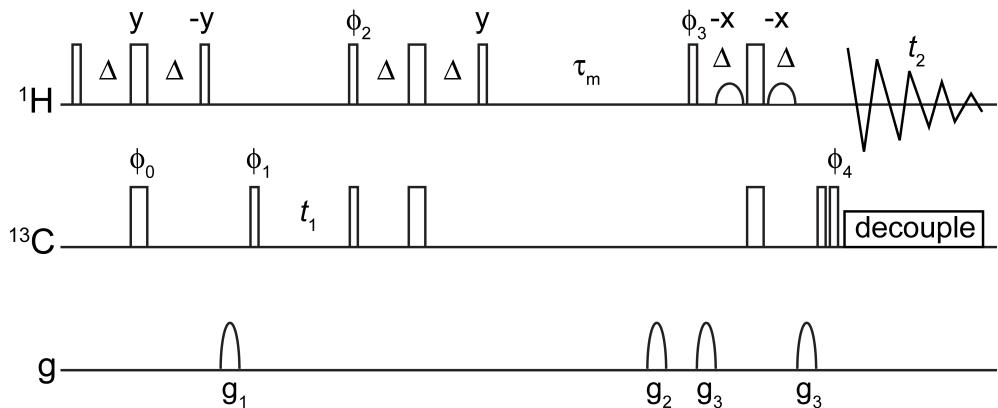
**Figure S6.** Superimpositions of  $[^{15}\text{N},^1\text{H}]$ -HSQC spectra recorded of 0.2 mM solutions of uniformly  $^{15}\text{N}$ -labelled IMP-1 N172C ligated with the C12 tag containing  $\text{Tb}^{+3}$  (red),  $\text{Tm}^{3+}$  (blue) or  $\text{Y}^{3+}$  (black) ions. (a) Spectra of free protein. (b) Spectra in the presence of 1.5-fold excess of captorpril.



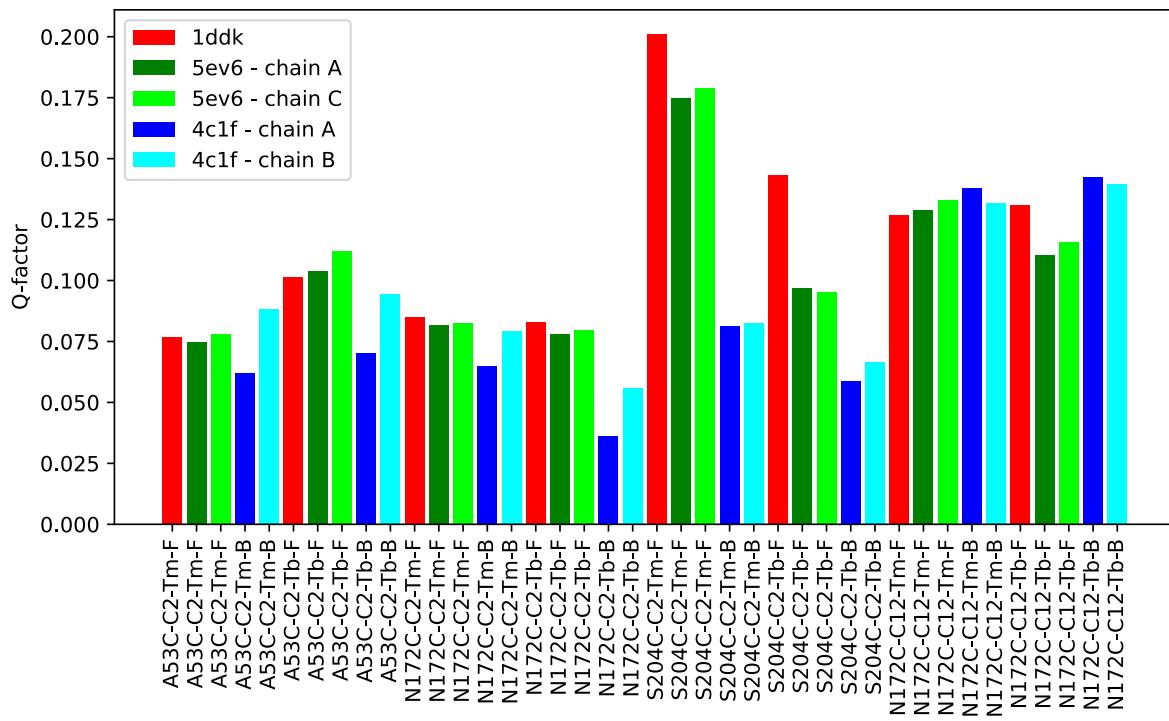
**Figure S7.** Spectral region of the  $[^{15}\text{N}, ^1\text{H}]$ -HSQC spectra of Figure S6 showing the indole  $\text{N}^{\varepsilon 2}-\text{H}^{\varepsilon 2}$  cross-peaks of the tryptophan residues in the absence (left) and presence (right) of captopril. The PCSs of Trp28  $\text{H}^{\varepsilon 1}$  are much smaller than for the same mutant (N172C) labelled with the C2 tag (Figure S5b).



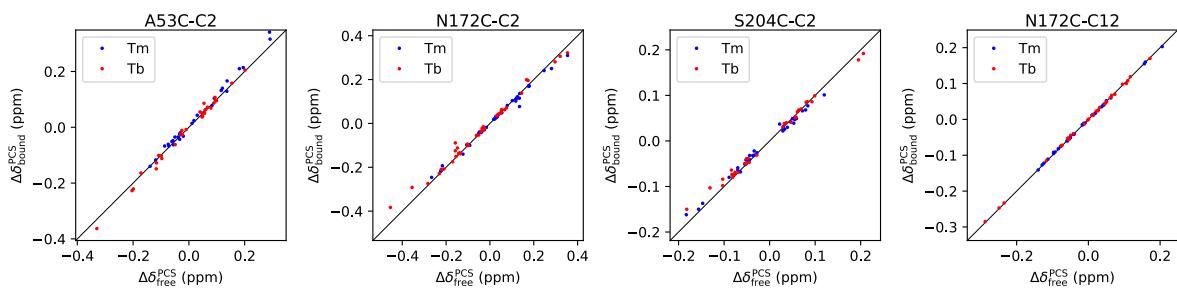
**Figure S8.** Pulse sequence of  $[^{13}\text{C}, ^1\text{H}]$ -HSQC with S<sup>3</sup>E filter designed to select the low-field TROSY component in the  $^{13}\text{C}$  dimension. Narrow and wide rectangles denote 90° and 180° pulses, respectively. Round pulse shapes indicate water-selective 90° pulses of 0.9 ms duration with the shape of the centre lobe of a sinc function. Pulse phases are x unless indicated otherwise. Delays:  $\Delta = 2\delta = 1/(4^1J_{\text{HC}}) = 1.56$  ms. Pulsed field gradients:  $g_1 = 1$  ms at 4 G/cm;  $g_2 = 1$  ms at 19 G/cm;  $g_3 = 0.4$  ms at 3.5 G/cm;  $g_4 = 0.4$  ms at 12.5 G/cm. Phase cycle:  $\phi_1 = \text{x}, -\text{x}$ ;  $\phi_2 = 22.5^\circ$  relative to x;  $\phi_3 = \text{y}, \text{y}, -\text{y}, -\text{y}$ ;  $\phi_4 = \text{x}$ ; receiver = x, -x. Quadrature detection in the  $F_1$  dimension is achieved by decrementing the phase  $\phi_4$  in the State-TPPI manner.



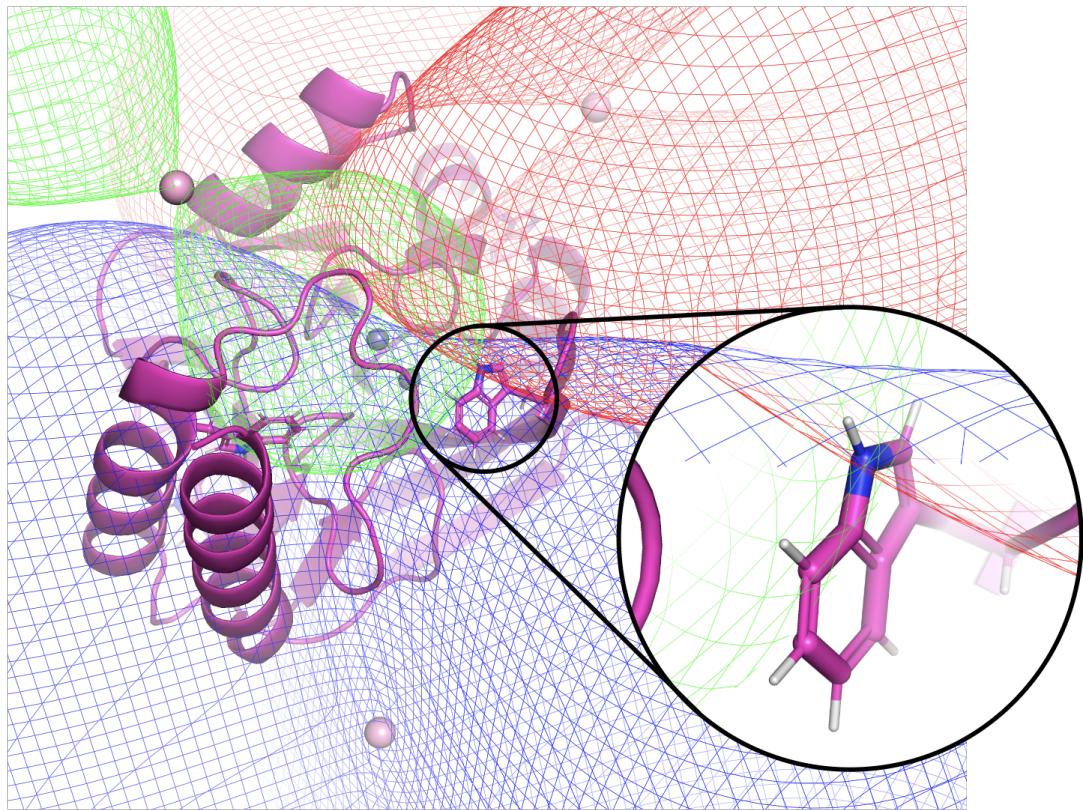
**Figure S9.** Pulse sequence of  $[^{13}\text{C}, ^1\text{H}]$ -HSQC with NOE relay and without decoupling in the  $^{13}\text{C}$ -dimension. The pulse sequence detects both doublet components in the  $^{13}\text{C}$ -dimension, but only the narrow TROSY component is readily detected in large proteins. Narrow and wide rectangles denote 90° and 180° pulses, respectively. Round pulse shapes indicate water-selective 90° pulses of 1 ms duration with the shape of the centre lobe of a sinc function. Pulse phases are x unless indicated otherwise. Delays:  $\Delta = 1/(4^1J_{\text{HC}}) = 1.56$  ms. Pulsed field gradients:  $g_1 = 0.6$  ms at 25 G/cm;  $g_2 = 0.6$  ms at 10 G/cm;  $g_3 = 0.6$  ms at 40 G/cm. Phase cycle:  $\phi_1 = \text{x}, -\text{x}$ ;  $\phi_2 = 2(-\text{x}), 2(\text{x})$ ;  $\phi_3 = 4(\text{x}), 4(-\text{x})$ ;  $\phi_4 = 8(\text{x}), 8(-\text{x})$ ; receiver = x, -x, -x, x, -x, x, x, -x.  $\phi_0$  and  $\phi_1$  are incremented in the State-TPPI manner.



**Figure S10.**  $Q$  factors of  $\Delta\chi$ -tensor fits to  $^1\text{H}$  PCSs measured of backbone amides for different PDB structures using the program Paramagpy (Orton et al., 2020). Hydrogen atoms were added to the crystal structures using the program PyMOL (Schrödinger, LLC, 2015). The x-axis identifies the dataset used; for example, A53C-C2-Tm-F used the  $^1\text{H}$  PCSs of backbone amides in the A53C mutant ligated with the C2-Tm $^{3+}$  tag. “F” and “B” refer to the free protein (in the absence of captopril) and bound protein (i.e. in the presence of captopril).

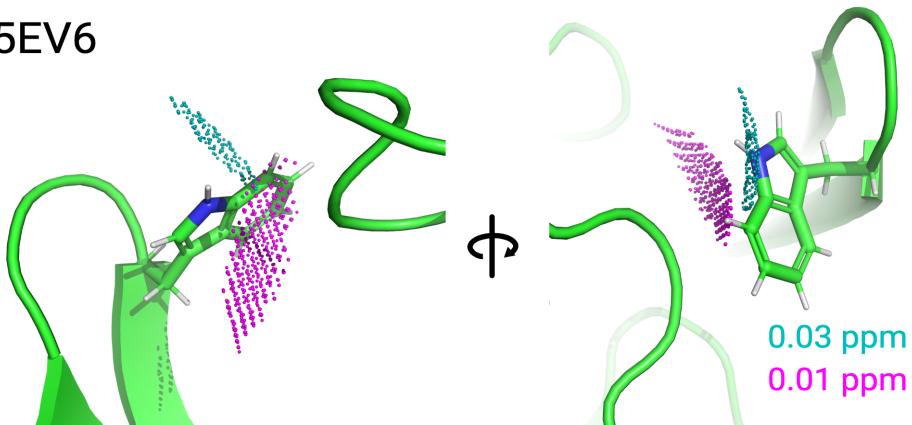


**Figure S11.** Correlations of  $^1\text{H}$  PCSs measured of backbone amides in the presence versus the absence of captopril.

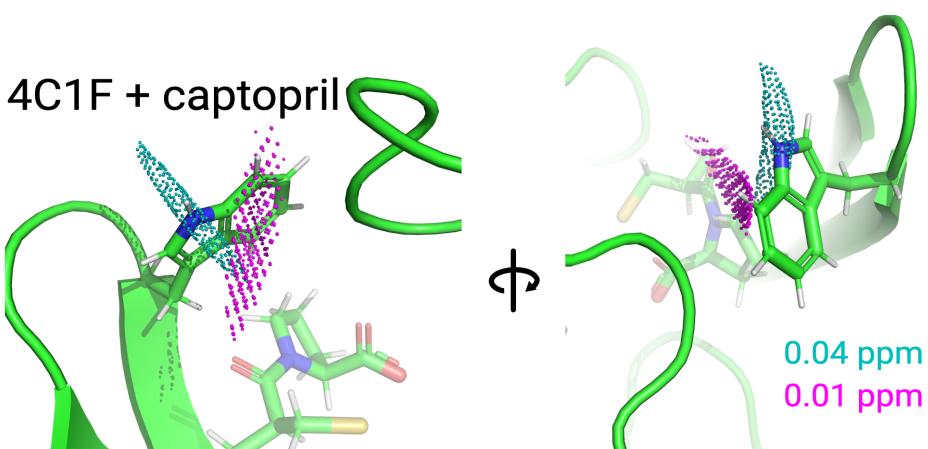


**Figure S12.** PCS isosurfaces plotted for the IMP-1 mutants A53C, N172C and S204C ligated with C2-Tb<sup>3+</sup> tags shown in red, green and blue, respectively, at a contour level equal to the experimental PCS measured for the sidechain H<sup>ε1</sup> atom of Trp28. Data recorded in the absence of captopril and plotted on the structure 5EV6. The protein is shown in a cartoon representation (magenta) with the Zn<sup>2+</sup> ions in the active site shown as grey spheres. The side-chain of Trp28 is highlighted by a stick representation. The figure shows that the three PCS isosurfaces intersect next to the Trp28 H<sup>ε1</sup> atom.

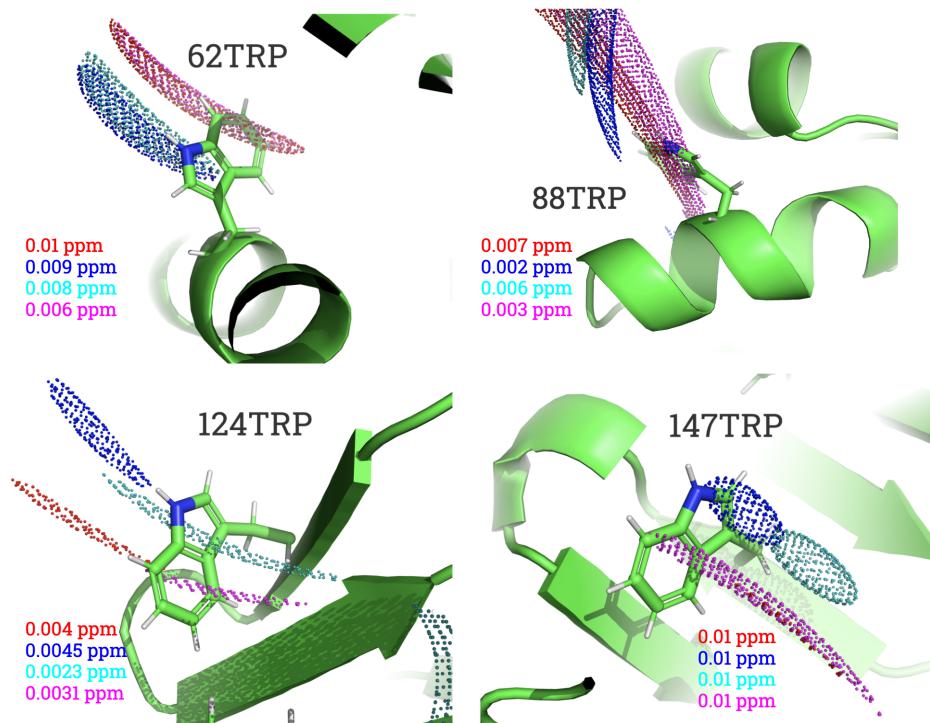
5EV6



4C1F + captoril



**Figure S13.** Localisation spaces of the  $H^{c2}$  and  $H^{e1}$  atoms of Trp28 defined by the PCSs obtained with  $Tm^{3+}$  tags. Data were obtained for the IMP-1 mutants A53C, N172C and S204C ligated with C2- $Tm^{3+}$  tags, both in the absence and presence of captoril. The localisation space of the  $H^{e1}$  atom was restricted further by a fourth PCS obtained for the mutant N172C with the C12- $Tm^{3+}$  tag. Red and blue points trace the localisation spaces of the  $H^{c2}$  and  $H^{e1}$  atoms, respectively. The maximal PCS RMSD values defining the boundaries of the localisation spaces are indicated in ppm. The top panel depicts the localisation spaces determined of the free protein, plotted on the crystal structure 5EV6 and depicted in two different orientations. The bottom panel depicts the localisation spaces determined in the presence of captoril, plotted on the crystal structure 4C1F. The localisation spaces are shown in two different orientations differing by a 180° rotation (about a tilted axis).



**Figure S14.** Localisation spaces of the  $H^{c2}$  and  $H^{e1}$  atoms of Trp62, Trp88, Trp124 and Trp147 in the absence of an inhibitor, plotted on the structure 5EV6. Red and blue points trace the localisation spaces of the  $H^{c2}$  (red) and  $H^{e1}$  (blue) atoms derived from PCS obtained with  $Tb^{3+}$  tags. PCS RMSD cutoffs used to trace the boundaries of the localisation spaces are indicated with the same colours. Corresponding localization spaces obtained with  $Tm^{3+}$  tags are shown in pink and cyan. The localization spaces are less well-defined than for Trp28, as some of the tagging sites produced very small PCSs for these atoms.

**Table S1.** Pseudocontact shifts measured in ppm for backbone amide protons of IMP-1 cysteine mutants ligated with lanthanoid tags.

Residue	A53C-C2		N172C-C2		S204C-C2		N172C-C12	
	Tb	Tm	Tb	Tm	Tb	Tm	Tb	Tm
4LEU	-0.344	0.289	-0.135	0.102	-	-0.044	-	-
8LYS	-0.428	0.365	-0.148	0.120	0.234	-0.188	0.110	-0.097
10GLU	-	-	-0.106	0.083	-	-	-	-
12LEU	-	-	-0.069	0.061	-	-	0.013	-0.017
14GLU	-0.029	0.036	-0.042	0.033	0.321	-0.244	-	-0.009
15GLY	-0.049	0.054	-0.030	0.025	-	-0.183	0.011	-0.013
17TYR	-0.097	0.091	-0.066	0.058	-	-	-	-
18VAL	-0.117	0.120	-0.103	0.081	-	-	0.022	-0.060
19HIS	-0.199	0.181	-0.155	0.125	0.245	-0.207	-	-
20THR	-0.330	0.291	-0.162	0.127	-	-	0.106	-0.097
22PHE	-	-	-0.231	0.178	-	-0.089	-	-
24GLU	-	-	-0.294	0.219	-	-0.027	-	-
25VAL	-	-	-0.417	0.310	-	-0.009	-	-
31VAL	-0.111	0.125	-0.389	0.291	-	-0.043	-	-
34HIS	-	-	-	-	-	-	-	-0.093
35GLY	-0.204	0.195	-0.222	0.178	0.195	-0.147	0.095	-0.113
36LEU	-	-	-	-	0.119	-0.082	0.031	-0.078
37VAL	-0.116	0.116	-0.158	0.135	0.206	-0.156	0.028	-0.058
40VAL	-0.008	0.015	-0.094	0.078	0.069	-0.054	-	-
42ALA	-	-	-	-	0.061	-0.045	-	-
44ALA	-	-	-	-	0.066	-0.040	-	-
45TYR	-0.017	0.034	-0.025	0.023	-	-	-	-
46LEU	-	-	-0.031	0.034	0.056	-0.046	-	-
47ILE	-0.083	0.093	-0.098	0.088	0.080	-0.064	-	-
48ASP	-0.021	0.056	-0.143	0.125	0.031	-0.024	0.074	-0.124
49THR	-0.099	0.137	-0.210	0.177	0.029	-0.034	-	-
52THR	-	-	-0.170	0.136	0.058	-0.048	-	-
55ASP	-	-	-0.099	0.079	0.064	-0.052	-	-
56THR	-	-	-	-	0.069	-0.057	-	-
59LEU	-	-	-	-	0.106	-0.083	-	-
61THR	-	-	-0.041	0.033	0.090	-0.069	-	-
64VAL	-0.229	0.179	-0.026	0.022	0.093	-0.070	0.061	-0.041
66ARG	-0.172	0.136	-0.026	0.017	0.099	-0.077	-	-
67GLY	-0.108	0.083	-	-	0.083	-0.070	0.026	-0.040
70ILE	0.040	-0.020	-	-	0.056	-0.044	0.047	-0.050
71LYS	0.061	-0.033	-	-	0.047	-0.037	-	-
72GLY	0.095	-0.055	-	-	0.037	-0.033	0.001	-0.040
73SER	-	-	-	-	0.018	-0.019	-	-
74ILE	-	-	-0.051	0.054	0.015	-0.017	-	-
76SER	-	-	-0.283	0.247	-	-	0.045	-0.140
77HIS	0.050	-	-0.465	0.409	-0.033	0.014	-	-
78PHE	0.083	-0.054	-	-	-0.028	0.022	-	-
82SER	-	-	-0.537	0.435	-	-	-	-
83THR	-	-	-0.373	0.302	-	-	-	-
84GLY	-	-	-0.256	0.212	0.024	-0.023	-	-
85GLY	-	-	-0.138	0.119	0.032	-0.028	-	-
88TRP	-	-	-	-	0.029	-0.025	-	-
89LEU	-	-	-	-	0.034	-0.026	-	-
90ASN	-	-	0.042	-0.030	-	-	0.172	-0.129
92ARG	-	-	0.037	-0.028	0.022	-0.017	0.117	-0.084
93SER	-	-	0.051	-0.039	0.014	-0.013	0.051	-0.057
94ILE	-	-	0.038	-0.025	0.022	-0.021	0.065	-0.051
97TYR	0.201	-0.139	-	-	-	-	-	-
99SER	0.090	-0.058	-	-	-0.030	0.018	-0.249	0.159
104GLU	0.067	-0.061	-	-	-0.019	0.010	-	-
106LEU	0.094	-0.087	-	-	-0.012	0.006	-	-
109ASP	0.054	-0.074	-	-	-	-	-	-
110GLY	-0.047	-	-	-	-	-	-0.002	-0.016
114ALA	-	-	0.174	-0.123	-	-	-	-
117SER	0.154	-0.119	0.234	-0.182	-	-	-	-
118PHE	0.099	-0.072	0.297	-0.225	-	-	-0.235	0.206
119SER	0.058	-0.050	0.354	-0.266	-0.031	0.021	-	-
120GLY	-	-	-	-	-0.051	0.031	-0.011	0.043
123TYR	0.036	-0.019	0.190	-0.131	-0.081	0.055	-	-

	A53C-C2	N172C-C2		S204C-C2		N172C-C12		
Residue	Tb	Tm	Tb	Tm	Tb	Tm	Tb	Tm
4LEU	-0.414	0.341	-0.138	0.104	0.054	-0.032	-	-
8LYS	-	-	-	-	-	-	0.108	-0.092
12LEU	-	-	-	-	-	-	0.014	-0.015
14GLU	-	-	-	-	-	-	-	-0.010
15GLY	-0.062	0.052	-0.015	0.026	0.236	-0.162	0.011	-0.011
16VAL	-	-	-	0.081	-	-	-	-
17TYR	-0.112	0.104	-	-	-	-	-	-
18VAL	-0.149	0.140	-0.094	-	-	-	0.023	-0.060
19HIS	-0.221	0.210	-0.151	0.121	-	-	-	-
20THR	-0.363	0.316	-	-	-	-	0.100	-0.095
22PHE	-	-	-0.229	0.170	0.098	-0.080	-	-
34HIS	-	-	-	-	-	-	-	-0.091
35GLY	-0.227	0.214	-0.209	0.173	0.178	-0.137	0.098	-0.111

**Table S2.** Pseudocontact shifts measured in ppm for backbone amide protons of IMP-1 cysteine mutants ligated with lanthanoid tags and in the presence of 1.5-fold excess of the inhibitor captopril.

	A53C-C2	N172C-C2		S204C-C2		N172C-C12		
Residue	Tb	Tm	Tb	Tm	Tb	Tm	Tb	Tm
4LEU	-0.414	0.341	-0.138	0.104	0.054	-0.032	-	-
8LYS	-	-	-	-	-	-	0.108	-0.092
12LEU	-	-	-	-	-	-	0.014	-0.015
14GLU	-	-	-	-	-	-	-	-0.010
15GLY	-0.062	0.052	-0.015	0.026	0.236	-0.162	0.011	-0.011
16VAL	-	-	-	0.081	-	-	-	-
17TYR	-0.112	0.104	-	-	-	-	-	-
18VAL	-0.149	0.140	-0.094	-	-	-	0.023	-0.060
19HIS	-0.221	0.210	-0.151	0.121	-	-	-	-
20THR	-0.363	0.316	-	-	-	-	0.100	-0.095
22PHE	-	-	-0.229	0.170	0.098	-0.080	-	-
34HIS	-	-	-	-	-	-	-	-0.091
35GLY	-0.227	0.214	-0.209	0.173	0.178	-0.137	0.098	-0.111

36LEU	-	-	-	-	-	-	-	0.033	-0.076
37VAL	-0.128	0.132	-0.089	0.077	0.192	-0.150	0.026	-0.060	
40VAL	-	-	-	-	0.070	-0.052	-	-	
42ALA	-	-	-	-	0.064	-0.043	-	-	
45TYR	-	-	-0.023	0.021	0.075	-	-	-	
46LEU	-	-	-0.031	0.031	0.051	-0.044	-	-	
47ILE	-	-	-	-	0.085	-0.068	-	-	
48ASP	-	-	-0.137	0.118	-	-	0.070	-0.122	
49THR	-0.102	0.166	-0.208	0.169	0.031	-0.022	-	-	
52THR	-	-	-0.175	0.137	0.052	-0.040	-	-	
55ASP	-	-	-0.101	0.082	0.068	-0.042	-	-	
61THR	-	-	-0.036	0.031	-	-	-	-	
64VAL	-	-	-0.026	0.022	0.086	-0.064	0.060	-0.040	
65GLU	-0.228	0.167	-	-	-	-	-	-	
66ARG	-0.164	0.129	-0.025	0.019	0.099	-0.073	-	-	
67GLY	-0.101	0.078	-0.015	0.011	0.086	-0.059	0.024	-0.040	
70ILE	0.055	-0.032	-	-	0.053	-0.041	0.045	-0.051	
71LYS	0.064	-0.043	-	-	0.041	-0.032	-	-	
72GLY	0.095	-0.063	-	-	0.040	-0.031	0.001	-0.042	
74ILE	-	-	-0.053	0.049	-	-	-	-	
76SER	-	-	-0.274	0.241	-	-	0.044	-0.141	
78PHE	-	-	-	-	-0.031	0.037	-	-	
82SER	-0.068	0.092	-	-	-	-	-	-	
85GLY	-	-	-0.134	0.111	0.038	-0.026	-	-	
90ASN	-	-	-	-	-	-	0.170	-0.127	
92ARG	-	-	0.042	-0.024	-	-	0.119	-0.082	
93SER	-	-	0.053	-0.041	-	-	0.049	-0.055	
94ILE	-	-	0.039	-0.029	-	-	0.067	-0.050	
97TYR	0.205	-0.140	-	-	-	-	-	-	
99SER	0.087	-0.052	-	-	-	-	-0.247	0.160	
104GLU	0.072	-0.051	-	-	-	-	-	-	
106LEU	0.107	-0.067	-	-	-	-	-	-	
109ASP	0.086	-0.061	-	-	-	-	-	-	
110GLY	-	-	-	-	-	-	-0.002	-0.015	
114ALA	-	-	0.196	-0.140	-	-	-	-	
117SER	0.158	-0.117	-	-	-	-	-	-	
118PHE	0.095	-0.067	0.281	-0.218	-	-	-0.233	0.203	
119SER	0.062	-0.035	0.322	-0.246	-	-	-	-	
120GLY	-	-	-	-	-0.050	0.025	-0.013	0.040	
123TYR	-	-	-	-	-0.080	0.047	-	-	
124TRP	0.051	-0.024	0.138	-0.098	-0.050	0.030	-	-	
125LEU	0.037	-0.025	0.071	-0.047	-0.039	0.022	-0.074	0.052	
128ASN	-	-	0.062	-0.039	-0.069	0.039	-0.055	0.016	
129LYS	-	-	0.045	-0.024	-0.039	0.026	-0.055	0.018	
130ILE	-	-	-	-	-	-	-0.040	0.035	
131GLU	-	-	-	-	-0.074	0.049	-0.073	0.040	
134TYR	-	-	0.306	-0.193	-0.084	0.066	-	-	
140THR	-	-	-	-	-0.074	0.049	-	-	
142ASP	0.039	-0.020	-	-	-0.047	0.031	-	-	
143ASN	-	-	-	-	-0.069	0.042	-	-	
144VAL	-	-	-	-	-0.064	0.048	-	-	
145VAL	-	-	-	-	-0.103	0.077	-0.285	0.155	
147TRP	-	-	-	-	-0.098	0.068	-0.115	0.060	
148LEU	-	-	-	-	-	-	-0.044	0.012	
153ILE	-	0.037	-0.023	0.028	-0.150	0.101	-0.052	0.011	
154LEU	-0.023	-	-	-	-	-	-	-	
160ILE	-0.026	0.043	-	-	-	-	-	-	
161LYS	-0.011	0.024	-	-	-	-	-	-	
169GLY	0.069	-0.048	-	-	-	-	-	-	
171ALA	0.050	-0.035	-	-	-	-	-	-	
180ALA	-	-	0.199	-0.100	-	-	-	-	
188GLY	-	-	0.063	-0.035	-	-	-	-	
189LYS	-	-	0.034	-0.018	-	-	-0.072	-	
191LYS	-	-	-0.023	0.032	-	-	-	-	
192LEU	-	-	-0.055	0.049	-	-	-	-	
210LEU	-0.008	0.014	-0.125	0.115	-	-	-	-	
211GLU	-	-	-0.112	0.102	-	-	-	-	
213ALA	-	-	-0.292	0.250	-	-	-	-	
216GLY	-	-	-0.383	0.310	-	-	-	-	
217LEU	0.018	-0.005	-	-	-	-	-	-	

**Table S3.** Pseudocontact shifts measured in ppm for the tryptophan H<sup>c2</sup> protons in the IMP-1 cysteine mutants ligated with C2-Tb<sup>3+</sup> or C2-Tm<sup>3+</sup> tags.

Residue	A53C-C2		N172C-C2		S204C-C2		N172C-C12	
	Tb	Tm	Tb	Tm	Tb	Tm	Tb	Tm
28TRP	0.048	-0.027	-1.228	0.918	-0.046	0.038	-	-
62TRP	-0.228	0.182	-0.056	0.046	0.227	-0.169	-	-
88TRP	0.392	-0.460	-0.014	0.014	0.038	-0.030	-	-
124TRP	0.018	-0.009	0.088	-0.058	-0.085	0.060	-	-
147TRP	-0.004	0.009	0.028	-0.010	-0.318	0.229	-	-
176TRP	0.003	0.004	-	-	-0.124	0.110	-	-

**Table S4.** Pseudocontact shifts measured in ppm for the tryptophan H<sup>c1</sup> protons in the IMP-1 cysteine mutants ligated with C2 and C12 lanthanoid tags.

Residue	A53C-C2		N172C-C2		S204C-C2		N172C-C12	
	Tb	Tm	Tb	Tm	Tb	Tm	Tb	Tm
28TRP	0.078	-0.049	-1.102	0.851	-0.040	0.028	-0.055	0.150
62TRP	-0.305	0.248	-0.057	0.047	0.175	-0.136	0.044	-0.042
88TRP	-	-	-0.010	0.010	0.029	-0.023	0.125	-0.087
124TRP	0.026	-0.018	0.104	-0.069	-0.068	0.049	-	-
147TRP	-0.001	0.003	0.049	-0.024	-0.209	0.150	-0.086	0.073
176TRP	0.004	0.002	-	-	-0.140	0.128	-	-

**Table S5.** Pseudocontact shifts measured in ppm for the tryptophan H<sup>c2</sup> protons in the IMP-1 cysteine mutants ligated with C2 lanthanoid tags and in the presence of captopril.

Residue	A53C-C2		N172C-C2		S204C-C2		N172C-C12	
	Tb	Tm	Tb	Tm	Tb	Tm	Tb	Tm
28TRP	0.073	-0.057	-1.327	0.952	-0.038	0.033	-	-
62TRP	-0.239	0.199	-0.057	0.045	0.219	-0.158	-	-
88TRP	0.138	-0.169	-0.010	0.009	0.039	-0.028	-	-
124TRP	0.012	-0.004	0.090	-0.059	-0.100	0.069	-	-
147TRP	-0.008	0.011	0.029	-0.014	-0.330	0.219	-	-
176TRP	0.002	0.003	-	-	-0.132	0.104	-	-

**Table S6.** Pseudocontact shifts measured in ppm for the tryptophan H<sup>c1</sup> protons in the IMP-1 cysteine mutants ligated with lanthanoid tags and in the presence of captopril.

Residue	A53C-C2		N172C-C2		S204C-C2		N172C-C12	
	Tb	Tm	Tb	Tm	Tb	Tm	Tb	Tm
28TRP	0.108	-0.089	-1.136	0.798	-0.025	0.022	0.001	0.020
62TRP	-0.319	0.268	-0.059	0.046	0.176	-0.129	0.057	-0.066
88TRP	-	-	-0.005	0.002	0.033	-0.025	0.147	-0.117
124TRP	0.020	-0.010	0.118	-0.079	-0.080	0.050	-0.199	0.147
147TRP	-0.002	0.007	0.049	-0.025	-0.210	0.140	-0.085	0.029
176TRP	0.003	0.002	-	-	-0.154	0.118	-	-

**Table S7.**  $\Delta\chi$  tensors fitted to the chain A of the structures 5EV6 and 4C1F using the  $^1\text{H}$  PCSs of backbone amides

PDBID	Captopril	Site	Tag	Ion	$\Delta\chi_{\text{ax}}^{\text{a}}$	$\Delta\chi_{\text{rh}}^{\text{a}}$	x (Å)	y (Å)	z (Å)	$\alpha$ (°)	$\beta$ (°)	$\gamma$ (°)
5EV6	no	A53C	C2	Tm	9.8	4.6	57.396	106.347	30.973	150	130	75
5EV6	no	A53C	C2	Tb	-12.6	-5.9	57.396	106.347	30.973	156	130	79
5EV6	no	N172C	C2	Tm	13.8	3.4	40.922	77.867	25.436	20	32	173
5EV6	no	N172C	C2	Tb	-17.9	-4.4	40.922	77.867	25.436	21	30	173
5EV6	no	S204C	C2	Tm	-6.3	-2.7	67.764	68.485	49.864	52	104	71
5EV6	no	S204C	C2	Tb	9.0	4.1	67.764	68.485	49.864	47	103	65
5EV6	no	N172C	C12	Tm	-10.5	-5.1	43.971	85.261	25.852	57	54	106
5EV6	no	N172C	C12	Tb	13.5	6.4	43.971	85.261	25.852	61	52	112
4C1F	yes	A53C	C2	Tm	10.4	5.5	2.684	31.225	13.791	167	19	114
4C1F	yes	A53C	C2	Tb	-12.8	-6.2	2.684	31.225	13.791	176	17	109
4C1F	yes	N172C	C2	Tm	13.8	4.0	10.430	1.144	23.907	43	136	40
4C1F	yes	N172C	C2	Tb	-17.9	-5.3	10.430	1.144	23.907	43	134	41
4C1F	yes	S204C	C2	Tm	-7.1	-0.9	21.056	0.080	-15.721	119	93	2
4C1F	yes	S204C	C2	Tb	9.6	1.8	21.056	0.080	-15.721	121	94	12
4C1F	yes	N172C	C12	Tm	-9.3	-4.3	9.353	8.135	21.272	85	118	30
4C1F	yes	N172C	C12	Tb	11.7	5.2	9.353	8.135	21.272	82	115	38

<sup>a</sup> In units of  $10^{-32} \text{ m}^3$ . Coordinates and Euler angles are reported with respect to the respective PDB coordinates. Euler angles are given in “ZYX” convention. See Figure S10 for the  $Q$  factors of the fits.

## References

- Orton, H. W., Huber, T., and Otting, G.: Software for fitting magnetic susceptibility tensors using paramagnetic effects measured in NMR spectra, Magn. Reson., 1, 1–12, <https://doi.org/10.5194/mr-1-1-2020>, 2020.
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