

1  
2  
3 Lipid Removal in Deuterium Metabolic Imaging (DMI)  
4 using Spatial Prior Knowledge.  
5

6 Robin A. de Graaf <sup>1,2</sup>, Yanning Liu <sup>2</sup>, Zachary A. Corbin <sup>3</sup>, Henk M. De Feyter <sup>1</sup>

7  
8 Departments of Radiology and Biomedical Imaging <sup>1</sup>, Biomedical Engineering <sup>2</sup> and Neurology <sup>3</sup>  
9 Magnetic Resonance Research Center  
10 Yale University School of Medicine  
11 New Haven, Connecticut, USA  
12  
13

14 Address correspondence to:

15 Robin A. de Graaf, Ph.D.

16 Magnetic Resonance Research Center

17 Department of Radiology and Biomedical Imaging

18 Yale University School of Medicine

19 300 Cedar Street, P.O. Box 208043

20 New Haven, CT 06520-8043, USA

21 Tel: (203) 785-6203

22 Fax: (203) 785-6643

23 E-mail: robin.degraaf@yale.edu  
24  
25  
26

27 Word count: 4,866 (abstract, main body, figure legends)

28 Short title: Robust Processing of Deuterium Metabolic Imaging

1 **ABSTRACT**

2 Deuterium Metabolic Imaging (DMI) is a novel method to generate spatial maps depicting dynamic metabolism  
3 of deuterated substrates, such as [6,6'-<sup>2</sup>H<sub>2</sub>]-glucose, and their metabolic products, like <sup>2</sup>H-lactate. While DMI  
4 acquisition methods are simple and robust, DMI processing still requires expert user interaction, for example in  
5 the removal of extracranial natural abundance <sup>2</sup>H lipid signals that interfere with metabolism-linked <sup>2</sup>H-lactate  
6 formation. Here we pursue the use of MRI-based spatial prior knowledge on brain and non-brain/skull locations  
7 to provide robust and objective lipid removal. Magnetic field heterogeneity was accounted for using DMI-derived  
8 surrogate B<sub>0</sub> and B<sub>1</sub> maps, as well as through subdivision of the skull region into smaller compartments.  
9 Adequate lipid removal with an average suppression of 90.5 ± 11.4 % is achieved on human brain *in vivo* without  
10 perturbation of the metabolic profile in brain voxels, thereby allowing the generation of distinct and reliable  
11 metabolic maps on patients with brain tumors.

12 Key words: Deuterium, Deuterium Metabolic Imaging (DMI), natural abundance lipids, human brain, spatial  
13 prior knowledge

14  
15  
16  
17  
18  
19  
20  
21  
22  
23  
24  
25  
26  
27  
28  
29

## 1. INTRODUCTION

Deuterium Metabolic Imaging (DMI) is a recent method to map the spatial distribution of  $^2\text{H}$ -enriched substrates and their metabolic products in health and disease (De Feyter et al., 2018; Kaggie et al., 2022; Adamson et al., 2023). The most commonly used substrate,  $[6,6\text{-}^2\text{H}_2]$ -glucose, has shown promise to offer unique metabolic insights in brain tumors (De Feyter et al., 2018; Adamson et al., 2023), stroke (Straathof et al., 2021), brown adipose tissue (Riis-Vestergaard et al., 2020), heart (Wang et al., 2021), preeclampsia (Markovic et al., 2021) and a range of tumors outside the brain (Kreis et al., 2020; Veltien et al., 2021). Other  $^2\text{H}$ -enriched substrates, such as  $^2\text{H}_9$ -choline and  $[2,3\text{-}^2\text{H}_2]$ -fumarate, can provide insights into tumor proliferation (Veltien et al., 2021; Ip et al., 2023) and cell death (Hesse et al., 2021), respectively. DMI sets itself apart from other metabolic imaging modalities such as  $^1\text{H}$ ,  $^{13}\text{C}$ , hyperpolarized  $^{13}\text{C}$  and  $^{31}\text{P}$  MRSI, through its simple and robust acquisition methods. The low natural abundance of deuterium eliminates the need for water and lipid suppression, whereas the sparsity of  $^2\text{H}$  MR spectra reduces the magnetic field homogeneity requirements, thereby enabling the acquisition of 3D DMI across the entire human head with high-quality spectra at all locations (De Feyter et al., 2018; Ruhm et al., 2021; Liu et al., 2022; Seres Roig et al., 2022).

Processing of DMI is generally straightforward, with high-quality quantification achieved using least-squares curve fitting with a limited number of Lorentzian lines (De Feyter et al., 2018). DMI of  $[6,6\text{-}^2\text{H}_2]$ -glucose metabolism generally requires four signals for  $^2\text{H}$ -labeled water, glucose (Glc), the combined signal from glutamate and glutamine (Glx) and lactate (Lac). A metabolic map of Lac or a ratio map of Lac/(Lac + Glx) provides high-contrast images of pathological metabolism as described for brain and other tumors (De Feyter et al., 2018), as well as stroke (Straathof et al., 2021). With sufficient sensitivity, as seen near radiofrequency (RF) coil receive elements, natural abundance  $^2\text{H}$  lipid signals originating from the skull will produce a detectable MR signal. While these small lipid signals do not cause the widespread contamination throughout the brain as seen for  $^1\text{H}$  MRSI (Tkáč et al., 2021), they can lead to artifactual intensity in Lac and Lac-Glx ratio maps as the Lac and lipids signals share near-identical chemical shifts. Lipid suppression in human brain MRSI studies can be achieved through pulse sequence modifications (inner volume selection, outer volume suppression, longer echo-times), additional hardware (higher-order gradient coils, crusher coils) or post-processing methods. While pulse sequence modifications and hardware solutions can give excellent lipid suppression (Tkáč et al., 2021), the requirements for DMI are generally modest, making post-processing methods a logical choice while still retaining the simple DMI acquisition method. Many post-processing methods utilize MRI-based prior knowledge on the lipid spatial location to achieve lipid removal and include dual density reconstruction (Hu et al., 1994; Metzger et al., 1999), data extrapolation (Haupt et al., 1996), L2 regularization (Bilgic et al., 2014) and Spectroscopic Localization by Imaging (SLIM, (Hu et al., 1988)) and its variants (Liang and Lauterbur, 1991; Von Kienlin and Mejia, 1991; Bashir and Yablonskiy, 2006; Khalidov et al., 2007; Zhang et al., 2012; Passeri et al., 2014; Adany et al., 2016, 2021).

Here we propose to use the SLIM algorithm (Hu et al., 1988) because it can (1) be applied to standard, 3D phase-encoded DMI without data acquisition modifications, (2) remove extracranial lipids without perturbing the brain metabolic profile and (3) be extended to provide regional brain signals from anatomy-matched compartments. The processing pipeline, including MRI brain/skull segmentation, generation of DMI-based surrogate  $B_0$  and  $B_1$  maps and SLIM-based regional signal removal, can be fully automated and provides a robust and objective tool to accelerate the inclusion of DMI in a clinical MR workflow.

## 2. METHODS

### 2.1. Workflow and algorithm

SLIM belongs to a class of post-processing methods that constrains the MRSI reconstruction with spatial prior knowledge derived from anatomical MRIs. The MRSI acquisition is formulated as a linear model whereby the acquired k-space data  $P$  is the linear sum of region of interest (ROI) specific signals  $C$  weighted by a gradient and position-dependent factor  $G$  according to

$$P = G.C \quad [1]$$

$P$  is the measured  $N_{\text{enc}} \times N_{\text{time}}$  k-space data matrix, with  $N_{\text{enc}}$  representing the total number of phase-encoding gradient combinations (i.e., number of k-space encodings) and  $N_{\text{time}}$  representing the number of complex, spectroscopic time domain points.  $C$  is an  $N_{\text{ROI}} \times N_{\text{time}}$  matrix containing the time-domain signals from each of the  $N_{\text{ROI}}$  compartments, the sum of which comprises the entire object under investigation (e.g., human head, including brain and skull areas). The shape of the ROIs can be arbitrary with  $N_{\text{ROI}} \leq N_{\text{enc}}$ .  $G$  is an  $N_{\text{enc}} \times N_{\text{ROI}}$  encoding matrix describing the amount of signal dephasing across a given compartment  $k$ , during phase-encoding step  $m$  according to

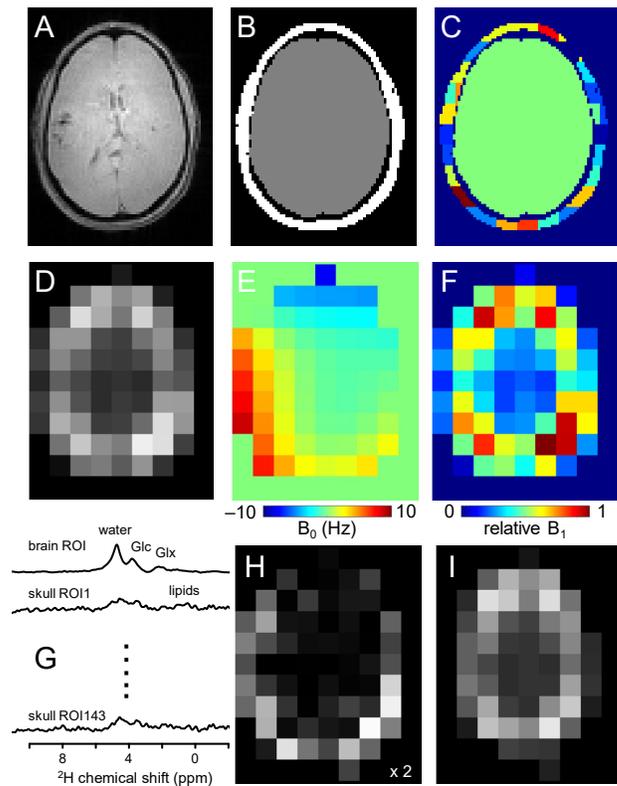
$$G_{k,m} = \sum_{r \in \text{ROI}_k} e^{2\pi i k_m r} \quad \text{with } k = 1 \dots N_{\text{ROI}}, m = 1 \dots N_{\text{enc}} \quad [2]$$

$k_m$  represents the time integral of phase-encoding gradient  $m$ . The SLIM algorithm calculates the ROI-specific signal  $C$  from the measured data  $P$  according to

$$C = G^{-1}.P \quad [3]$$

whereby the inverse of  $G$  can be obtained through singular value decomposition (SVD). If the object under investigation can be decomposed into  $C_{ROI}$  homogeneous compartments, without any requirements on homogeneity *between* compartments, then the SLIM algorithm produces  $C_{ROI}$  signals from the ROIs without any contamination from other compartments.

Fig. 1 summarizes how the outlined algorithm can be utilized to achieve lipid removal in DMI. Based on anatomical MRIs (Fig. 1A), the human head can be segmented into  $N_{brain}$  ROIs and  $N_{skull}$  ROIs (Fig. 1B/C), after which the gradient encoding matrix  $G$  can be calculated for each ROI. With the measured DMI data (Fig. 1D) as input  $P$ , the ROI-specific signals  $C$  can be calculated via Eq. [3] (Fig. 1G). The  $N_{skull}$  signals can be used to reconstruct, via Eq. [1], a skull MRSI dataset (Fig. 1H), which can be subtracted from the original MRSI to provide a skull-free, lipid-suppressed MRSI dataset (Fig. 1I).



**Figure 1 – Lipid removal workflow.** (A) Anatomical MRI and (B) brain and non-brain/skull ROIs. (C) To accommodate heterogeneity in the skull ROI, it is sub-divided into 125-175 ROIs of 3.5 – 8.5 mL. (D)  $^2H$ -water map obtained through numerical integration of each pixel in a  $9 \times 13 \times 11$  DMI dataset. (E)  $^2H$ -water line shift map and (F) relative  $^2H$ -water intensity map. (G) Compartment-specific signals from one brain ROI and 143 skull ROIs are the primary output of the SLIM algorithm. (H) DMI reconstructed from the 143 skull ROI signals, which can be subtracted from (D) the original DMI to yield (I) a skull (and lipid) free DMI dataset.

When all ROIs are homogeneous, the suppression of skull-based signals is expected to be perfect. Unfortunately, the requirement for homogeneous compartments is rarely encountered experimentally due to variations in metabolic composition and  $B_0$  and  $B_1$  magnetic fields across the sample. This heterogeneity is a violation of the linear model (Eq. [1]) and will lead to contamination (or ‘bleeding’) between ROIs (Liang and

Lauterbur, 1993; Von Kienlin and Mejia, 1991) and thus an incomplete removal of skull-based lipids. Several strategies have been developed to address the issue of signal heterogeneity. Multiple methods incorporate prior knowledge on magnetic field heterogeneity from  $B_0$  and/or  $B_1$  map into the SLIM processing pipeline (Bashir and Yablonskiy, 2006; Khalidov et al., 2007; Passeri et al., 2014; Adany et al., 2016). Other methods seek to optimize the k-space encoding scheme to minimize bleeding between ROIs (Von Kienlin and Mejia, 1991; Zhang et al., 2012), whereas another strategy rests on subdividing larger ROIs to decrease the heterogeneity across any one ROI (Adany et al., 2021; Dong and Hwang, 2006).

Here we employ two strategies, namely (1) the subdivision of the non-brain ROI into smaller, more homogeneous compartments and (2) the incorporation of prior knowledge on known  $B_0$  and  $B_1$  magnetic field heterogeneity. Following the segmentation of brain and non-brain ROIs, the latter is sub-divided further (Fig. 1C) by multiplying the ROI with an equidistant 3D grid. A minimum ROI volume is enforced by combining adjacent ROIs for which at least one falls below a minimum threshold. The effect of the ROI size and number of ROIs on the localization performance can be quantitatively evaluated through calculation of the spatial response function (SRF, (Von Kienlin and Mejia, 1991)) according to

$$SRF_k(r) = \sum_{m=1}^{N_{enc}} G_{k,m}^{-1} e^{2\pi i k_m r} \quad \text{with } k = 1 \dots N_{ROI} \quad [4]$$

The SRF is a complex function that depicts the spatial extent of each ROI given a set of k-space encodings, whereby the net SRF contribution of a given ROI is zero across all other ROIs.

The incorporation of  $B_0$  and  $B_1$  magnetic field heterogeneity is achieved through a modification of the encoding matrix (Eq. [2]) according to

$$G_{k,m,n} = \sum_{r \in ROI_k} B_1(r) e^{2\pi i k_m r} \cdot e^{2\pi i B_0(r) t_n} \quad \text{with } n = 1 \dots N_{time} \quad [5]$$

Note that  $G$  essentially converts from a time-independent  $N_{enc} \times N_{ROI}$  matrix to a time-dependent  $N_{enc} \times N_{ROI} \times N_{time}$  matrix, providing unique encoding for each (spectroscopic) time point  $n$ . In Eq. [5]  $B_0(r)$  represents a spatial  $B_0$  magnetic field maps (in Hz) and  $B_1(r)$  represents a relative signal intensity (0 ... 1) with contributions from the transmit  $B_1^+$  and receive  $B_1^-$  magnetic fields. While high-resolution  $B_0$  and  $B_1$  maps can be obtained with routine MR methods for  $^1H$  MRSI, the low-sensitivity DMI data prevents a comparable implementation. The calculation of surrogate  $B_0$  and  $B_1$  maps (Figs. 1E and F) from the measured DMI data will be described next.

## 2.2. Simulations

To evaluate the effectiveness of extracranial lipid suppression and intracranial metabolite retention, simulations were performed on brain and non-brain ROIs segmented from  $T_2$ -weighted MRIs of five human subjects. To limit the overall calculation times, all simulations were performed on a single 2D slice. The brain ROIs were manually segmented into gray matter (GM), white matter (WM) and cerebrospinal fluid (CSF). A pathological ROI (e.g., tumor, stroke) with a random size (72 – 148 % of a nominal MRSI voxel), shape and position was placed inside the brain ROI. With ten random pathology ROI variations per subject, a total of fifty datasets were created for simulation.

Heterogeneity in the  $B_0$  and  $B_1$  magnetic fields can significantly affect the reconstruction performance and simulations were extended with typical distributions found across the human head *in vivo*. As  $^2\text{H}$ -based  $B_0$  and  $B_1$  maps are not readily available, DMI-derived  $B_0$  and  $B_1$  maps were determined on five subjects by measuring the  $^2\text{H}$  water line shift and  $^2\text{H}$  water intensity in each pixel across a 2D slice, respectively. The maps were parameterized with a fourth order 2D polynomial fit to set the  $B_0$  and  $B_1$  distribution average and range. Each of the 50 simulated datasets was constructed with unique  $B_0$  and  $B_1$  distributions, calculated from randomly selected polynomial coefficients characterizing the *in vivo* ranges. To further accommodate  $B_0$  and  $B_1$  heterogeneity, the skull ROI was divided into multiple smaller ROIs. The division was initiated by multiplying the skull ROI by a grid of uniform voxels (e.g., 20 x 20 mm). The resulting skull ROIs were combined with the nearest neighbor ROI when the ROI volume was below a minimum threshold volume (e.g., 40% of a nominal volume). By adjusting the grid size, the number of skull ROIs was varied from 1 to 50 to determine the optimal setting.

Using knowledge on the spatial ROIs,  $B_0$  and  $B_1$  distribution and MRSI k-space encoding, DMI datasets were calculated as a 9 x 13 spatial matrix and 512 complex points acquired over a 1.0 kHz spectral width using a single, well-resolved resonance line per compartment in addition to a water signal present in all compartments. The linewidths were chosen based on previously reported *in vivo* values ((De Feyter et al., 2018),  $T_2 = 30$  ms). Unless specified otherwise, the amplitude per unit volume was identical for all signals. Optional Gaussian noise could be added to the entire 9 x 13 x 512 time domain DMI dataset. SLIM processing (i.e., Eq. [3]) resulted in ROI-specific signals  $C$ , from which the non-brain ROI signals were selected to calculate a non-brain DMI dataset. Subtraction of the non-brain DMI dataset from the experimentally measured DMI produced a lipid-free, brain-only DMI dataset. All data analysis is based on numerical integration of the well-separated spectral MR signals representing water, lipids, and metabolites.

## 2.3. Human studies *in vivo*

All human studies were approved by the Yale University Institutional Review Board. All scans were performed on a 4 T Magnex magnet (Magnex Scientific Ltd.) interfaced to a Bruker Avance III HD spectrometer running on ParaVision 6 (Bruker Instruments). The system was equipped with 67-cm-diameter Magnex gradients capable

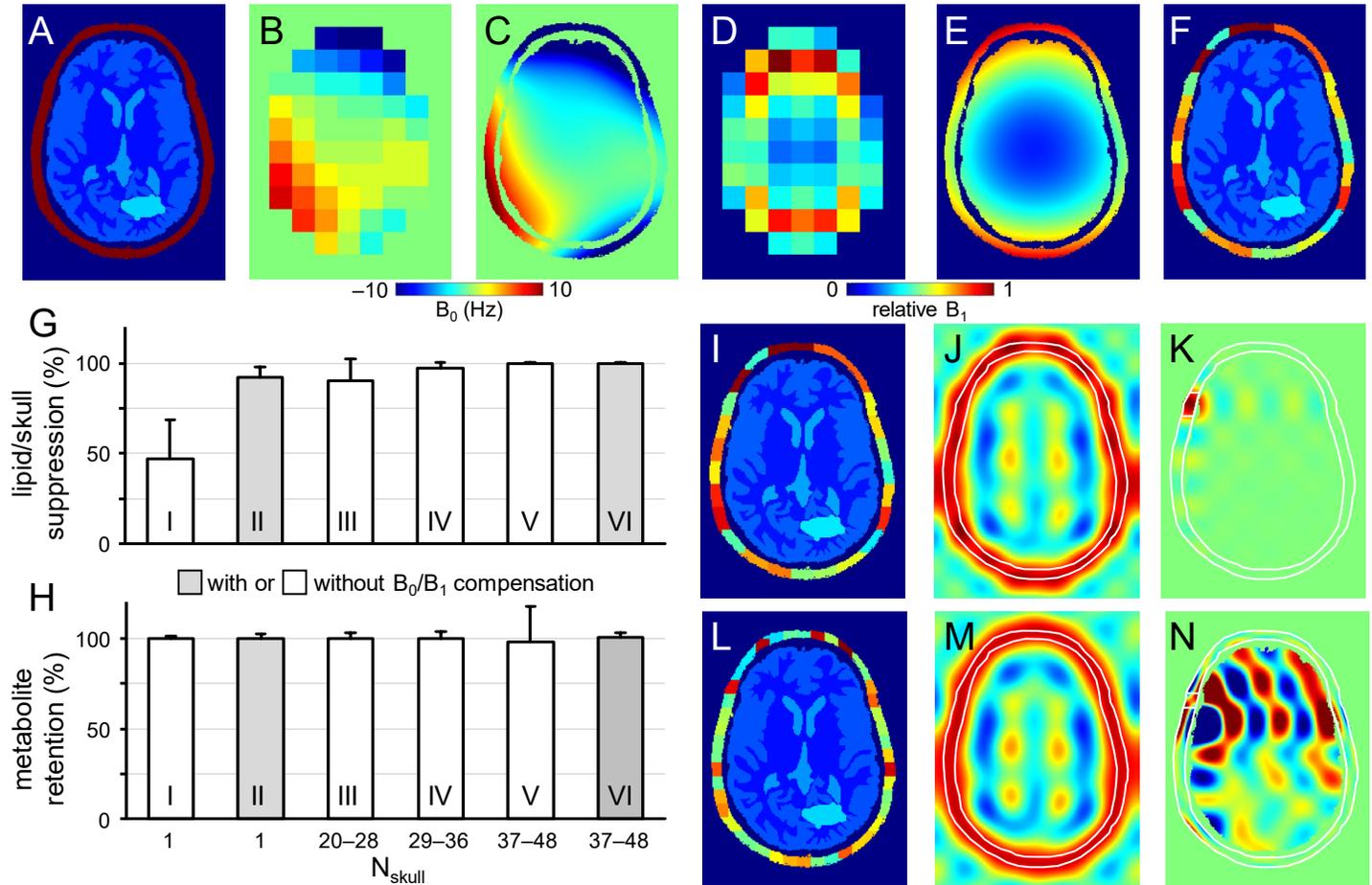
1 of switching 30 mT/m in 1.1 ms. RF transmission and reception were conducted with a 28.5-cm-diameter  
2 transverse electromagnetic (TEM) volume coil tuned to the proton frequency (170.5 MHz) for MRI and shimming.  
3 Deuterium RF reception at 26.2 MHz was achieved with a four-coil phased array that was driven as a single RF  
4 coil during RF transmission. The four  $8 \times 10$  cm rectangular  $^2\text{H}$  array elements were positioned equidistantly on  
5 an  $18 \times 25$  cm elliptical former that was positioned within the  $^1\text{H}$  TEM coil.

6 DMI was acquired with a pulse-acquire method, extended with 3D phase-encoding according to a spherical k-  
7 space sampling pattern. A total of 491 phase encoding steps were acquired to generate a  $9 \times 13 \times 11$  DMI  
8 dataset in circa 27 min with 8 mL nominal resolution (TR = 333 ms, 10 averages). All DMI scans were preceded  
9 by anatomical MRIs and second-order SH shimming over the entire brain. Natural abundance DMI data was  
10 acquired on three subjects without the administration of deuterated substrates. On two subjects, including one  
11 patient with an intracranial glioblastoma tumor,  $[6,6\text{-}^2\text{H}_2]$ -glucose dissolved in water was administered orally  
12 (0.75 g/kg) after which the metabolic profile was sampled with DMI 90 – 120 min later. Brain and non-brain (i.e.,  
13 skull) ROIs were semi-automatically segmented from 3D gradient-echo MRIs (TR/TE = 25/6 ms).

14 Processing of the *in vivo* DMI datasets is similar to that described for simulated data with a few noticeable  
15 differences. Firstly, whereas all *in vivo* DMI data is acquired as a  $9 \times 13 \times 11$  matrix over a field-of-view of  $180 \times$   
16  $260 \times 220$  mm (XYZ), the limited length (80 mm) of the  $^2\text{H}$  RF elements only provided  $^2\text{H}$  signal in 3 to 4 planes  
17 in the Z direction. As a result, ROIs were only generated for spatial positions where the  $^2\text{H}$  water intensity was  
18 at least 5% of the maximum intensity in the entire 3D dataset. Secondly, the brain ROI was not divided into  
19 tissue-specific compartments (e.g., GM, WM, CSF) as simulations (see Results) demonstrated that this step did  
20 not affect lipid suppression and metabolite retention. Thirdly, the skull ROI was divided according to a 3D grid of  
21 nominal DMI voxels without further optimization. ROIs below a minimum volume threshold (40% of nominal)  
22 were merged with nearest neighbor ROIs, resulting in 128-177 skull ROIs across all subjects. Fourthly, signal  
23 quantification is based on non-linear least-squares fitting using a linear combination model of Lorentzian-shaped  
24 signals for  $^2\text{H}$ -labeled water, glucose, glutamate+glutamine (Glx) and lactate.  $^2\text{H}$  MR spectra are fitted with a  
25 (fixed) linear phase roll due to the delayed  $^2\text{H}$  acquisition caused by the phase-encoding gradient. While the  
26 overall linewidths are unconstrained to accommodate magnetic field heterogeneity, the linewidth of metabolites  
27 are linked (within a  $[-2 \dots +5]$  Hz range) to the water linewidth. All spectral fitting was performed in DMIWizard,  
28 a graphical user interface programmed in Matlab (The MathWorks, Natick, MA, USA) and freely available to the  
29 MR community. Finally, in the evaluation of metabolite retention, only pure-brain pixels are considered. For lipid  
30 suppression, partial skull pixels are also considered even though they can lead to an underestimation of the real  
31 suppression.

### 3. RESULTS

Fig. 2A shows the brain and non-brain ROIs used for one of the simulations, together with DMI-derived  $B_0$  (Fig. 2B) and  $B_1$  (Fig. 2D) maps, as well as surrogate  $B_0$  (Fig. 2C) and  $B_1$  (Fig. 2E) obtained by fitting the low-resolution



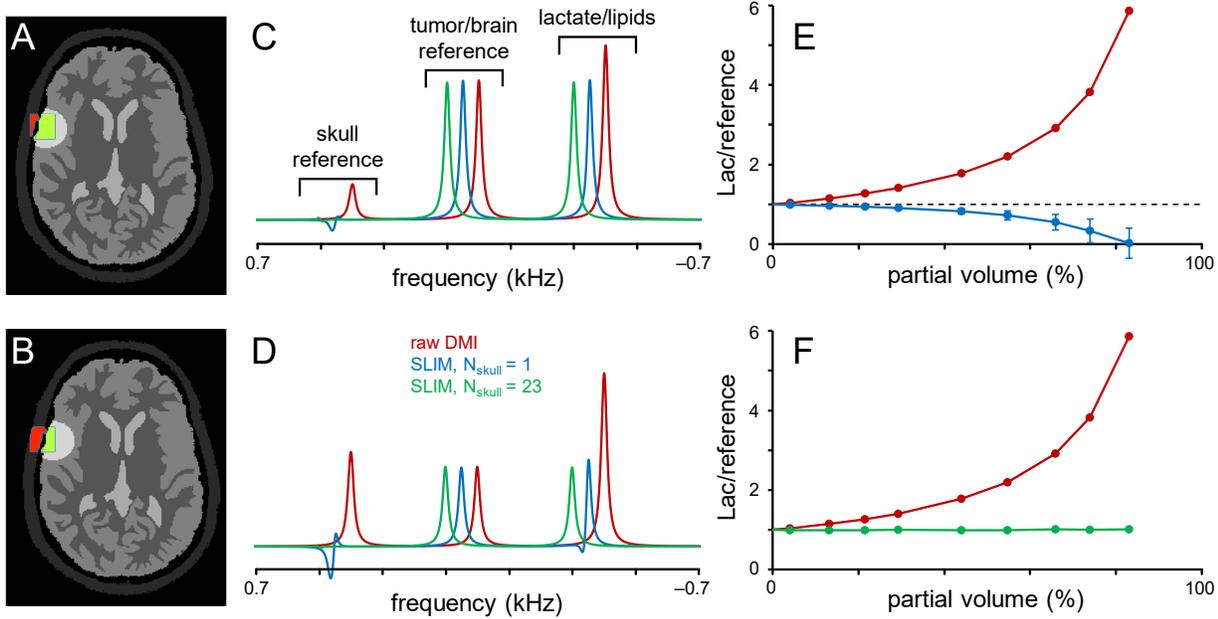
**Figure 2 – Performance of SLIM-based lipid signal removal on phantoms *in silico*.** (A) Brain and skull tissue constellation for one out of 50 permutations. (B) Water shift and (D) intensity maps as extracted from DMI data, providing (C) surrogate  $B_0$  and (E)  $B_1$  maps following low-order polynomial fitting. (F) ROI map with the skull ROI subdivided into 23 smaller compartments (i.e.,  $N_{\text{skull}} = 23$ ) to accommodate signal heterogeneity. (G) Lipid suppression and (H) metabolite retention under different scenarios including  $N_{\text{skull}} = 1$  without (column I) and with (column II)  $B_0$  and  $B_1$  compensation,  $N_{\text{skull}} = 20-28$  (column III),  $29-36$  (column IV),  $37-48$  (columns V and VI) without (columns III-V) and with  $B_0$  and  $B_1$  compensation (column VI). Results for the separate brain ROIs (GM, WM, CSF and pathology) are summarized in Supplemental Figure S2 and essentially minor the whole brain results shown in (H). (I, L) Head constellations with the skull ROI subdivided into (I) 23 and (L) 44 smaller compartments. (J, K, M, N) Spatial response function (SRF) of (J, M) the summed skull ROIs and (K, N) a single skull ROI for  $N_{\text{skull}} = 23$  (J, K) and  $N_{\text{skull}} = 44$  (M, N). All SRFs have the same vertical scale, spanning -1 to +1. While the integrated, *phase-sensitive* SRF intensity of a single skull ROI is zero across the brain ROI for any  $N_{\text{skull}}$ , the integrated, *absolute-valued* SRF intensity will be much larger for  $N_{\text{skull}} = 44$  (N) compared to  $N_{\text{skull}} = 23$  (K). Supplemental Figure S3 gives a summary of the absolute-valued SRF across the brain ROI for  $N_{\text{skull}}$  ranging from 1 to 50.

maps in Figs. 2B/D with third order 2D polynomials. The  $B_0$  magnetic field varied between  $-10$  and  $+10$  Hz over the areas analyzed, whereby the relative  $B_1$  amplitude varied from 0.15 in the center of the brain to 1.00 near the periphery.

1 Fig. 2F shows the subdivision of the non-brain ROI into 23 smaller ROIs, whereby separate simulations  
2 investigated the effect of the number of non-brain ROIs. Supplemental Figure S1 show five additional datasets  
3 used for simulation (out of a total of 50 datasets). All simulations achieved 100% skull-based signal removal and  
4 100% brain signal retention in the absence of  $B_0$  and/or  $B_1$  heterogeneity. Even in the presence of  $B_0$  and  $B_1$   
5 heterogeneity, perfect results were obtained provided that the exact  $B_0$  and  $B_1$  spatial distributions were  
6 incorporated according to Eq. [5]. However, in the presence of  $B_0$  and/or  $B_1$  heterogeneity, skull-based signal  
7 suppression was only  $47 \pm 21$  % when the skull ROI was not subdivided further (Fig. 2G, column I). When the  
8 reconstruction was supplemented with DMI-derived surrogate  $B_0$  and  $B_1$  maps, the skull-based signal  
9 suppression improved to  $92 \pm 6$  % (Fig. 2G, column II). It was observed that the combined effect of incorporating  
10 surrogate  $B_0$  and  $B_1$  maps (6.6 x reduction in residual lipids) was larger than that of the separate  $B_0$  (2.9 x  
11 reduction) or  $B_1$  (1.1 x reduction) maps.  $B_0$  compensation becomes less important, and the overall lipid  
12 suppression improves, as the spectroscopic linewidths becomes broader. Fig. 2G/H (columns III to VI) explores  
13 the effect of subdividing the skull into  $N_{\text{skull}}$  ROIs, with  $N_{\text{skull}}$  ranging from 20-28 (column III), 29-36 (column IV)  
14 and 37-48 (columns V and VI). Without compensation for  $B_0/B_1$  heterogeneity, the skull suppression increases  
15 with increasing  $N_{\text{skull}}$  as the smaller ROIs exhibit less heterogeneity across their volume (lipid suppression equals  
16  $90 \pm 12$ ,  $97 \pm 3$  and  $99 \pm 1$  % for column III - V). However, for higher  $N_{\text{skull}}$  the variability in metabolite retention  
17 increases from 3% (column III) to 4% (column IV) to 20% (column V). Supplemental Figure S2 summarizes  
18 metabolite brain retentions across separate brain ROIs (GM, WM, CSF, and pathology) and essentially mirror  
19 the trends seen across the entire brain (Fig. 2H). This effect can be understood by considering the SRF (Eq. [4])  
20 as demonstrated for  $N_{\text{skull}} = 23$  (Fig. 2I-K) and  $N_{\text{skull}} = 44$  (Fig. 2L-N). In the case of  $N_{\text{skull}} = 23$ , both the overall  
21 skull SRF (Fig. 2J), being the phase-sensitive sum of all  $N_{\text{skull}}$  ROIs, and an exemplary single skull ROI (Fig. 2K)  
22 are well-behaved with the bulk SRF intensity within the intended skull ROI locations. Small contributions inside  
23 the brain have a zero net integral due to phase cancellation. In the case of  $N_{\text{skull}} = 44$ , the overall skull SRF (Fig.  
24 2M) looks like that for  $N_{\text{skull}} = 23$  (Fig. 2J). However, the single skull ROI (Fig. 2N) has the bulk SRF intensity  
25 inside the brain. Even though the integrated SRF intensity across the brain is zero, the strong reliance on phase-  
26 sensitive signal cancellation can lead to alteration of the brain metabolite signal distribution in the presence of  
27 compartmental heterogeneity, such as simulated in Fig. 2H (column V). Supplemental Figure S3 summarizes  
28 the absolute-valued SRF brain contribution for every skull ROI when  $N_{\text{skull}}$  varies from 1 to 50. The SRF  
29 contribution across the brain ROI for any skull ROI is small and well-behaved for  $N_{\text{skull}}$  less than circa 35.  
30 However, when  $N_{\text{skull}} > 35$ , the SRF contribution sharply rises, resulting in localization that is highly dependent  
31 on phase cancellation – a condition that is violated in the presence of heterogeneity. When the  $B_0/B_1$   
32 heterogeneity is compensated with surrogate  $B_0/B_1$  maps (Fig. 2H, column VI), the metabolite retention variability  
33 is greatly reduced (from 20% (column V) to 3% (column VI)) as the large SRF intensity within the brain is properly  
34 phase-canceled. Overall, the simulation results in Fig. 2 indicate that the effects of  $B_0/B_1$  heterogeneity can be  
35 reduced through (1) compensation with surrogate  $B_0/B_1$  maps, (2) subdivision of the skull region into smaller  
36 ROIs, or both, and that the skull ROI subdivision needs to balance improvement in skull suppression with

1 increased variability in metabolite retention. Since the optimal ROI subdivision in Fig. S3 is fairly broad, all 3D  
 2 human DMI data was divided according to a grid of nominal DMI voxels without further optimization. It should be  
 3 noted that the brain ROI was not divided into smaller, tissue-specific ROIs (GM, WM, CSF, pathology) in any of  
 4 the simulations. Preliminary simulations have shown that metabolite retention in separate tissue-specific ROIs  
 5 is only marginally different if the brain is considered as one compartment ( $N_{\text{brain}} = 1, 98.9 \pm 3.7\%$  across all tissue  
 6 ROIs) or as four compartments ( $N_{\text{brain}} = 4, 99.7 \pm 1.5\%$ ). An intuitive explanation for this observation is that even  
 7 though signal leakage between brain compartments does occur, the final step in the algorithm is the  
 8 reconstruction and subtraction of a skull-only DMI dataset, leaving the brain signals largely unperturbed. It should  
 9 also be noted that compartmental differences found *in vivo* are much smaller than those used in simulations (i.e.,  
 10 either signal or no signal) leading to reduced leakage. As the focus of the current work is on lipid removal, the  
 11 decision was made to consider the brain as one compartment.

12 The results in Fig. 2 were primarily focused on lipid/skull suppression and metabolite retention in voxel locations  
 13 with minimal partial voluming. However, one of the most important clinical applications of the proposed algorithm  
 14 is Lac retention in pathologies immediately adjacent to the skull where partial voluming can be significant. Fig. 3  
 15 summarizes Lac retention in the presence of partial voluming by using skull and brain-specific reference signals  
 16 to monitor the amount of partial volume and the quality of lipid suppression.

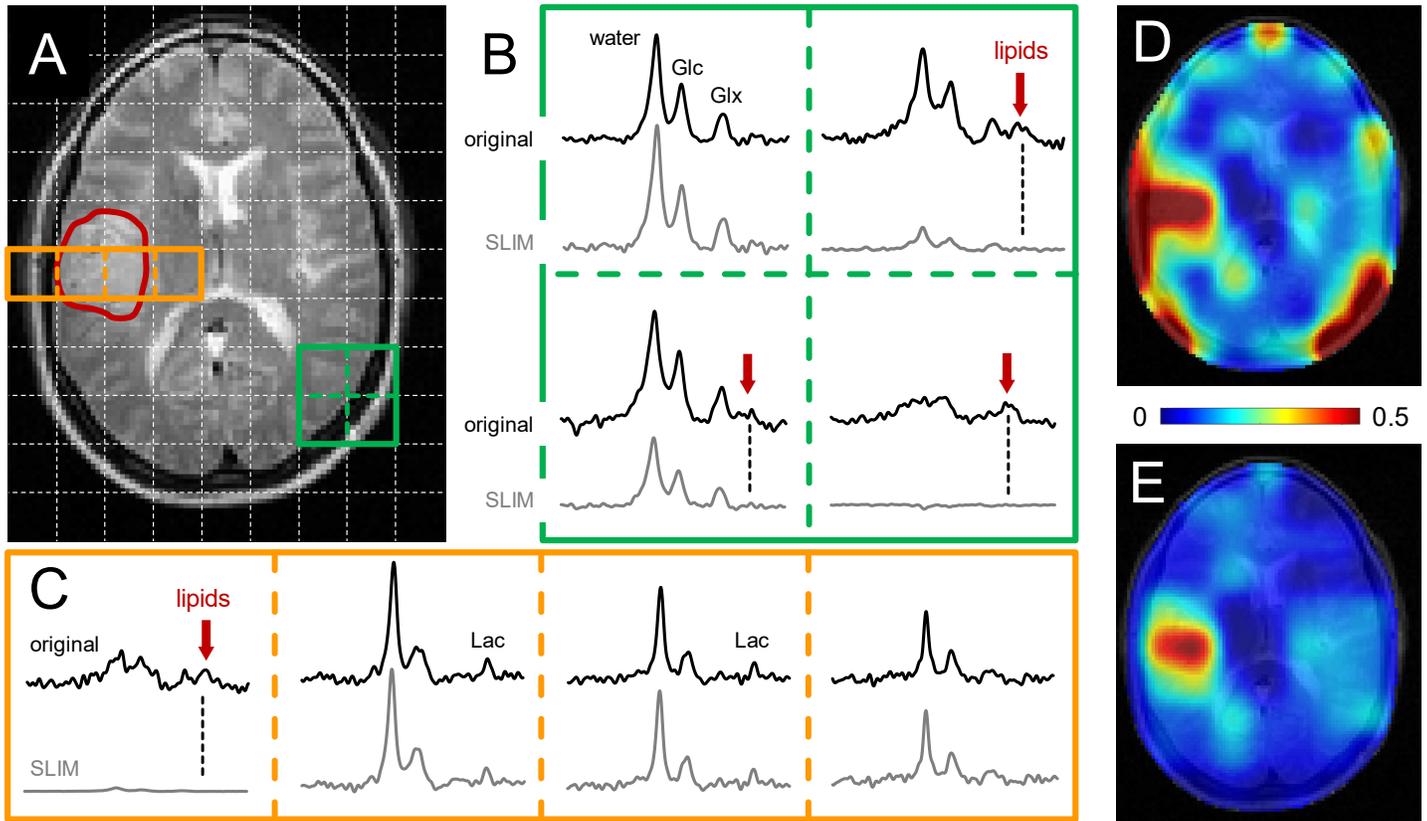


**Figure 3 - Performance of SLIM-based lipid signal removal in the presence of partial volume averaging.** (A, B) Brain, tumor and skull ROI constellation used for all simulations. The amount of partial volume averaging is varied by shifting the ROIs relative to the MRSI grid, giving partial volumes of (A) ~20% and (B) ~55%, whereby partial volume is defined as  $100 \cdot S_{\text{skull}} / (S_{\text{skull}} + S_{\text{tumor}})$  with  $S_{\text{skull}}$  (red in (A, B)) and  $S_{\text{tumor}}$  (green in (A, B)) representing the amount of signal from a reference signal specific for the skull and tumor ROIs, respectively. (C, D) MR spectra obtained from the MRSI voxel indicated in (A, B) with standard FFT processing (red) and SLIM-based processing without (blue) and with (green) skull ROI subdivision. For enhanced visualization, MR spectra are shifted by 50 Hz each. (E, F) Lactate-to-reference ratio as a function of partial volume for standard FFT processing (red) or SLIM processing without (blue) or with (green) skull subdivision. As the lipid removal without skull subdivision (blue) is compromised by  $B_0$  and  $B_1$  magnetic field homogeneity, the lactate-to-reference changes with increasing partial volume. With skull subdivision (green) the lipid removal becomes less dependent on spatial homogeneity, leading to a stable lactate-to-reference ratio, independent of the partial volume. Error bars represent standard deviations calculated over 20  $B_0$  and  $B_1$  magnetic field distributions. Note that the error bars for the red and green curves fall within the marker.

1 The amount of partial voluming was adjusted by spatially shifting the ROI compartments relative to the fixed  
2 MRSI grid. Figs. 3A and B show the ROI constellations to achieve ~20% (Fig. 3A) and ~55% (Fig. 3B) skull  
3 contribution to the indicated MRSI voxel. Figs. 3C and D show MR spectra at the indicated MRSI voxel location  
4 extracted from a 2D MRSI dataset obtained with standard or SLIM processing. With standard, FFT-based  
5 processing (red line) the spectra contain four signals corresponding to a skull reference signal, a brain/pathology  
6 reference signal (e.g., Glx) and a combined signal from Lac and lipids. As the reference signals have equal  
7 amplitudes per unit volume, the intensity of the skull and brain reference signals is indicative of the partial volume  
8 effect. After employing the SLIM algorithm with  $N_{\text{skull}} = 1$  (blue line), the skull reference and lipid signals are  
9 largely removed. However, in agreement with Fig. 2, the removal is incomplete in the presence of  $B_0$  and  $B_1$   
10 heterogeneity, leading to an overestimation of the lipid signal and thus an underestimation of the Lac signal. Fig.  
11 3E shows the Lac-to-brain reference ratio as a function of the partial volume. With standard processing (red line),  
12 the ratio quickly increases as the large lipid contribution is included in the estimated Lac. With SLIM processing  
13 ( $N_{\text{skull}} = 1$ , blue line) the ratio stays close to one for small partial volumes but decreases as the lipid contribution  
14 is overestimated. When the SLIM algorithm is executed with skull subdivision ( $N_{\text{skull}} = 23$ , green line in Figs. 3C-  
15 F) the lipid removal becomes much less sensitive to  $B_0$  and  $B_1$  heterogeneity, leading to near-complete lipid  
16 removal and a correct Lac retention. The Lac-to-brain reference ratio stays close to one for all partial volumes  
17 (Fig. 3F). Note that a partial volume of 90% only contains 10% brain/pathology for which the  $^2\text{H}$  sensitivity would  
18 typically be too low to provide useful data.

19 Fig. 4 shows the performance of SLIM-based skull removal on a patient with a glioblastoma tumor. Fig. 4A shows  
20 a  $T_2$ -weighted spin-echo MRI with the approximate tumor location outlined in red. The white dotted lines indicate  
21 the in-plane DMI grid (8 mL nominal resolution). Fig. 4B shows a grid of  $^2\text{H}$  MR spectra extracted from the 3D  
22 DMI dataset at the location of the green 2 x 2 grid (Fig. 4A). The top spectra (black) represent the original,  
23 measured  $^2\text{H}$  MR spectra, whereas the bottom spectra (gray) are obtained following SLIM-based skull  
24 suppression. Signals from the pure skull voxel (lower right) are completely removed, whereas signals from the  
25 pure brain voxel (upper left) are fully retained. The two remaining voxel locations have both brain and skull  
26 contributions. The extracranial signals (primarily lipids, both also glucose and water) are suppressed, whereas  
27 the intracranial signals (glucose, glutamate, and water) are retained. Fig. 4C shows a grid of  $^2\text{H}$  MR spectra  
28 extracted from the 3D DMI dataset at the location of the red 4 x 1 grid (Fig. 4A) through the tumor region. Only  
29 the skull voxel shows a pronounced lipid signal that is removed with SLIM processing. The remaining three  
30 spectra look visually identical with or without SLIM processing, with the increased lactate signal within the tumor  
31 region remaining visibly unperturbed. However, the removal of extracranial lipid signal has a significant effect on  
32 the resulting Lac/(Lac + Glx) metabolic maps (Fig. 4D/E). Without skull suppression, the lipid signals lead to  
33 several hotspots surrounding the brain (Fig. 4D), including an artifactual elevation of the lactate signals within  
34 the tumor. After SLIM-based skull removal, the lipid hotspots are removed, resulting in a high-contrast 'Warburg  
35 effect' image of aberrant tumor metabolism (Fig. 4E). The lipid suppression across all subjects was  $90.5 \pm 11.4\%$   
36 with 66% and 99% of all skull pixels achieving suppression factors of at least tenfold and threefold, respectively.

1 The signal retentions for Glc and Glx were  $99.8 \pm 7.5$  and  $100.2 \pm 7.8$  %, respectively. Lac retention cannot be  
 2 quantitatively determined due to spectral overlap with lipid signals. However, the outlined simulation results (Figs.  
 3 2 and 3) on high metabolite retentions together with a close visual agreement of Lac in Fig. 4C provide confidence  
 4 that the Lac signal is preserved by the lipid removal algorithm.



**Figure 4 – Performance of SLIM-based lipid signal removal on human brain *in vivo*.** (A) T<sub>2</sub>-weighted spin-echo MRI of a patient harboring a glioblastoma brain tumor with the approximate location outlined in red. The white dotted lines indicate the in-plane DMI grid (8 mL nominal resolution, circa 30 min total scan time). (B) 2 x 2 x 1 sub-grid extracted from the 9 x 13 x 11 DMI dataset showing the original (black, top) and SLIM-processed (gray, bottom) <sup>2</sup>H MR spectra. In three spectra a clear natural abundance lipid signal (red arrow) is present in the original data which is completely removed following SLIM processing. (C) 4 x 1 x 1 sub-grid covering tumor and adjacent skull and brain tissue extracted from the 9 x 13 x 11 DMI dataset. Only the skull voxel shows a pronounced lipid signal that is removed with SLIM processing. While the remaining three spectra looks visually identical with or without SLIM processing, the removal of the extracranial lipid signal has a significant effect on the (D, E) resulting Lac/(Lac + Glx) metabolic maps.

5

#### 6 4. DISCUSSION

7 The use of spatial prior knowledge in DMI processing allowed the removal of extracranial lipid signals in datasets  
 8 acquired of human head to below the spectral noise level. Similar approaches have previously been used for  
 9 lipid signal removal in <sup>1</sup>H MRSI (Dong and Hwang, 2006; Adany et al., 2016). However, <sup>1</sup>H MRSI requires lipid  
 10 signal suppression factors well in excess of 100. This puts much higher demands on accurate prior knowledge  
 11 and signal homogeneity and may ultimately place a limit on the robustness for use on <sup>1</sup>H MRSI data. In contrast,

1 the low natural abundance of deuterium leads to much lower amplitude lipid signals in the DMI data which were  
2 adequately removed with suppression factors below 10, thereby eliminating ambiguities in detecting elevated  
3 lactate levels. While lipid removal is desirable for all spatial positions to eliminate 'hot spots' in the resulting  
4 lactate map (see Fig. 4D), it is especially prudent for skull areas immediately adjacent to the pathology (e.g.,  
5 brain tumor) where partial volume effects would lead to a distorted lactate signal. The SLIM algorithm is based  
6 on spatial prior knowledge from high-resolution MRIs and is intrinsically suitable to separate skull from brain  
7 signals even in the presence of partial volume effects caused by the lower DMI resolution. This feature was  
8 systematically investigated with simulations (Fig. 3) and experimentally demonstrated (Fig. 4C) and confirmed  
9 constant lipid suppression performance in the absence or presence of partial volume effects.

10 The basic SLIM algorithm that uses two ROIs (one brain, one skull) without compensation for  $B_0$  and  $B_1$  magnetic  
11 field heterogeneity only provides modest lipid suppression ( $47 \pm 21$  % in simulations). Subdividing the skull ROI  
12 into smaller compartments quickly improves the lipid suppression to  $>90$ ,  $>97$  and  $> 99.5$  % for  $\sim 25$ ,  $\sim 35$  and  
13  $\sim 45$  ROIs, respectively. However, improved lipid suppression is accompanied by more variable metabolite  
14 retention as the SRF outside any given ROI becomes more dominant and localization increasingly relies on  
15 phase cancelation. The inclusion of  $B_0$  and  $B_1$  maps greatly improved the lipid suppression (to  $92 \pm 6$  % in  
16 simulations for one brain and one skull ROI) without increased variability in metabolite retention. The use of  
17 surrogate  $B_0$  and  $B_1$  maps based on the  $^2\text{H}$  water shift and intensity provide a practical solution to absence of  
18 high-resolution maps. Based on these results, it is recommended that  $B_0$  and  $B_1$  compensation should be used  
19 whenever possible, ideally in combination with a moderate subdivision of the skull ROI. It should be noted that  
20 while  $B_1$  heterogeneity was treated as a nuisance in the current implementation, it is possible to use  $B_1$  receive  
21 profiles for signal encoding to accelerate and/or improve data acquisition (An et al., 2011). While the average  
22 metabolite retention in the human brain *in vivo* is essentially 100%, the standard deviations on Glc and Glx are  
23 somewhat higher than anticipated compared to simulation results (Fig. 2). Several factors can explain the  
24 increased variability. Firstly, removal of extracranial signals, including water and Glc, will perturb brain signals  
25 as voxel bleeding due to the MRSI point spread function is also removed. Secondly, MRSI voxels with significant  
26 partial volume effects will be perturbed as extracranial contributions are selectively removed. Thirdly,  
27 compensation of  $B_0$  and/or  $B_1$  heterogeneity with DMI-based surrogate  $B_0$  and  $B_1$  maps will greatly decrease the  
28 variability in metabolite retention. However, since the surrogate maps are only an approximation of the actual  $B_0$   
29 and  $B_1$  heterogeneity, the compensation is necessarily incomplete, thus leading to some residual variability.  
30 Finally, the SNR of *in vivo* DMI data is generally low, such that small perturbations in signal shape or noise level  
31 can lead to large relative changes in fitted signal amplitude.

32 In the current study the anatomical MRI and DMI datasets were acquired with the same RF coil assembly without  
33 subject movement. As a result, no spatial co-registration between MRI and DMI was necessary, and the MRI-  
34 derived spatial prior knowledge was accurate. However, in the presence of significant subject movement, or  
35 when using MRIs acquired from a separate scan session (e.g., clinical MRIs), a mismatch between MRI-derived  
36 spatial prior knowledge and DMI can lead to greatly diminished lipid suppression performance and even a

1 distorted metabolite profile. Simulations (data not shown) have indicated that a mismatch of 10-20% of the  
2 nominal DMI voxel size can still retain a high level of lipid suppression, but that performance quickly degrades  
3 with larger mismatches. It is therefore recommended to acquire MRIs and DMI during the same session with an  
4 identical subject position. Spatial co-registration between MRI and DMI is possible but was not further pursued  
5 in this study. When subject movement during MRI/DMI scanning is anticipated, additional measures (e.g., motion  
6 tracking, additional immobilization) need to be taken to ensure accurate spatial prior knowledge.

7 The SLIM implementation as presented here for DMI can be modified and extended in several ways to enhance  
8 the immunity to signal heterogeneity and improve the reliability. Sub-division of the skull ROI can be optimized  
9 further. The number of ROIs and ROI shapes can be based on (1) optimizing the SRF contributions, or (2) DMI-  
10 based lipid maps or (3) high-resolution MRI-based lipid maps. The SLIM algorithm is also ideally suited to allow  
11 regional analysis of brain ROIs during which  $^2\text{H}$  MR spectra can be obtained from anatomy-matched ROIs  
12 without partial volume effects. Development of automated definition of anatomical (GM, WM) and pathological  
13 (tumor core, tumor rim, FLAIR-enhanced) ROIs is currently in progress.

14 Similar levels of lipid suppression could have been achieved with alternative methods, such as dual density  
15 reconstruction (Hu et al., 1994; Metzger et al., 1999), data extrapolation (Haupt et al., 1996) or L2 regularization  
16 (Bilgic et al., 2014). Whereas L2 regularization and data extrapolation can operate on standard MRSI data, dual  
17 density reconstruction requires a modification of the data acquisition scheme, such that high k-space coordinates  
18 are sampled in addition to the standard low k-space coordinates typically acquired for MRSI. L2 regularization is  
19 based on the conditions of spatial and spectral orthogonality in which the desired and nuisance signals originate  
20 from spatially distinct compartments without spectral overlap. As the spectral overlap requirement cannot be  
21 fulfilled for lipids and lactate, L2 regularization will remove brain lactate signals, thus making it unsuitable for  
22 DMI. Like SLIM, data extrapolation only requires spatial separability of signals and would thus also be applicable  
23 to DMI. However, in the current investigation we focused on SLIM-based processing as the algorithm is readily  
24 extended to obtain signal from anatomy and/or pathology-based ROIs.

## 26 **CODE AVAILABILITY**

27 The Matlab code to process 3D DMI data, referred to as DMIWizard, is freely available for download from the  
28 Yale website or via GitHub (<https://github.com/radegraaf/DMIWizard>).

## 29 **COMPETING INTERESTS**

30 At least one of the (co-)authors is a member of the editorial board of Magnetic Resonance.

## 31 **ACKNOWLEDGEMENTS**

1 This research was funded, in part, by NIH grants NIBIB R01-EB025840 and R01- EB033764 and by CTSA grant  
2 KL2 TR001862 from the National Center for Advancing Translational Science (NCATS), component of the NIH  
3 and NIH roadmap for Medical Research.

#### 4 REFERENCES

- 5 Adamson, P. M., Datta, K., Watkins, R., Recht, L., Hurd, R., and Spielman, D.: Deuterium Metabolic Imaging (DMI) for 3D  
6 mapping of glucose metabolism in humans with central nervous system lesions at 3T, *Magn Reson Med*, 2023.
- 7 Adany, P., Choi, I. Y., and Lee, P.:  $B_0$ -adjusted and sensitivity-encoded spectral localization by imaging (BASE-SLIM) in the  
8 human brain *in vivo*, *Neuroimage*, 134, 355-364, 10.1016/j.neuroimage.2016.04.016, 2016.
- 9 Adany, P., Choi, I. Y., and Lee, P.: Method for fast lipid reconstruction and removal processing in  $^1\text{H}$  MRSI of the brain,  
10 *Magn Reson Med*, 86, 2930-2944, 10.1002/mrm.28949, 2021.
- 11 An, L., Warach, S., and Shen, J.: Spectral localization by imaging using multielement receiver coils, *Magn Reson Med*, 66,  
12 1-10, 10.1002/mrm.22783, 2011.
- 13 Bashir, A. and Yablonskiy, D. A.: Natural linewidth chemical shift imaging (NL-CSI), *Magn Reson Med*, 56, 7-18,  
14 10.1002/mrm.20917, 2006.
- 15 Bilgic, B., Chatnuntawech, I., Fan, A. P., Setsompop, K., Cauley, S. F., Wald, L. L., and Adalsteinsson, E.: Fast image  
16 reconstruction with L2-regularization, *J Magn Reson Imaging*, 40, 181-191, 10.1002/jmri.24365, 2014.
- 17 De Feyter, H. M., Behar, K. L., Corbin, Z. A., Fulbright, R. K., Brown, P. B., McIntyre, S., Nixon, T. W., Rothman, D. L., and  
18 de Graaf, R. A.: Deuterium metabolic imaging (DMI) for MRI-based 3D mapping of metabolism *in vivo*, *Sci Adv*, 4,  
19 eaat7314, 10.1126/sciadv.aat7314, 2018.
- 20 Dong, Z. and Hwang, J. H.: Lipid signal extraction by SLIM: application to  $^1\text{H}$  MR spectroscopic imaging of human calf  
21 muscles, *Magn Reson Med*, 55, 1447-1453, 2006.
- 22 Haupt, C. I., Schuff, N., Weiner, M. W., and Maudsley, A. A.: Removal of lipid artifacts in  $^1\text{H}$  spectroscopic imaging by data  
23 extrapolation, *Magn Reson Med*, 35, 678-687, 1996.
- 24 Hesse, F., Somai, V., Kreis, F., Bulat, F., Wright, A. J., and Brindle, K. M.: Monitoring tumor cell death in murine tumor  
25 models using deuterium magnetic resonance spectroscopy and spectroscopic imaging, *Proc Natl Acad Sci U S A*, 118,  
26 e2014631118, 10.1073/pnas.2014631118, 2021.
- 27 Hu, X., Patel, M., and Ugurbil, K.: A new strategy for spectroscopic imaging, *J Magn Reson B*, 103, 30-38, 1994.
- 28 Hu, X., Levin, D. N., Lauterbur, P. C., and Spraggins, T.: SLIM: spectral localization by imaging, *Magn Reson Med*, 8, 314-  
29 322, 1988.
- 30 Ip, K. L., Thomas, M. A., Behar, K. L., de Graaf, R. A., and De Feyter, H. M.: Mapping of exogenous choline uptake and  
31 metabolism in rat glioblastoma using deuterium metabolic imaging (DMI), *Front Cell Neurosci*, 17, 1130816,  
32 10.3389/fncel.2023.1130816, 2023.
- 33 Kaggie, J. D., Khan, A. S., Matys, T., Schulte, R. F., Locke, M. J., Grimmer, A., Frary, A., Menih, I. H., Latimer, E., Graves, M.  
34 J., McLean, M. A., and Gallagher, F. A.: Deuterium metabolic imaging and hyperpolarized  $^{13}\text{C}$ -MRI of the normal human  
35 brain at clinical field strength reveals differential cerebral metabolism, *Neuroimage*, 257, 119284,  
36 10.1016/j.neuroimage.2022.119284, 2022.
- 37 Khalidov, I., Van De Ville, D., Jacob, M., Lazeyras, F., and Unser, M.: BSLIM: spectral localization by imaging with explicit  
38  $B_0$  field inhomogeneity compensation, *IEEE transactions on medical imaging*, 26, 990-1000, 10.1109/tmi.2007.897385,  
39 2007.
- 40 Kreis, F., Wright, A. J., Hesse, F., Fala, M., Hu, D. E., and Brindle, K. M.: Measuring tumor glycolytic flux *in vivo* by using  
41 fast deuterium MRI, *Radiology*, 294, 289-296, 10.1148/radiol.2019191242, 2020.
- 42 Liang, Z. and Lauterbur, P. C.: A theoretical analysis of the SLIM technique, *J Magn Reson B*, 102, 54-60, 1993.
- 43 Liang, Z. P. and Lauterbur, P. C.: A generalized series approach to MR spectroscopic imaging, *IEEE transactions on*  
44 *medical imaging*, 10, 132-137, 10.1109/42.79470, 1991.
- 45 Liu, Y., De Feyter, H. M., Fulbright, R. K., McIntyre, S., Nixon, T. W., and de Graaf, R. A.: Interleaved Fluid-attenuated  
46 Inversion Recovery (FLAIR) MRI and Deuterium Metabolic Imaging (DMI) on human brain *in vivo* *Magn Reson Med*, 88,  
47 28-37, 2022.

1 Markovic, S., Roussel, T., Neeman, M., and Frydman, L.: Deuterium Magnetic Resonance Imaging and the Discrimination  
2 of Fetoplacental Metabolism in Normal and L-NAME-Induced Preeclamptic Mice, *Metabolites*, 11, 376,  
3 10.3390/metabo11060376, 2021.

4 Metzger, G., Sarkar, S., Zhang, X., Heberlein, K., Patel, M., and Hu, X.: A hybrid technique for spectroscopic imaging with  
5 reduced truncation artifact, *Magn Reson Imaging*, 17, 435-443, 10.1016/s0730-725x(98)00187-8, 1999.

6 Passeri, A., Mazzuca, S., and Bene, V. D.: Radiofrequency field inhomogeneity compensation in high spatial resolution  
7 magnetic resonance spectroscopic imaging, *Phys Med Biol*, 59, 2913-2934, 10.1088/0031-9155/59/12/2913, 2014.

8 Riis-Vestergaard, M. J., Laustsen, C., Mariager, C., Schulte, R. F., Pedersen, S. B., and Richelsen, B.: Glucose metabolism in  
9 brown adipose tissue determined by deuterium metabolic imaging in rats, *Int J Obes (Lond)*, 44, 1417-1427,  
10 10.1038/s41366-020-0533-7, 2020.

11 Ruhm, L., Avdievich, N., Ziegs, T., Nagel, A. M., De Feyter, H. M., de Graaf, R. A., and Henning, A.: Deuterium metabolic  
12 imaging in the human brain at 9.4 Tesla with high spatial and temporal resolution, *Neuroimage*, 244, 118639,  
13 10.1016/j.neuroimage.2021.118639, 2021.

14 Seres Roig, E., De Feyter, H. M., Nixon, T. W., Ruhm, L., Nikulin, A. V., Scheffler, K., Avdievich, N. I., Henning, A., and de  
15 Graaf, R. A.: Deuterium metabolic imaging of the human brain in vivo at 7 T, *Magn Reson Med*, 89, 29-39,  
16 10.1002/mrm.29439, 2022.

17 Straathof, M., Meerwaldt, A. E., De Feyter, H. M., de Graaf, R. A., and Dijkhuizen, R. M.: Deuterium metabolic imaging of  
18 the healthy and diseased brain, *Neuroscience*, 474, 94-99, 10.1016/j.neuroscience.2021.01.023, 2021.

19 Tkáč, I., Deelchand, D., Dreher, W., Hetherington, H., Kreis, R., Kumaragamage, C., Považan, M., Spielman, D. M.,  
20 Strasser, B., and de Graaf, R. A.: Water and lipid suppression techniques for advanced  $^1\text{H}$  MRS and MRSI of the human  
21 brain: Experts' consensus recommendations, *NMR Biomed*, 34, e4459, 10.1002/nbm.4459, 2021.

22 Veltien, A., van Asten, J., Ravichandran, N., de Graaf, R. A., De Feyter, H. M., Oosterwijk, E., and Heerschap, A.:  
23 Simultaneous Recording of the Uptake and Conversion of Glucose and Choline in Tumors by Deuterium Metabolic  
24 Imaging, *Cancers (Basel)*, 13, 10.3390/cancers13164034, 2021.

25 von Kienlin, M. and Mejia, R.: Spectral localization with optimal pointspread function, *J Magn Reson*, 94, 268-287, 1991.

26 Wang, T., Zhu, X. H., Li, H., Zhang, Y., Zhu, W., Wiesner, H. M., and Chen, W.: Noninvasive assessment of myocardial  
27 energy metabolism and dynamics using in vivo deuterium MRS imaging, *Magn Reson Med*, 86, 2899-2909,  
28 10.1002/mrm.28914, 2021.

29 Zhang, Y., Gabr, R. E., Schar, M., Weiss, R. G., and Bottomley, P. A.: Magnetic resonance Spectroscopy with Linear  
30 Algebraic Modeling (SLAM) for higher speed and sensitivity, *J Magn Reson*, 218, 66-76, 10.1016/j.jmr.2012.03.008, 2012.

## 32 FIGURE LEGENDS

33 **Figure 1 – Lipid removal workflow.** (A) Anatomical MRI and (B) brain and non-brain/skull ROIs. (C) To  
34 accommodate heterogeneity in the skull ROI, it is sub-divided into 125-175 ROIs of 3.5 – 8.5 mL. (D)  $^2\text{H}$ -water  
35 map obtained through numerical integration of each pixel in a  $9 \times 13 \times 11$  DMI dataset. (E)  $^2\text{H}$ -water line shift  
36 map and (F) relative  $^2\text{H}$ -water intensity map. (G) Compartment-specific signals from one brain ROI and 143 skull  
37 ROIs are the primary output of the SLIM algorithm. (H) DMI reconstructed from the 143 skull ROI signals, which  
38 can be subtracted from (D) the original DMI to yield (I) a skull (and lipid) free DMI dataset.

39

40 **Figure 2 – Performance of SLIM-based lipid signal removal on phantoms *in silico*.** (A) Brain and skull tissue  
41 constellation for one out of 50 permutations. (B) Water shift and (D) intensity maps as extracted from DMI data,  
42 providing (C) surrogate  $B_0$  and (E)  $B_1$  maps following low-order polynomial fitting. (F) ROI map with the skull ROI  
43 subdivided into 23 smaller compartments (i.e.,  $N_{\text{skull}} = 23$ ) to accommodate signal heterogeneity. (G) Lipid

1 suppression and (H) metabolite retention under different scenarios including  $N_{\text{skull}} = 1$  without (column I) and with  
2 (column II)  $B_0$  and  $B_1$  compensation,  $N_{\text{skull}} = 20-28$  (column III), 29-36 (column IV), 37-48 (columns V and VI)  
3 without (columns III-V) and with  $B_0$  and  $B_1$  compensation (column VI). Results for the separate brain ROIs (GM,  
4 WM, CSF and pathology) are summarized in Supplemental Figure S2 and essentially mirror the whole brain  
5 results shown in (H). (I, L) Head constellations with the skull ROI subdivided into (I) 23 and (L) 44 smaller  
6 compartments. (J, K, M, N) Spatial response function (SRF) of (J, M) the summed skull ROIs and (K, N) a single  
7 skull ROI for  $N_{\text{skull}} = 23$  (J, K) and  $N_{\text{skull}} = 44$  (M, N). All SRFs have the same vertical scale, spanning -1 to +1.  
8 While the integrated, *phase-sensitive* SRF intensity of a single skull ROI is zero across the brain ROI for any  
9  $N_{\text{skull}}$ , the integrated, *absolute-valued* SRF intensity will be much larger for  $N_{\text{skull}} = 44$  (N) compared to  $N_{\text{skull}} = 23$   
10 (K). Supplemental Figure S3 gives a summary of the absolute-valued SRF across the brain ROI for  $N_{\text{skull}}$  ranging  
11 from 1 to 50.

### 13 **Figure 3 – Performance of SLIM-based lipid signal removal in the presence of partial volume averaging.**

14 (A, B) Brain, tumor and skull ROI constellation used for all simulations. The amount of partial volume averaging  
15 is varied by shifting the ROIs relative to the MRSI grid, giving partial volumes of (A) ~20% and (B) ~55%, whereby  
16 partial volume is defined as  $100 \cdot S_{\text{skull}} / (S_{\text{skull}} + S_{\text{tumor}})$  with  $S_{\text{skull}}$  (red in (A, B)) and  $S_{\text{tumor}}$  (green in (A, B))  
17 representing the amount of signal from a reference signal specific for the skull and tumor ROIs, respectively. (C,  
18 D) MR spectra obtained from the MRSI voxel indicated in (A, B) with standard FFT processing (red) and SLIM-  
19 based processing without (blue) and with (green) skull ROI subdivision. For enhanced visualization, MR spectra  
20 are shifted by 50 Hz each. (E, F) Lactate-to-reference ratio as a function of partial volume for standard FFT  
21 processing (red) or SLIM processing without (blue) or with (green) skull subdivision. As the lipid removal without  
22 skull subdivision (blue) is compromised by  $B_0$  and  $B_1$  magnetic field homogeneity, the lactate-to-reference  
23 changes with increasing partial volume. With skull subdivision (green) the lipid removal becomes less dependent  
24 on spatial homogeneity, leading to a stable lactate-to-reference ratio, independent of the partial volume. Error  
25 bars represent standard deviations calculated over 20  $B_0$  and  $B_1$  magnetic field distributions. Note that the error  
26 bars for the red and green curves fall within the marker.

### 28 **Figure 4 – Performance of SLIM-based lipid signal removal on human brain *in vivo*.**

29 (A)  $T_2$ -weighted spin-echo MRI of a patient harboring a glioblastoma brain tumor with the approximate location outlined in red. The  
30 white dotted lines indicate the in-plane DMI grid (8 mL nominal resolution, circa 30 min total scan time). (B)  $2 \times 2 \times 1$   
31 sub-grid extracted from the  $9 \times 13 \times 11$  DMI dataset showing the original (black, top) and SLIM-processed  
32 (gray, bottom)  $^2\text{H}$  MR spectra. In three spectra a clear natural abundance lipid signal (red arrow) is present in  
33 the original data which is completely removed following SLIM processing. (C)  $4 \times 1 \times 1$  sub-grid covering tumor  
34 and adjacent skull and brain tissue extracted from the  $9 \times 13 \times 11$  DMI dataset. Only the skull voxel shows a  
35 pronounced lipid signal that is removed with SLIM processing. While the remaining three spectra look visually

- 1 identical with or without SLIM processing, the removal of the extracranial lipid signal has a significant effect on
- 2 the (D, E) resulting Lac/(Lac + Glx) metabolic maps.