Determining large hyperfine interactions of a model flavoprotein in the semiquinone state by pulse-EPR techniques

Jesús I. Martínez[1,3], Susana Frago[2], Milagros Medina[2], Inés García-Rubio[3,4]

[1]Departmento de Física de la Materia Condensada, Universidad de Zaragoza, Zaragoza, 50009, Spain
[2]Departmento de Bioquímica y Biología Molecular y Celular and Instituto de Biocomputación y Física de Sistemas Complejos (BIFI), Universidad de Zaragoza, Zaragoza, 50009, Spain
[3]Instituto de Ciencia de Materiales de Aragón, CSIC-Universidad de Zaragoza, 50009, Spain
[4]Institute for Molecular Physical Science, ETH Zurich, 8093 Zürich, Switzerland

Correspondence to: Inés García-Rubio (inesgr@unizar.es)

Abstract.

Flavoproteins are a versatile class of proteins involved in numerous biological processes, including redox reactions, electron transfer, and signal transduction, often relying on their ability to stabilize different oxidation states of their flavin cofactor. A critical feature of flavin cofactors is their capacity to achieve, within particular protein environments, a semiquinone state that plays a pivotal role in mediating single-electron transfer events and is key to understanding flavoprotein reactivity. Hyperfine interactions between the unpaired electron in the semiquinone state and magnetic nuclei in the isoalloxazine ring provide valuable insights into the electronic structure of this intermediate and its mechanistic roles. This study investigates the hyperfine interactions of isotopically labeled flavodoxin (Fld) with ^{13}C and ^{15}N at specific positions of the flavin mononucleotide (FMN) ring using advanced electron paramagnetic resonance (EPR) techniques. The combination of Continuous wave (CW) EPR at X-band and ELDOR-detected NMR and HYSCORE at Q-band revealed a strong and anisotropic hyperfine interaction with the nucleus ^{13}C at 4a and yielded principal tensor values of 40, -13.5 and -9 MHz, the first of which is associated to the axis perpendicular to the flavin plane. On the other hand, as predicted, the hyperfine interaction with the ^{13}C nucleus at position 2 was minimal. Additionally, HYSCORE experiments on ^{15}N-FMN labeled Fld provided precise~~ly~~ axial hyperfine parameters, (74, 5.6, 5.6) MHz for $^{15}N(5)$ and (38, 3.2, 3.2) MHz for $^{15}N(10)$. These were used to refine quadrupole tensor values for ^{14}N nuclei via isotope-dependent scaling. These results showcase the potential of combining CW-EPR, ELDOR-detected NMR, and HYSCORE with isotopic labelling to probe electronic and nuclear interactions in flavoproteins. The new data complete and refine the existing experimental map for the electronic structure of the flavin cofactor and expose systematic divergences between the calculated and experimental values of hyperfine couplings of the atoms most contributing to the SOMO. This could indicate a slight but significant shift of the unpaired electron density from position 4a towards the central nitrogens of the pyrazine ring as compared to the calculations. These results highlight the importance of integrating computational and experimental approaches to refine our understanding of flavin cofactor reactivity.

1 Introduction

Flavoproteins constitute an extensive and versatile family of proteins which are involved in a wide range of biochemical and physiological processes. ~~These include, among others, the catalysis of different redox reactions, the support of electron transfer chains, the ability to act as sensors of the redox state of the cell, oxygen or voltage, and the specific capability to activate catalytic processes upon their activation by blue light or magnetoreception (Walsh &and Wencewicz, 2013;, Minjáarez-Sáaenz et al., 2022;, Calloni &and Vabulas, 2023;, Moreno, A. et al., 2024;, Kaya et al., 20215;, Matysik et al., 2023;, Fraaije &and Mattevi, 2000)~~ These include mainly oxido-reduction processes for the catalysis of different redox reactions (Minjarez-Saez et al., 2022, Walsh and Wencewicz, 2013), as well as the support of electron transfer chains and biosynthetic processes (Curtabbi et al., 2024, Minjarez-Saenz et al., 2025, Buey et al., 2021). Moreover, some flavoproteins have also the specific capability to act as sensors of the redox state of the cell, oxygen or voltage (Fraaije and Mattevi, 2000, Villanueva et al., 2019, Tsukuno et al, 2019), and to activate catalytic processes upon their activation by blue light or magnetoreception (Calloni and Vabulas, 2023, Kaya et al., 2015)~~.~~

Flavoproteins have for~~as~~ cofactor a group that includes the 7,8-dimethyl-isoalloxazine ring, usually in the form of Flavin mononucleotide (FMN) or Flavin adenine dinucleotide (FAD) (see Fig. 1). This ring is able to reach three different oxidation states: fully oxidized, fully reduced by two electrons, and an intermediate semiquinone state holding an additional electron over the oxidized state and, therefore, having an unpaired electron and being paramagnetic. The semiquinone state is hardly~~usually not~~ detectable in redox processes involving free flavins (see Rostas et al., 2018, and references therein), but in some flavoproteins it is highly stabilized by the protein environment (hydrogen bonding and electrostatic interactions). This is particularly the case of electron transferring flavoproteins like flavodoxins (Fld), flavoproteins containing Fld-like domains (as cytochrome P450 reductases and nitric oxide synthase) or bifurcating electron transferring flavoproteins, wich pla~~l~~y crucial roles in anaerobic metabolism, energy conservation, and maintaining redox balance by efficiently distributing electrons (Mayhew & Ludwig, 1975; Iyanagi, 2005; Bestsova et al., 2019; Mohamed-Raseek & Miller 2022; González-Viegas, 2023)~~.~~ This is an indication that the ~~combination of the~~ unique properties of is~~x~~oalloxazine ring are modulated by the protein environment ~~and of the modulation of~~in order to select its redox and electronic properties ~~by the protein environment configures its ability~~ to develop different functions in each specific flavoprotein. In particular, the ability of some flavoproteins to stabilize the semiquinone state during their redox cycle makes them unique molecules to mediate processes involving donor/acceptors with the ability to~~of~~ exchange only one electron at a time (in general, metal centers), with those that necessa~~obligato~~rily exchange two electrons (as for example pyridine nucleotides) (Lans et al., 2012;~~,~~ Young et al, 2020; Moreno et al., 2024).

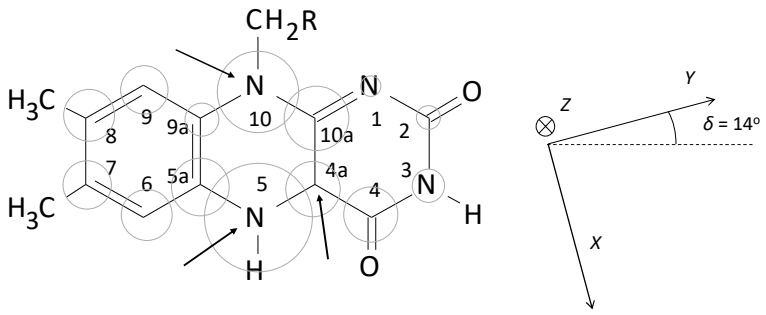


Figure 1: Sketch of the molecular structure and IUPAC numbering of a (7,8-dimethyl) isoalloxazine ~~neutral semiquinone radical~~ring. Grey circles are proportional to the isotropic hyperfine interaction of ^{14}N and ^{13}C nuclei in the neutral semiquinone state. Arrows in the sketch indicate positions where experimental hyperfine interactions are determined in this work. The p~~P~~rincipal axes of the g tensor used in the simulations (from Kay et al. 2005) are displayed on the~~at~~ right.~~.~~

Con formato: Izquierda

Con formato: Fuente: Sin Negrita

Con formato: Inglés (Reino Unido)

The spin density distribution in the semiquinone state of flavin cofactors has a clear relation with protein reactivity and its characterization can provide insights about electron transfer pathways. A better knowledge of the orbital composition of the SOMO of the semiquinone state is of great mechanistic relevance, because the reactivity of the fully reduced state to generate it and the topology of the electron transfer process depend entirely on it. Also the life time of this semiquinone intermediate and the way it transits to the completely oxidized state for processes occurring in the different flavoproteins are intimately related to such electronic structure. Hyperfine interactions between the unpaired electron and the magnetic nuclei in the cofactor are directly related to the orbital composition of the semioccupied orbital (SOMO) and can be used to probe electron density. Moreover, hyperfine interactions are also relevant for the elucidation of the flavoproteins mechanisms themselves as they influence, for instance, flavin magnetochemistry in magnetorecepteion or the lifetime of semiquinone intermediates. Therefore, numerous reports have aimed at the characterization of hyperfine structure using different techniques of Electron Paramagnetic Resonance on several flavoproteins (Weber et al., 2005; Schleicher et al., 2009; Martínez et al., 2012; Schleicher & Weber~~et al.~~, 2012; Brosi et al., 2014; Nohr et al., 2019; Schleicher~~Weber~~ et al., 2021).

The effect of hyperfine interactions can be observed already in the CW-EPR spectra of the flavin, however, characterizations by hyperfine spectroscopy methods able to provide higher resolution (ESEEM-HYSCORE or ENDOR) have revealed more details about the anisotropy of the interaction. The use of different techniques and microwave frequencies, together with model flavoproteins containing isotopically enriched flavins in selected positions or flavin analogues, has allowed to obtain the complete hyperfine tensors for many nuclei at the isoalloxazine ring. Most reported hyperfine interactions refer to protons ~~within~~ bound to the ring. Although they provide relevant information, these interactions are just indirectly related to spin densities at the sites of the ring to which they are attached and, therefore, are less significant compared to the

interactions with nuclei directly part of the ring. To probe the electronic distribution on the carbon atoms on the isoalloxazine, the use of flavin samples with flavins isotopically labelled is necessary, since the most abundant carbon nucleus ^{12}C has no nuclear spin, $I_{12C} = 0$. For nitrogen positions, over 99% of nuclei are ^{14}N, with a nuclear spin $I_{14N} = 1$ that usually displays experimental evidence that is difficult to interpret because of the presence of quadrupole interaction and many nuclear transitions and correlations (see 2. Materials and Methods, 2.4 Spin Hamiltonian section below). Therefore, labelling with ^{15}N ($I_{15N} = 1/2$) is also helpful. As a result of the combination of high-resolution methods and isotopic labelling, some information on couplings to all the nitrogen nuclei in the ring has been obtained through HYSCORE experiments at X band, CW-EPR at W band and pulse ENDOR at W band (Martínez et al., 1997; Barquera et al., 2003; Weber et al., 2005). Additionally, a detailed study has also been published on a protein with a flavin cofactor selectively enriched with ^{13}C at different carbon sites, using the ENDOR pulse technique at W band (Schleicher et al., 2021). However, the characterization of the hyperfine interaction with some of the nuclei at the sites with the highest spin densities, namely positions C(4a) and N(5) (see Fig. 1 for site numbering), still remains incomplete, preventing the experimental determination of the spin population. This information is also of particular interest from the flavoproteins and flavoenzymes functional point of view, because the N(5)-C(4a) locus of the flavin concentrates most of its chemical prowess (Lans et al., 2012; Beaupre & Moran, 2020). In fact, the N(5) of the isoalloxazine is known to be relevant during redox processes and substrate oxygenation, while C(4a) is a recognized site for one-electron chemistry during flavin reoxidation processes (Beaupre & Moran, 2020; Sucharitakul et al., 2011; Ghisla & Massey, 1989; Visitsatthawong et al., 2015; Saleem-Batcha et al., 2018). Furthermore, the reactivity of N(5) and C(4a) allows formation of covalent intermediates contributing to increase the chemical repertoire of natural flavin derivatives within flavoproteins (Leys &and Scrutton, 2016, Beaupre &and Moran, 2020).

Código de campo cambiado

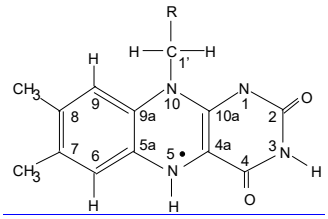
In parallel to these studies, calculations, mainly based on DFT computational methods, have been published in the last twenty-five years to improve the knowledge of the electronic structure of the flavin cofactor in its three oxidation states (HOMO and LUMO of the completely oxidized and reduced states, and SOMO of the semiquinone state) aiming to predict their physicochemical properties (Domratcheva et al., 2014). Although these methods have a great capacity to develop realistic electronic structures, considering in detail the effect of the environment, obviously their validity must be contrasted with experimental results and the discrepancy must be used to improve or refine the results of those calculations. Hyperfine interactions of the unpaired electron in the semiquinone state with nuclei in the vicinity of the isoalloxazine ring are very valuable experimental control parameters to contrast the result of the calculations, as these interactions are directly related to the spin density distribution in the SOMO. They provide an appropriate tool to test the predictions of the calculations for the electronic structure of the semiquinone state, as the magnetic nuclei act as local probes for drawing a map of the electronic spin density within the ring.

Over these years it has been recognized through the calculation of different flavoproteins (such as Fld and DNA photolyase) that the values for the hyperfine parameters obtained through calculations are very sensitive to the level of calculation and

the functional used, as well as to the environment of the flavin that is considered (Weber et al., 20012;, García et al., 2002;, Schleicher et al., 2021). However, experimental evidences indicate that hyperfine interactions are very similar in different flavoproteins (Barquera et al., 2003; Martínez et al., 2012,; Schleicher & Weberet al., 2012; Paulus et al., 2014; Nohr et al., 2019; Pompe et al., 2022), and that the substitution of residues in the flavin environment or in the ring atoms barely affects the hyperfine splittings (Medina et al., 1999;, Martínez et al., 2016). In general, the experimental differences between different flavoproteins are in the same order or smaller than those that appear between calculations depending on their technical details (level, functional used and environment considered). The agreement between the calculations and the known experimental values of a_{iso} is fair for interactions with ^{1}H nuclei (most of the calculated values are within ± 20% of the experimental values) but the hyperfine couplings with ^{14}N(5) and ^{14}N(10) nuclei appear to be systematically underestimated, whereas the relative error remains within ± 20% for the completely characterized ^{13}C nuclei within the flavin ring[1] (Schleicher eat al, 2021).

In the analysis of the hyperfine interactions describing the electronic structure of the flavin semiquinone radical for a better understanding of its role in flavoprotein-catalyzed reactions, it is certainly worth including the hyperfine interactions with nitrogen nuclei. Nitrogen atoms occupyies four positions within the ring, namely N(1), N(3), N(5) and N(10), for which previously reported experimental evidence of the hyperfine couplings is available (Martínez et al., 1997; Barquera et al., 2003; Weber et al., 2005; Martínez et al., 2012). Furthermore, the anisotropic part of the hyperfine also provides relevant information that should not be neglected. In a pure π radical, the hyperfine interaction with nuclei on the ring is axial, being the axis perpendicular to the plane of the ring. The detection of orthorhombic hyperfine matrices or hyperfine principal values that show different proportions between the isotropic and anisotropic part implies the mixture of π and σ orbitals linked to distortion of the molecular and/or electronic structure that can have a relevant effect on the mechanisms where flavin is involved, as it has been shown in Fld variants where the naturally occurring FMN cofactor has been substituted by different analogs (Lans et al., 2012; Martínez et al., 2016). Besides, although this evidence does not directly inform the electronic structure of the fully reduced and oxidized states, the disparity between the calculated and measured hyperfine splitting values offers an indirect indication about differences that may exist between the electronic structures calculated for those states and the real ones.

[1] The relative error for some weakly coupled nuclei is larger than this value. However, since the absolute values of the discrepancies are small, they are considered not very relevant.



~~**Figure 1: Sketch of the molecular structure** and IUPAC numbering of a (7,8-dimethyl) isoalloxazine neutral semiquinone radical.~~

In this work we present an X and Q band study of the neutral semiquinone of *Anabaena* Fld combining CW-EPR, ELDOR detected NMR and HYSCORE experiments with selective ^{13}C and ^{15}N isotope labelling of the flavin. Fld was chosen as a model system due to its feasibility to replacing its FMN cofactor with modified flavins and to its ability to stabilize a large proportion of its neutral semiquinone state (Martínez et al., 2012; Martínez et al., 2014; Martínez et al., 2016, Lans et al., 2012). This particular combination of experiments and microwave frequencies (about 9 and 34 GHz) has turned out to be especially suited for the detection of couplings at the C(4a), N(5) and N(10) positions of the isoalloxazine, allowing the complete characterization of the hyperfine interactions for the nuclei $^{13}C(4a)$, $^{15}N(5)$ and $^{15}N(10)$. These results provide ~~with~~ a suitable protocol to experimentally access these couplings and ~~an~~ thus estimat~~ion of~~ the spin density distribution in the isoalloxazine ring for the Fld model system.

The~~se~~ results are discussed based on the predictions of published calculations and on our knowledge of electron transfer processes involving flavoproteins.

2 Materials and Methods

2.1 Biological Material

Riboflavin (RF) analogues $^{13}C(2)$-RF and $^{13}C(2, 4a)$-RF were converted into the corresponding FMN forms using the mutant H28A of FAD synthase (FADS) from *Corynebacterium ammoniagenes* (Frago et al., 20084~~10;~~ Frago et al., 20104~~08~~), Reaction mixtures containing 50 µM of the RF analogue, 0.5 mM ATP, 1 mM $MgCl_2$ and 1.5–3 µM H28A FADS in 50 mM Tris/HCl, pH 8.0, were incubated in the dark at 37 °C overnight. Full conversion of RF into FMN was checked by thin layer chromatography in silica-gel plates. Once the reaction was completed, FADS was separated from the flavin by ultrafiltration (Amicon Ultra, Millipore, 10000 MW cut-off). ^{15}N-labeled FMN was produced as previously described (Martinez et al., 1997), *Anabaena* Fld was over-expressed in *Escherichia coli* and purified as described in Genzor ~~et~~al., 1996. ApoFld was prepared by treatment with 3% trichloroacetic acid at 4 °C in the presence of dithiothreitol. The precipitated apoprotein was separated from FMN by centrifugation and dissolved in 500 mM MOPS pH 7.0 before dialysis against 50 mM MOPS pH

Con formato: Justificado

Con formato: Justificado, Espacio Después: 0 pto, Interlineado: 1,5 líneas

Con formato: Epígrafe, Izquierda, Interlineado: 1,5 líneas, Punto de tabulación: No en 1,62 cm + 3,23 cm + 4,85 cm + 6,46 cm + 8,08 cm + 9,69 cm + 11,31 cm + 12,92 cm + 14,54 cm + 16,16 cm + 17,77 cm + 19,39 cm + 21 cm + 22,62 cm + 24,23 cm + 25,85 cm

Con formato: Inglés (Estados Unidos)

Con formato: Justificado, Interlineado: 1,5 líneas

Con formato: Inglés (Estados Unidos)

Con formato: Inglés (Estados Unidos)

Con formato: Inglés (Estados Unidos)

Con formato: Inglés (Estados Unidos)

Con formato: Inglés (Estados Unidos)

Con formato: Espacio Después: 0 pto

7.0. Finally, *Anabaena* ApoFld was incubated with a 1.5-fold molar excess of each FMN analogue (namely ^{13}C(2)-FMN, ^{13}C(2, 4a)-FMN or ^{15}N-FMN) in 50 ~~μm~~M MOPS, pH 7.0 for 1 h at 25 °C. Excess flavin was then removed by ultrafiltration and the reconstituted Flds stored at −20 °C. Samples with a protein concentration of 400–800 mM in 50 mM MOPS, pH 7.0, were placed in 3 mm EPR tubes and anaerobically reduced under an argon atmosphere to the semiquinone state at 4 °C by light irradiation with a 150 W Barr & Stroud light source, approximately 7.5 cm from the sample, in the presence of 20 mM EDTA and 2.5 μM 5-deazariboflavin. Once maximal production of the neutral semiquinone was obtained, samples were frozen and stored in liquid nitrogen (at 77 K) until used in EPR measurements.

Con formato: Fuente: Cursiva

2.2 EPR spectroscopy

X-band EPR experiments were performed in a Bruker Elexys E580 spectrometer (microwave (mw) frequency ~ 9.7 GHz) equipped with a cylindrical dielectric cavity and a helium gas-flow cryostat from Oxford Inc. Q-band pulse EPR measurements were carried out on a home-built spectrometer operational in the frequency range of 34.5−35.5 GHz (Gromov et al., 2001) equipped with a custom-made resonator allowing the use of 3 mm sample tubes (Tschaggelar et al., 2009). The spectra were taken at 50 or 90 K. The repetition rate was generally 3 kHz. ~~The~~ HYSCORE and ELDOR-detected NMR experiments were carried out at different observer positions that correspond to different selections of orientations of the molecules with respect to the magnetic field.

2.2.1 CW-EPR and other field-swept spectra~~Electron Spin Echo detected EPR~~

The X-band CW-EPR spectra were acquired at a mw frequency of 9.714 GHz and ~~at~~ a temperature of 50 K, using a modulation amplitude of 2 G and a ~~microwave~~ mw power of 0.32 μW. The Q-band Electron Spin Echo (ESE)-detected EPR spectra were detected using a π/2−τ−π−τ−*echo* sequence with pulse lengths of 16 and 32 ns for the π/2 and π pulses. The Q-band Free Induction Decay (FID)-detected EPR spectra were recorded with the pulse sequence π –*FID,* where the mw pulse was 1μs and the FID was integrated over its whole duration.

2.2.2 HYSCORE

HYSCORE experiments (Höfer, 1994~~;~~, Schweiger &~~et~~ Jeschke, 2001) were performed at Q-band frequency (34.3 GHz) at a temperature of 50 K (^{13}C-labelled samples) or 90 K (^{15}N-labelled samples) using the pulse sequence π/2−τ−π/2−t_1−π−t_2−π/2−τ−*echo*. Different τ values were used and specified in the corresponding figure captions. Unless stated otherwise, pulse lengths of 12 ns for all pulses were~~was~~ used in the experiments performed at the echo maximum to obtain maximum excitation width and 24 and 16 ns pulse lengths were programmed for π/2 and π pulses respectively for the experiments performed at the high-field or low-field flanks of the CW spectrum to obtain better orientation selectivity. The time intervals t_1 and t_2 were varied in steps of 8, 12 or 16 ns starting from 96 ns. A standard phase cycle of eight steps (Gemperle et al., 1990) was used to eliminate unwanted echoes. The experimental time traces were baseline corrected,

apodized with a Hamming or Gaussian window, and zero filled. After a Fourier transformation in the two time dimensions, the absolute-value spectra were calculated and plotted with Matlab.

Con formato: Fuente: Sin Negrita

2.2.3 ELDOR-detected NMR

The Q-band *ELDOR-detected NMR* (Schosseler et al., 1994, Schweiger &~~and~~ Jeschke, 2001) experiments were performed using the pulse sequence $(HTA)_{mw2}$–τ–$(\pi)_{mw1}$–FID. Two rectangular pulses were used. The pulse lengths were 1 µs for the first pulse with variable mw frequency (mw_2), and 1 µs for the second pulse with fixed mw frequency mw_1. Pulse length and power of the detection pulse was set by optimizing FID integrated intensity, whereas ELDOR pulse length and power (which was finally set using an ELDOR channel attenuation of 0 db) were chosen by optimization of the ELDOR detected NMR spectra. The separation between the two pulses was τ = 1.5 µs. The FID generated after the second pulse was integrated over a width of 800 ns. The real and imaginary parts were acquired, baseline shifted, and the absolute value was calculated. The spectra were inverted (multiplied by -1) for display.

2.3 Spectral simulations

~~CW EPR and~~ HYSCORE spectra were simulated using the toolbox for MATLAB EasySpin (Stoll &~~and~~ Schweiger, 2006), version 6.0.6 freely downloadable from www.easyspin.org, using the functions *pepper* and *saffron* respectively and the spin Hamiltonian specified below. For the HYSCORE simulations, in a first step the nuclear frequencies of individual nuclei were computed using the function *endorfreq* and the Hamiltonian given in eq 1, whereby the orientation selection of the experiment was taken into account. From these frequencies, the position and shape of the HYSCORE correlation ridges of the individual nuclei can be deduced, but no information is obtained about the intensity of the cross-peaks. In a second step, ~~when~~ once all the transitions were identified, HYSCORE simulations of either a single nucleus or a set of nuclei ~~the complete nuclear system~~ were done using the function saffron of EasySpin, which provides the intensities of all the ridges, including the~~providing the~~ combination ~~ridges~~ lines that may appear when two or more nuclei are simulated~~and the intensities of all the lines~~. The time-domain simulations were processed and plotted using the same procedure as the experimental data described above.

2.4 Spin Hamiltonian

The Spin Hamiltonian (SH) that was used to analyze the experimental spectra and characterize the hyperfine interactions of the semiquinone radical ($S = 1/2$) with n different nuclear spins (I_i) in the isoalloxazine ring consists of several terms:

$$H = \mu_B \vec{B} \hat{g} \vec{S} + \sum_{j,i} \mu_{N~~n~~} \vec{B} g_{N j~~n~~} \vec{I}_j \vec{I}_i + \sum_j \vec{S} \hat{A}_j \vec{I}_j ~~\vec{S} \hat{A}_i \vec{I}_i~~ + \sum_{I_i > 1/2} \vec{I}_j \vec{I}_i \hat{Q}_j \hat{Q}_i \vec{I}_j \vec{I}_i \quad (1)$$

The first and second terms of the SH represent the electron and nuclear Zeeman interactions, respectively. In the case of semiquinonnme radicals, the electron g-tensor is close to the free electron g-factor, as expected for a radical. However, at higher mw frequencies some anisotropy in the tensor can be resolved (Fuchs et al., 2002; Kay et al., 2005; Okafuji et al., 2008). In this article we assumed (g_{Z} = 2.0022, g_{Y} = 2.0036 and g_{X} = 2.0043), and principal axes as shown in Fig. 1 (Kay et al, 2005****ref***). The third term takes into account the hyperfine interactions with the different magnetic nuclei. If I > 1/2, as is the case for ^{14}N (I = 1), the nuclear-quadrupole interaction has to be included (fourth term). $\hat{A}_i$ and $\hat{Q}_j$ are the hyperfine and nuclear quadrupole tensors, respectively, of nucleus i.

Reflecting the planar symmetry of the molecule, the hyperfine tensors of magnetic nuclei directly in the flavin ring have been reported to be mostly axial, with the separate axis perpendicular to the ring (normally called z). This z axis coincides with Z principal axis of the $\hat{g}$ tensor. Since most of the electron density is located in π orbitals, the hyperfine interaction along this direction uses to be sensiblyis usually noticeably larger than in the other two directions contained in the plane. For moderate mw frequencies (X Band and below) this results in a CW-EPR spectrum with a central intense line corresponding to the perpendicular features and low- and high-field wings that are contributed mainly by the molecules oriented with their axis parallel to the magnetic field. On these wings, some ripples due to large (parallel) hyperfine couplings can be resolved (Martínez et al., 2016). The distance between the two outermost features (highest-field, labelled O2 in Fig. 2, and lowest-field, labelled O1 in Fig. 2) is the sum of the couplings of all magnetic nuclei in the direction perpendicular to the isoalloxazeine plane. For flavins with natural isotopic abundance, the three nuclei with the largest couplings in semiquinone radicals are ^{14}N(5), ^{1}H(5) and ^{14}N(10), therefore, neglecting smaller couplings, the distance between the two outermost shoulders is approximately

$$\Delta B_{\text{out}}^{\text{wt}} = [B(\text{O2})\text{-}B(\text{O1})] \approx C\ \{2\ [A_{iZi}(^{14}\text{N}(5)) + A_{Zi}(^{14}\text{N}(10))] + A_Z(^{1}\text{H}(5))\} \quad (2)$$

where $C = \frac{g_e \mu_B}{h} = 3.57\text{x}10^{-2}$ mT/MHz is a constant for translating the couplings observed in the spectrum (in mT) to MHz consideringtaking into account that the g-factor is close to the one of the free electron, $g \approx g_e$. AsAlso, since hyperfine interactions for N(5) and N(10) are almost axial, we use $A_{||} \equiv A_z$, $A_{\perp} \equiv A_x, A_y$.

Expression (2)This, as it will be shown in the next section, can be used to estimate unknown large hyperfine couplings if the others are known or if a reference spectrum without the particular nucleus of interest is available.

Con formato: Fuente: Cursiva

Con formato: Español (España, internacional)

Con formato: Español (España, internacional)

Con formato: Fuente: Sin Cursiva

3 Experimental results

3.1 Flavodoxin selectively labeled with ^{13}C at positions 2 and 4a of the FMN ring

3.1.1 CW EPR

The X-band CW-EPR spectra for [^{13}C(2)-FMN]-Fld and [^{13}C(2,-4a)-FMN]-Fld samples are shown in Fig. 2. The hyperfine splittingsexperimental trace of non-isotopically labelled Fld (WT Fld), described in a previous work (Martinez et al., 2016),

is also shown here for comparison. According to what was said above, if the hyperfine interaction with the ^{13}C nuclei in the labeled samples were~~as~~ large in the direction perpendicular to the isoalloxazine plane, the distance between the outermost O1 and O2 shoulders is predicted to increase. It can be seen that, for [$^{13}C(2)$-FMN]-Fld, the spectrum does not change with respect to the WT, nor the O1-O2 distance, which indicates that the coupling of ^{13}C nucleus at position 2 is small. On the other hand, an increase in the separation between the two outermost shoulders is clearly seen for [$^{13}C(2, 4a)$-FMN]-Fld. This indicates that the X-band CW-EPR experiments can provide a first estimate of the hyperfine splitting due to the $^{13}C(4a)$ nucleus in the direction perpendicular to the isoalloxazine ring, since the broadening can be directly attributed to the hyperfine coupling of $^{13}C(4a)$:

$$\Delta B_{out}^{13C(4a)FMN} \approx C \{2[A_z(^{14}N(5)) + A_z(^{14}N(10))] + A_z(^{1}H(5)) + A_z(^{13}C(4a)) \} \quad (3)$$

The differences between the B(O2)-B(O1) splitting of the WT Fld (or [$^{13}C(2)$-FMN]-Fld) and [$^{13}C(2,4a)$-FMN]-Fld samples is~~are~~:

$$\Delta B_{out}^{13C(4a)FMN} - \Delta B_{out}^{wt} = (8.4 \pm 0.3 \text{ mT}) - (7.1 \pm 0.3 \text{ mT}) = 1.3 \pm 0.4 \text{ mT} \approx C\, A_z(^{13}C4a) \quad (4),$$

which gives a first estimate of the hyperfine coupling in the direction perpendicular to the isoalloxazine plane ~~of~~ $A_z(^{13}C(4a))$ ≈ 36 MHz.

It should be noted that calculations of the hyperfine coupling of this nucleus in flavoproteins (Weber et al., 2001;~~,~~ García et al., 2002) indicated that it is an anisotropic, nearly axial, interaction, with the largest splitting in the direction perpendicular to the plane. On the other hand, these calculations predicted a value for $A_{z\parallel}(^{13}C(4a))$ around double of that obtained in our experiment ($A_{z\parallel}^{calc}(^{13}C(4a))$ ≈ 70-90 MHz). This discrepancy will be discussed later.

In order to gain accuracy and further information about ~~the~~ $^{13}C(4a)$ hyperfine coupling, advanced EPR techniques were also used.

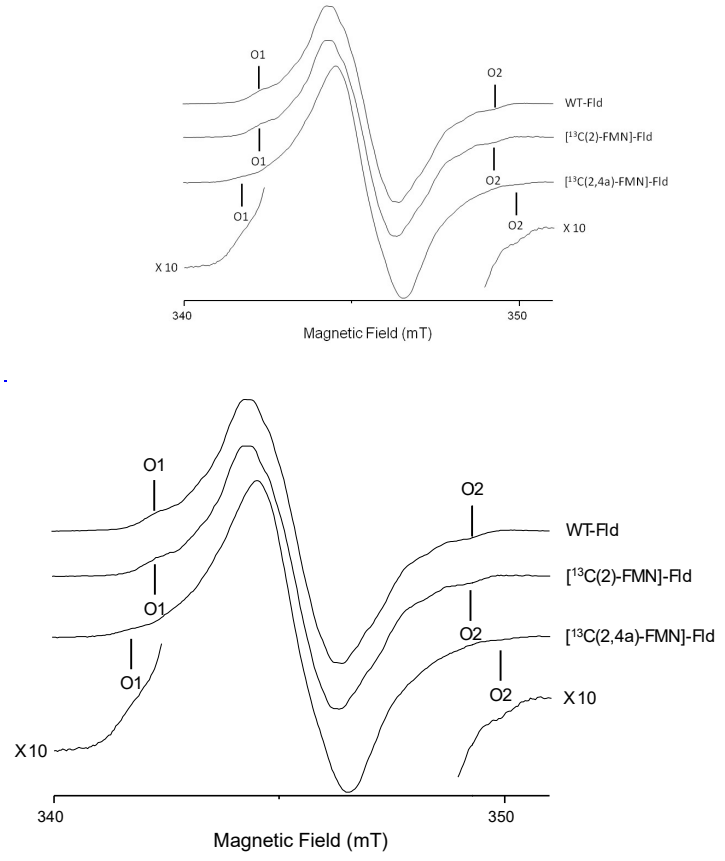


Figure 2: X-band CW-EPR spectra of ^{13}C labelled Fld variants at 50 K.

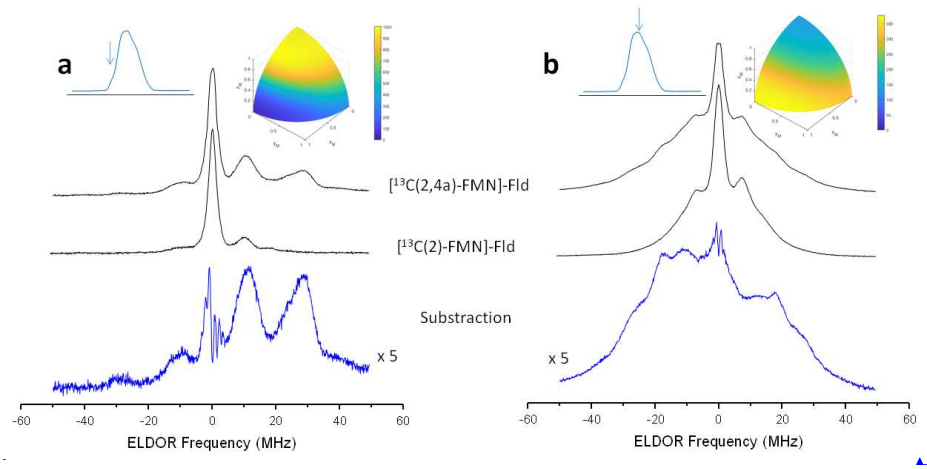
Con formato: Fuente: Sin Negrita

3.1.2 ELDOR-detected NMR

The ELDOR-detected NMR technique can be used to detect nuclear frequencies in systems for which EPR transitions are partially allowed due to, for example, hyperfine anisotropy and/or quadrupole interaction. Pumping an EPR forbidden transition with a variable frequency pulse burns a hole in the polarization that is detected by a decrement ofn the ~~spin echo~~integrated FID intensity generated by ~~the~~ a detection ~~sequence at a variable frequency~~pulse when it hits an allowed EPR transition. Negative peaks associated with nuclear frequencies are obtained when plotting the echo intensity as a function of the ~~detection~~ pump frequency (ELDOR frequency), at symmetric positions with respect to the detection frequency. Since this

experiment is based on driving the polarization, the signal obtained is free of blind spots and other distortions, although it has zero intensity for the principal directions of the hyperfine tensor, ~~(~~since in those directions, EPR transitions are completely allowed~~)~~.

Figure 3 shows the ELDOR detected NMR experiments of [^{13}C(2)-FMN]-Fld and [^{13}C(2,~~-~~4a)-FMN]-Fld. For the ^{13}C(4a) nucleus, the values of the hyperfine coupling and the Larmor frequency at Q-band (13.0 MHz) are comparable. Therefore, at intermediate orientations there will be partially forbidden transitions involving ^{13}C(4a) nuclear levels suitable to be detected with Q-band ELDOR-detected NMR. ~~is especially suitable for detecting nuclear frequencies of ^{13}C(4a).~~ The experiments were performed with the magnetic field set to the Q-band Echo-detected EPR ~~absorption~~ maximum (Fig. 3.b), as well as on the high-field tail~~s~~ of the ~~CW~~ spectrum (Fig. 3.a). As mentioned before, in the second case orientation selection occurs since only molecules oriented with the isoalloxazine plane approximately perpendicular to the direction of the magnetic field will contribute to the spectrum (Martínez et al., 2014). On the other hand, experiments ~~recorded of~~ with the magnetic field at the center of the spectrum are contributed by all possible orientations in the disordered sample with a preference for molecules with the magnetic field oriented close to the isoaloxacine plane (see insets in Fig. 3). The spectra obtained by the subtraction of the two ^{13}C labelled samples measured under identical conditions are~~is~~ also shown for both magnetic field positions and the obtained lines correspond to the nuclear frequencies of the ^{13}C(4a) nucleus (Fig. 3, bottom spectra).



Con formato: Inglés (Estados Unidos)

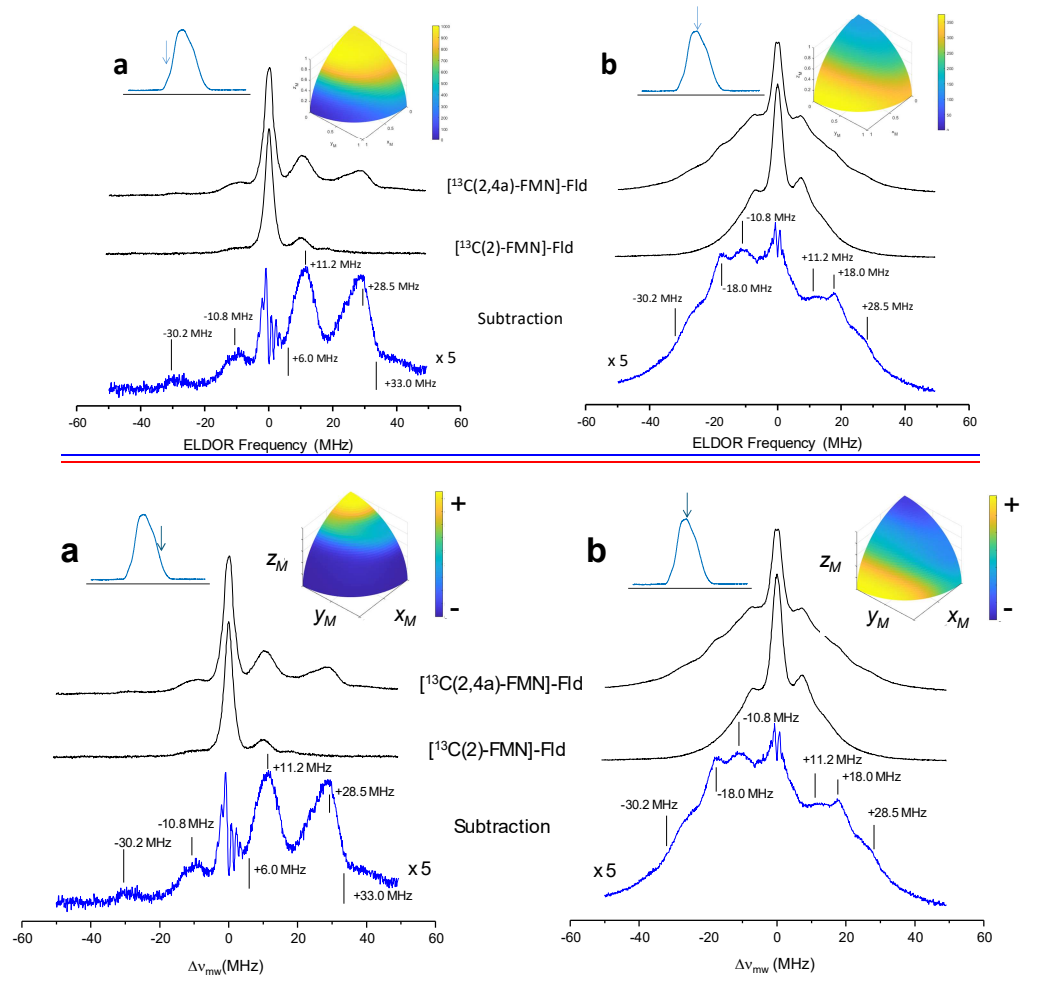


Figure 3: ELDOR-detected NMR spectra of ^{13}C labelled Fld variants. a) Spectra taken at the ~~low~~high-field tail of the Q-band Echo-detected EPR ~~CW~~ spectrum, B = 1225~~1.4~~ mT, ~~for [^{13}C(2,4a)-FMN]-Fld and B = 1222.2 mT for [^{13}C(2)-FMN]-Fld, both~~ corresponding to selective excitation of molecules with the magnetic field oriented perpendicular to the isoalloxazine ring ($A_{\perp}$ since the magnetic field is parallel to the axis). b) Spectra taken at the maximum ~~absorption~~ of the Q-band Echo-detected EPR~~CW~~ spectrum, B = 1221.4 mT~~5~~ for [^{13}C(2,4a)-FMN]-Fld and B = 1222.2 mT for [^{13}C(2)-FMN]-Fld, ~~mT for both samples,~~ which yields ~~no~~ poor orientation selection. For all spectra, T = 50 K. The upper inserts on the left shows the Echo-detected EPR spectrum of the sample with the magnetic field setting of the experiment indicated by an arrow. The inserts on the right show~~s~~ the pattern of excited orientations in a sphere octave according to the colours of the accompanying scale. For the calculation of these patterns a spin Hamiltonian containing the Zeeman anisotropic term and anisotropic hyperfine couplings of $A_{\perp}^{14}$N(5), $A_{\perp}^{14}$N(10) and $A_{\perp}^{1}$H(5) was used. See Table 1 and Fuchs et al. 2002 for hyperfine values. The pulse excitation bandwidth corresponds to 1 MHz.

Con formato: Superíndice

Con formato: Superíndice

Con formato: Superíndice

For the tail spectra, two wide signals are obtained, respectively centered around ± 30 MHz and ± 11 MHz. The value of the largest one is larger than $2\nu_L(^{13}C)$ = 2613.21 MHz, which indicates that the nucleus is in the strong coupling regimen at this particular orientation ($|A_z| > 2\nu_L(^{13}C)$). In this regimen, nuclear frequencies for a nucleus with I = 1/2 are approximately

$$\nu_+ = \left|\frac{A_z}{2} + \nu_L\right| \ , \ \nu_- = \left|\frac{A_z}{2} - \nu_L\right| \qquad (5)$$

and therefore

$$\nu_+ - \nu_- = 2\nu_L \ , \ \nu_+ + \nu_- = |A_z| \qquad (6)$$

The difference between the frequency of the detected peaks is not far fromclose to $2\nu_L(^{13}C)$, which confirms the assignment of the peaks to the nuclear frequencies of ^{13}C(4a). SOn the other hand, since the selection by field is not perfect (see insets in Fig. 3), the peaks in the spectrum are contributed by molecules for which the direction perpendicular to the isoalloxazine plane presents a wide distribution of orientations around that of the magnetic field. It should be also taken into account that the direction perpendicular to the isoalloxazine plane (z) is likely one of the principal axes of the hyperfine tensor of this nucleus (García et al., 2002; Weber et al., 2001). Thus, so molecules perfectly aligned with the field do not contribute to the ELDOR detected NMR spectra, as they display completely allowed hyperfine transitions. Because of this, tThe edge of the detected broad signals (ca. 6 MHz and 33 MHz) cwould then be used in Eqee. 6 to provide a first estimation of the value for the nuclear transitions. The difference between these values (Eqee. 5) is not far from $2\nu_L(^{13}C)$, which confirms the assignment of the peaks to the nuclear frequencies of ^{13}C(4a). Using Eqee. 6, we estimate in this direction $|A_z[^{13}C(4a)]| \approx 40$ 39 MHz, compatible with the estimations from X-band CW-EPR experiments. The signals being wide indicate that the hyperfine tensor of ^{13}C(4a) is quite anisotropic.

In the experiment at the center field very wide signals are also distinguished, as expected for an orientationally disordered sample. The main distinct feature of the spectrum is a peak at ± 18 MHz. It could correspond to the perpendicular feature of the largestr nuclear frequency since many of the orientations are close to the perpendicular plane. In such a case, its absolute value would be approximately $|A_{x,y}[^{13}C(4a)]| \approx 10$ MHz. Assuming that the hyperfine tensor is very anisotropic, it is likely that $A_z[^{13}C(4a)]$ and $A_{x,y}[^{13}C(4a)]$ will exhibit opposite signs. The evidence from the ELDOR detected NMR experiments does not allow resolving A_x and A_y of ^{13}C(4a) but since the hyperfine interactions in the flavin ring tend to be nearly axial, we can assume that $A_x[^{13}C(4a)]$ would be close to $A_y[^{13}C(4a)]$. Then a first estimation from the analysis of these experiments would be:

$A_z[^{13}C(4a)] \approx +40$ 39 MHz $A_\perp[^{13}C(4a)] \approx -10$ MHz

These values can be further refined from evidence obtained from Q-band HYSCORE experiments

Con formato: Fuente: Sin Cursiva

3.1.2 HYSCORE

Q-band HYSCORE experiments were also carried out ~~o~~in [^{13}C(2)-FMN]-Fld and [^{13}C(2,4a)-FMN]-Fld samples, both at the center (Fig. 4 and Fig. S1) and at the tail of the EPR line (Figure S.2 of the Supplementary Material). Again, the larger Larmor frequency of ^{13}C at Q-band allows some weak features from echo modulation with ^{13}C(4a) nuclear frequencies to be seen in the spectra.

The HYSCORE spectra of both samples show intense ridges in the negative quadrant due to ^{14}N that will be discussed later. Since the focus here is on the ^{13}C signals, Fig. 4 only shows the positive quadrant of the 2D measurement, set at the center of the EPR line. The spectrum of [^{13}C(2)-FMN]-Fld, on the left~~right~~, shows a small elongated ridge on the antidiagonal that crosses the diagonal at the ^{13}C Larmor frequency. This line is attributed to the hyperfine coupling of ^{13}C(2), which was estimated smaller than 2 MHz. The low frequency correlations are assigned to ^{14}N. The spectrum of [^{13}C(2,4a)-FMN]-Fld is shown in the center and besides the described lines, a weak and broad bow-shaped~~ribbon-shaped~~ symmetric feature is distinguished on the diagonal, with the knot at about 17 MHz. The feature has been surrounded by a dotted line in Fig. 4.b ~~The long streamer of this ribbon reaches the ^{13}C antidiagonal at about (6, 20) MHz and a shorter inner ridge is distinguished almost perpendicular to the axis.~~ We interpret this feature, undoubtedly associated to ^{13}C(4a), to be the crossing part of two long ridges starting (although not visible) at (40,11) MHz, point corresponding to *z*, the orientation perpendicular to the isoalloxazine plane. The rhombicity of the hyperfine tensor within the plane is manifested by the width of the feature (red area in Fig. 4.c), especially in the two ridges reaching to the antidiagonal indicated by arrows in Fig. 4.c ~~which are the outer borders of the "ribbon"~~. The points where these structures cross the antidiagonal allow estimating the two principal values of the hyperfine tensor in the flavin plane and confirm the hyperfine couplings in the plane and the hyperfine coupling perpendicular to the plane have opposite signs.

Using the value for A_z estimated from the analysis of the ELDOR detected NMR experiments, the following values are obtained from the analysis and simulation of the spectra for the three principal values of the hyperfine tensor:

A_z[^{13}C(4a)] = (+40 ± 2) MHz A_{y2}[^{13}C(4a)] = (-13.5 ± 1)MHz |A_{y2}[^{13}C(4a)]| = (-9 ± 1) MHz

~~The hyperfine couplings within the isoalloxazine plane could not be assigned to particular directions in the plane, therefore, they have been labeled A_2 and A_3 instead of A_y and A_x.~~ The solid red lines superimposed to the spectra on the right spectrum are the HYSCORE patterns calculated with the couplings given above. Proper simulations of the spectrum performed with *saffron* ~~spectral intensities~~ are shown in Figure S.1 of the Supplementary Material.

Con formato: Fuente: Cursiva

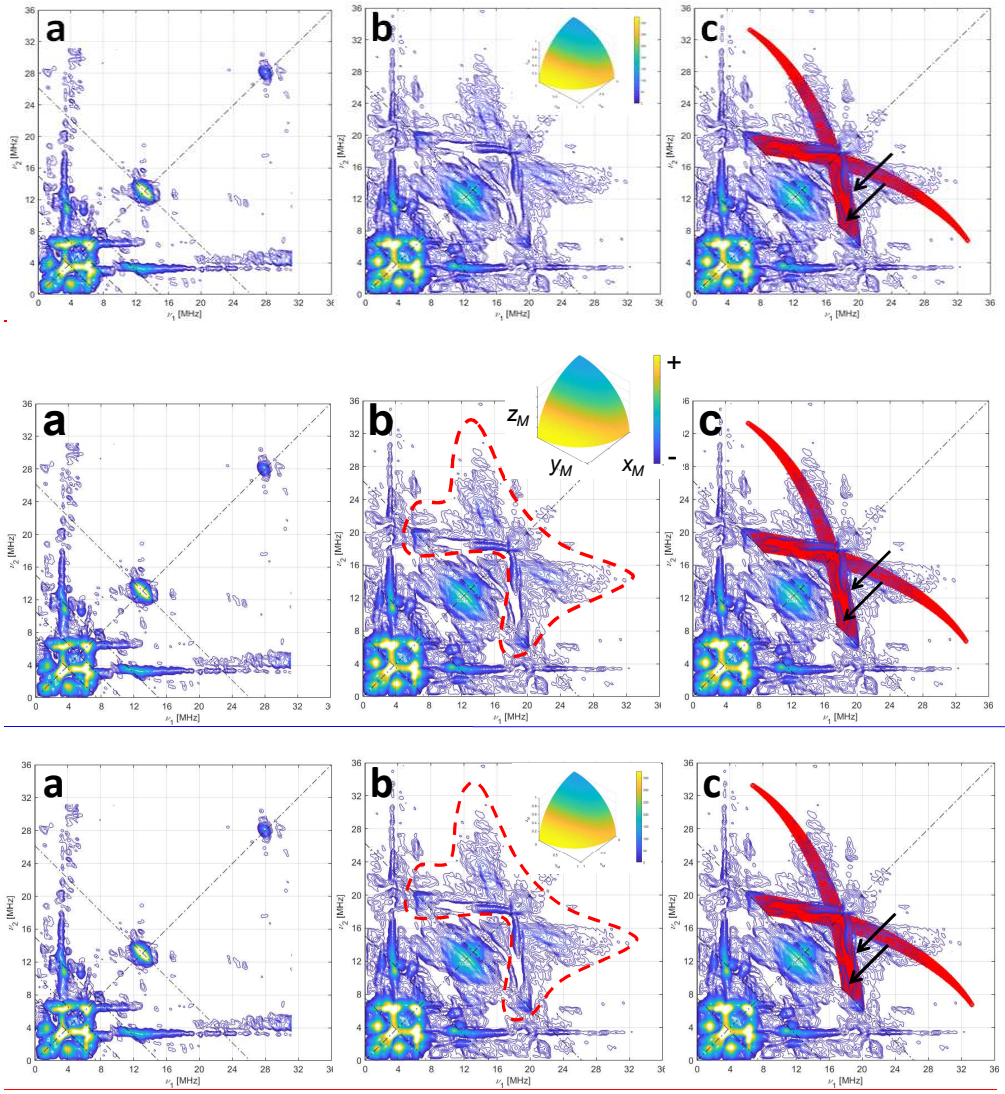


Figure 4: HYSCORE of ^{13}C labelled Fld variants. a) [^{13}C(2)-FMN]-Fld spectrum, b) [^{13}C(2,4a)-FMN]-Fld spectrum, and c) [^{13}C(2,4a)-FMN]-Fld spectrum with the calculated HYSCORE pattern for a ^{13}C nucleus with the parameters specified in the text superimposed in red. Both experimental spectra were taken at the maximum absorption of the ~~CW~~ EPR ~~spectrum~~ absorption~~corresponding to no orientation selection~~, B = 1226.5 mT, τ = 112 ns and T = 50 K. Antidiagonal lines cross the diagonal at the Larmor frequencies ν_{14N}, $2\cdot\nu_{14N}$ and ν_{13C}. Red dotted line in b) shows the feature assigned to ^{13}C(4a) interaction. The two arrows ~~on the right point~~in c) point at the two ridges that

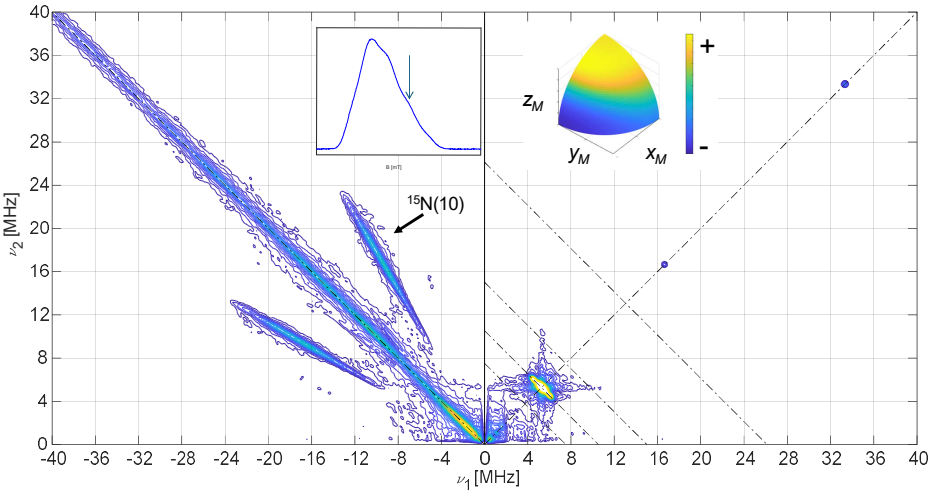
evidence the rhombicity of the ^{13}C(4a) hyperfine tensor. The inset in b) shows the orientation selection of the experimental spectra in a sphere octave according to the colours of the accompanying scale.

3.2 Flavodoxin isotopically labeled with ^{15}N at the FMN ring

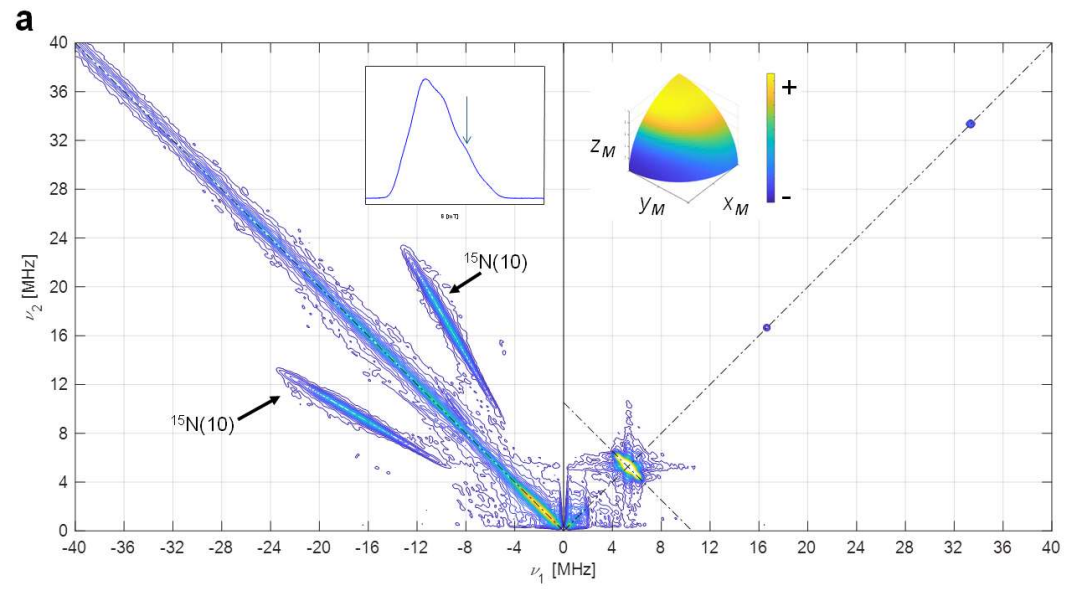
3.2.1 HYSCORE

Although the Q-band HYSCORE spectra of samples with natural abundance of nitrogen nuclei present intense signals due to hyperfine interactions with these nuclei, their interpretation is difficult, because the ^{14}N nucleus has a nuclear spin $I = 1$ and an appreciable quadrupole contribution, which causes the appearance of multiple correlation features (see Fig. 6). The use of samples labelled with ^{15}N-FMN greatly simplifies the analysis, since its I = 1/2 nucleus presents a single nuclear transition per electron spin manifold, and therefore a single pair of correlated features per nucleus. The hyperfine parameters obtained from ^{15}N are directly convertible to those of the ^{14}N nucleus at the same position.

a

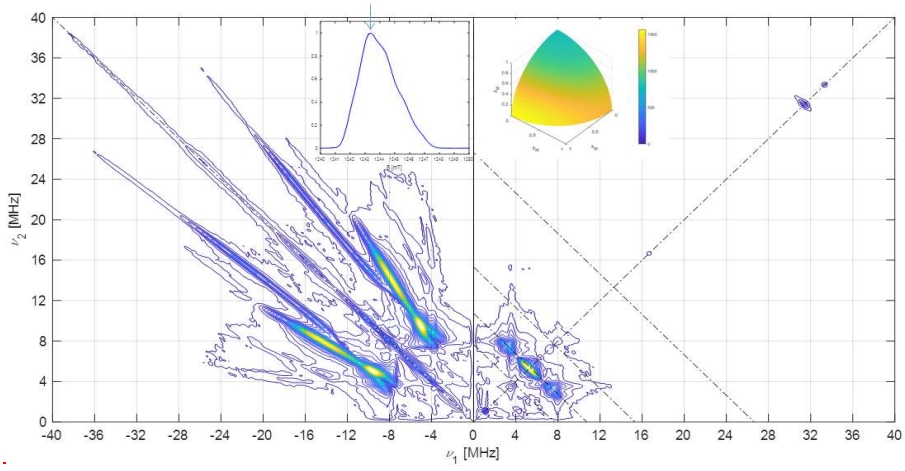

40
36
32
28
24
20
16
12
8
4
0
-40 -36 -32 -28 -24 -20 -16 -12 -8 -4 0 4 8 12 16 20 24 28 32 36 40
ν_2 [MHz]
ν_1 [MHz]
^{15}N(10)
z_M
y_M
x_M
+
-

a



410

b



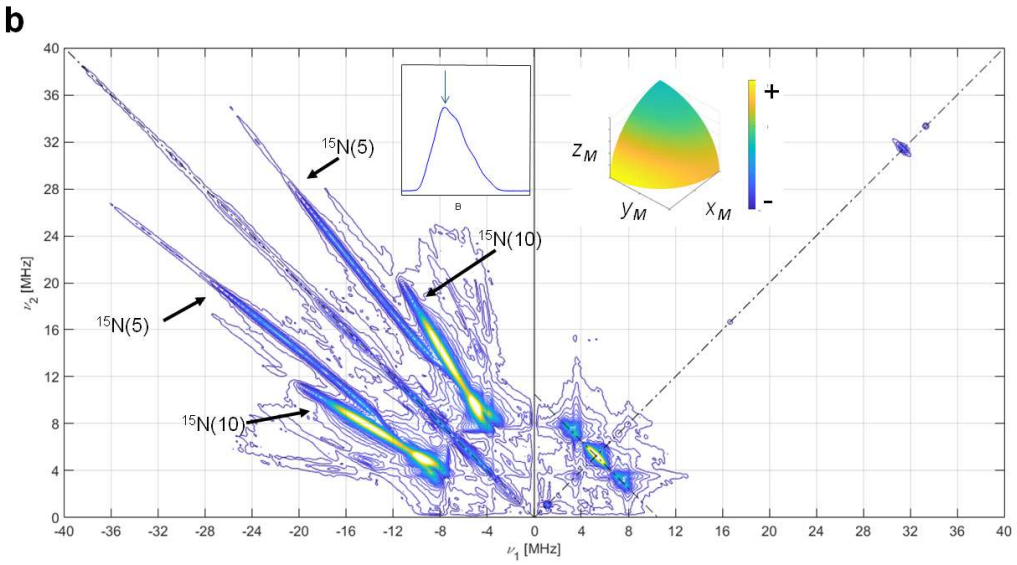


Figure 5: HYSCORE of [^{15}N-FMN]-Fld. a) Spectrum taken at the high-field tail of the CW spectrum, B = 1219.7 mT, the displayed spectrum is the sum of spectra taken at τ values of 96, 124, 144 and 168 ns. b) Spectrum taken at the absorption maximum of the CW spectrum, B = 1217.2 mT, the displayed spectrum is the sum of spectra taken at τ values of 96, 144 and 168 ns. T = 98 K for all spectra. The antidiagonal line crossing the (+,+) diagonal at the Larmor frequency ν_{15N} has been included for reference. The upper insert on the left shows the eEcho-detected EPR spectrum of the sample with the magnetic field setting of the experiment indicated by an arrow. The insert on the right shows the pattern of excited orientations for each spectrum.

The Q-band HYSCORE experiments performed on a [^{15}N-FMN]-Fld sample are displayed in Fig. 5, both, at the upper tail of the EPR spectrum (Fig. 5.a) and at its maximum (Fig. 5.b). To the first spectrum, only molecules with the magnetic field approximately perpendicular to the flavin plane contribute (see insert in Fig. 5.a). The hyperfine coupling of N(5) at this orientation is too large (Martínez et al., 1997; Weber et al., 2005) for the excitation bandwidth of the mw pulses to be enough to excite its nuclear frequencies. The spectrum, therefore, only shows a pair of ridges, symmetrical with respect to the diagonal and approximately parallel to it. This features can be associated with a hyperfine interaction with a nucleus of spin I = ½ and attributed to N(10). From the distance between these ridges, the hyperfine interaction of N(10) close to *z* can be estimated. The short ridge on the ^{15}N antidiagonal in the positive quadrant is assigned to weakly interacting nuclei N(1) and N(3) whose hyperfine couplings have been reported somewhere else (Martínez et al. 2012).

The HYSSYCORE spectrum at the maximum of the EPR line (Fig. 5.b) displays two crossing pairs of correlated ridges, one of them is very long, associated with a highly anisotropic and strong interaction, and assigned to N(5) according to previous results (Martínez et al., 1997; Weber et al., 2005). The other is shorter and overlaps partially with the ridge seen in the

parallel spectrum and is assigned to N(10). Additionally, two pairs of peaks on the ^{15}N antidiagonal come out in the positive quadrant at the low-frequency edge of the two ridges, allowing ~~to identify~~identifying the hyperfine coupling in the plane as isotropic within the plane. Satisfactory simulations (see Fig. S.3 and S.4 in the Supplementary Material) were produced using the following axial hyperfine parameters:

$A_z[^{15}N(5)] = (+74 \pm 3)$ MHz $A_\perp[^{15}N(5)] = (+5.6 \pm 0.3)$ MHz

$A_z[^{15}N(10)] = (+38.0 \pm 1.0)$ MHz $A_\perp[^{15}N(10)] = (+3.2 \pm 0.3)$ MHz

Here, and in the rest of the article, the errors in the hyperfine parameters were estimated with the aid of multiple calculations of the ridge positions. The optimum parameters were varied from their optimum value, one by one and the error was set observing the parameter value that gave a calculated feature whose position was clearly not coincident with the experimental one. The orientation of the tensor cannot be obtained from our results, the one published by Kay and coworkers was used (Fuchs et al., 2002~~;~~, Kay et al., 2005) for the simulations.

3.3 HYSCORE signals of ^{14}N at the FMN ring

Once the hyperfine coupling parameters of ^{15}N are estimated with high precision, they can be directly transformed to the ones of ^{14}N in the same position by just applying a factor $g_N(^{14}N)/ g_N(^{15}N) = - 0.71$. With the hyperfine already established, the experimental data on the ^{14}N-FMN can be used to refine the values of the quadrupole tensor. In Fig. 6, the complete set of spectra for [^{13}C(2)-FMN]-Fld are shown. As mentioned before, they are dominated by ^{14}N ridges. The spectrum at the high-field tail shows two short pairs of ridges assigned to N(10) for the reason mentioned above. It allows obtaining a value of 0.8 MHz for $|Q_z|$, the principal value of the quadrupole tensor in the direction perpendicular to the isoalloxazine ring. The spectrum recorded at the ~~center~~ maximum of the EPR absorption~~line~~ contains ridges due to N(10) and N(5) (Fig. 6.b). The best simulation of the spectra, shown in Fig. S.6 of the Supplementary Material, was produced with the following quadrupole parameters:

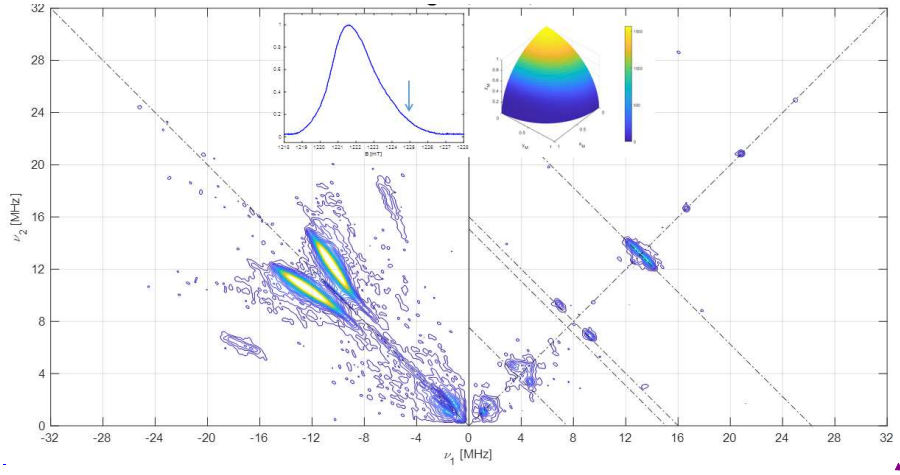
$Q_z[^{14}N(10)] = -0.8 \pm 0.1$ MHz $Q_{\perp,1}[^{14}N(10)] = 2.4 \pm 0.1$ MHz $Q_{\perp,2}[^{14}N(10)] = -1.6 \pm 0.1$ MHz

$Q_z[^{14}N(5)] = 1.8 \pm 0.1$ MHz $Q_{\perp,1}[^{14}N(5)] = -0.8 \pm 0.1$ MHz $Q_{\perp,2}[^{14}N(5)] = -1.0 \pm 0.1$ MHz

Note that the errors in the quadrupole parameters have to be correlated so that the sum of all three principal values sum up to zero. With our data, these principal values cannot be associated with a particular axis in the plane and we have, therefore, labeled the principal values in the flavin plane as $Q_{\perp,1 \text{ or } 2}$.

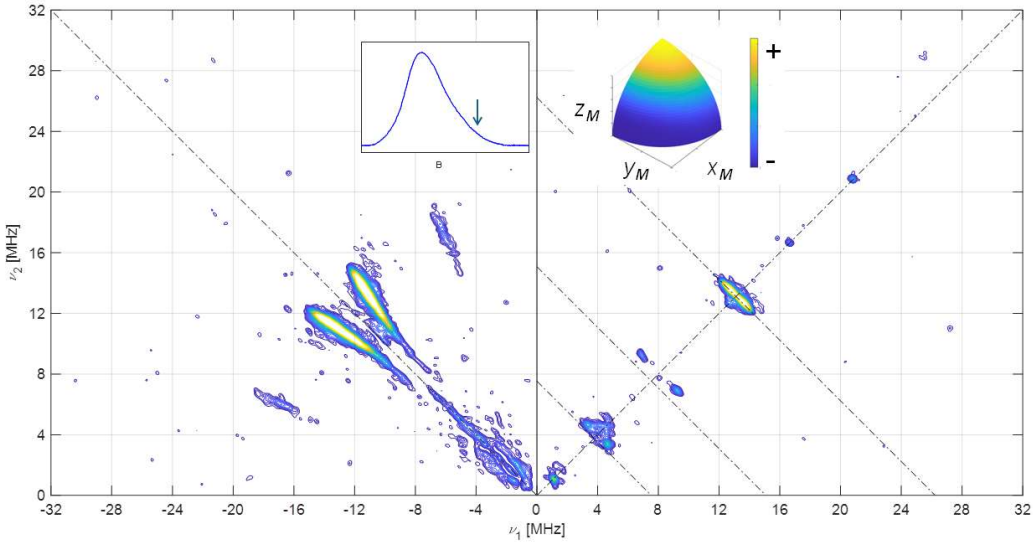
Con formato: Fuente: Cursiva

~~a~~

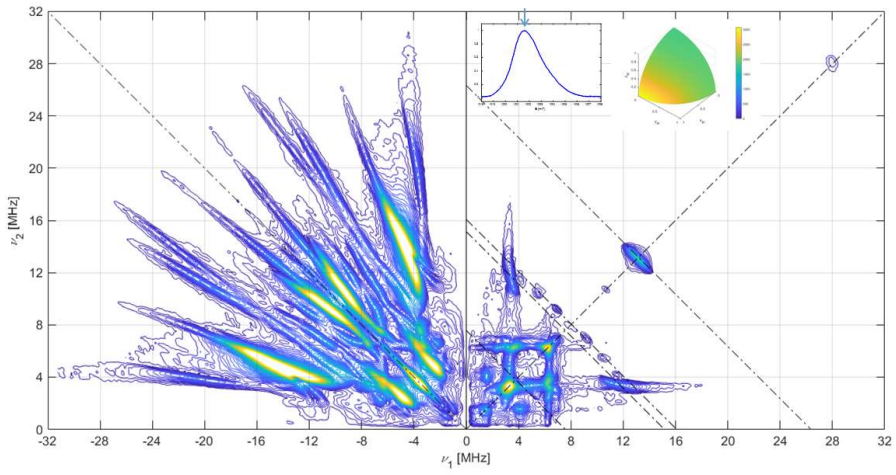


Con formato: Inglés (Estados Unidos)

a



~~b~~



b

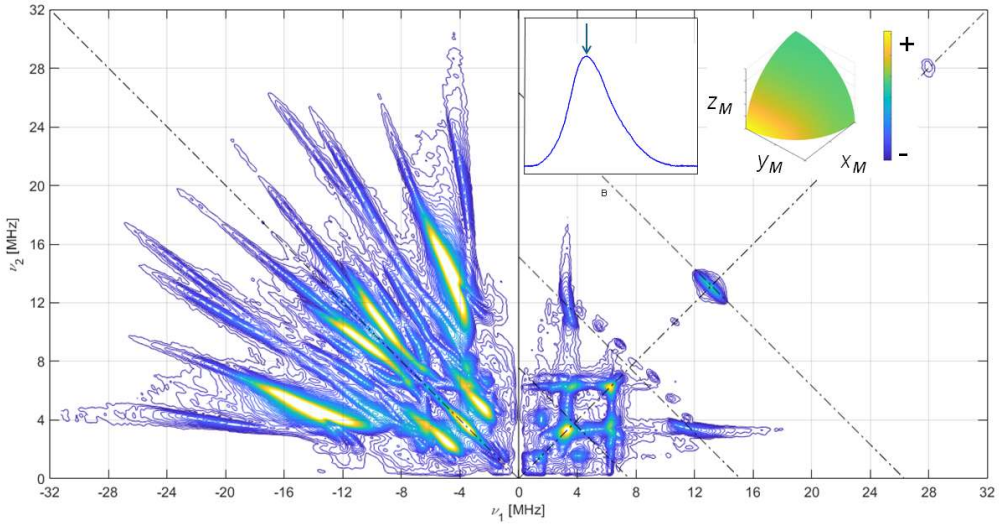


Figure 6: ^{14}N HYSCORE signals Fld samples with naturally abundant nitrogen in the FMN. Spectra recorded using the sample [^{13}C(2)-FMN]-Fld. a) Spectrum taken at the tail of the ~~CW spectrum~~EPR absorption, B = 1225.0 mT~~.~~. T~~t~~he displayed spectrum is the sum of spectra taken at τ values of 96, 112, 128, 144 and 176 ns, b) Spectrum taken at the EPR absorption maximum ~~of the CW spectrum~~, B = 1221.0 mT, T~~t~~he displayed spectrum is the sum of spectra taken at τ values of 96, 128 and 208 ns. T = 50 K for all spectra. The antidiagonal lines crossing the (+,+) diagonal at the Larmor frequencies $\nu_{14N,}$ $2\cdot\nu_{14N}$~~, ν_{2H}~~ and ν_{13C} have been included for reference. The

upper insert on the left shows the Echo-detected EPR spectrum of the sample with the magnetic field setting of the experiment indicated by an arrow. The insert on the right shows the pattern of excited orientations for each spectrum.

3 Discussion

In this work, the use of hyperfine spectroscopy techniques at~~in~~ Q-band, in combination with selective isotopic labeling, has~~ve~~ proven very useful to experimentally determine the hyperfine couplings of the unpaired electron of flavoproteins in the semiquinone state with the nuclei of the isoalloxaz~~e~~ine ring bearing the highest spin densities. The complete set of principal hyperfine values of ^{13}C(4a), ^{15}N(5) and ^{15}N(10) nuclei have been determined. Additionally, the spectroscopic effects of the nuclear quadrupole interaction of ^{14}N(5) and ^{14}N(10) have been analyzed. Despite the relevance of these positions in the ring, until now only partial experimental evidence of hyperfine interactions was available. ~~The interaction with ^{15}N(10) and ^{14}N(10) in this protein had been previously determined by us using X-band HYSCORE experiments (Martínez et al., 1997).~~ The Q-band results presented here show that both~~,~~ hyperfine and quadrupolar interactions are compatible with and allow refinement of the ones previously reported for N(10) determined using X-band HYSCORE experiments (Martínez et al., 1997)~~.~~. Concerning N(5), the hyperfine structure was never obtained for Fld~~this protein~~ but the hyperfine matrix was determined by an analysis of the X-band and W-band CW-EPR spectra assisted by simulation in the flavoprotein Na^+-NQR (Barquera et al., 2003). Here we have shown that direct evidence for the hyperfine interaction with N(5) can also be obtained using Q-band HYSCORE experiments providing very similar results in ~~f~~Fld. Estimations for the quadrupolar interaction for this nucleus were also obtained for the first time due to the high resolution of 2D HYSCORE experiments. In spite of the symmetric positions of N(5) and N(10) in the pyrazine central ring, the nuclear quadrupole values differ considerably, possibly indicating a significant contribution of the unpaired electron density, considerably larger in the former atom.

Regarding the hyperfine interaction with ^{13}C, our X-band CW-EPR, and Q-band ELDOR~~-~~detected NMR and ~~and Q-band~~ HYSCORE experiments allowed us to determine the complete set of the principal values for~~of~~ the hyperfine interaction. Previously, it was only ~~possible to determine~~reported for DNA photolyase the two smallest principal values (Schleicher et al., 2021), which are in the same range of the ones found here. For the hyperfine couplings of ^{13}C(2), we find also compatible values for F~~f~~ld, always smaller than 2 MHz.

Our findings can be used for a wide comparison of measured and calculated hyperfine parameters for the main isoallosazine ring positions in flavoproteins. Table 1 displays~~shows~~ a~~the~~ comparison between experiments and calculations for ~~the ring~~ nuclei directly on the ring, bearing~~with~~ the highest hyperfine couplings (4, 4a, 5, 5a, 10, 10a), where both the isotropic and the anisotropic parts of the hyperfine principal values are considered. As indicated in the Introduction section, measured hyperfine values for different flavoproteins exhibiting neutral semiquinone show just small differences, comparable to the variation between reported calculations (Weber et al., 2001; García et al., 2002; Schleicher et al., 2021).

~~It is worth noting that only the complete characterization of the hyperfine structure allows obtaining the isotropic and anisotropic components of ^{13}C(4a), see Table 1, the former related to the spin density at the nucleus and the second one directly related to the spin population in the π orbitals of the atom. In addition, accurate values of the largest hyperfine splittings in the isoalloxazine ring are critical to characterize the magnetochemistry involved in the magnetoreception of avian cryptochromes (Hore et al., 2016), so the reported experimental values of should be very useful for modelling this mechanism.~~

~~For other positions in the ring, experimental evidence shows only small differences in the hyperfine structure between different flavoproteins exhibiting neutral semiquinone, even when their functions are very different. It remains to be proved whether this holds true for the newly estimated spin densities of C(4a) or they vary among the different flavoproteins.~~

Table 1.- Comparison between measured and calculated hyperfine parameters for the flavin ring sites with the largest hyperfine couplings.

	Measured Isotropic hyperfine parameter a (MHz)[a]	Calculated Isotropic hyperfine parameter a (MHz)[a]	Relative inaccuracy in a (exp-calc, %)	Measured anisotropic hyperfine parameters T_x, T_y, T_z[b] (MHz)	Calculated anisotropic hyperfine parameters T_1, T_2, T_3[b] (MHz)
^{13}C(4)[c]	-9.7	-11.2	-16%	-4.1, -1.5, +5.7	-0.4, -1.3, +1.8
^{13}C(4a)	+5.8[d]	+13.2[c]	126%	-14.8, -19.4, +34.2[d]	-23.3, -22.7, +46.0[c]
^{14}N(5)	+20.2[d,e]	+13.6[f]	-33%	-16.2, -16.2, +32.4[d,e]	-14.6, -14.6, +29.2[f]
^{13}C(5a)[c]	-13.2	-12.3	7%	-1.9, +0.2, +1.6	+1.7, +2.3, -3.9
^{14}N(10)	+10.5[d,e]	+7.6[f]	-28%	-8.2, -8.2, +16.5[e]	-7.6, -7.6, +15.2[f]
^{13}C(10a)[c]	-14.0	-13.6	3%	-1.4, +0.4, +0.9	+1.1, -0.7, -0.5

a. Isotropic hyperfine parameter $a = \frac{(A_x+A_y+A_z)}{3}$

b. $T_i = A_i - a\,, i = x, y, z$

c. Ref. ~~({~~Schleicher et al., 2021)~~}~~

d. This work, in Fld

e. The hyperfine parameters for ^{14}N nucleus have been scaled from ^{15}N ones (see text).

f. Ref. ~~({~~García et al., 2002)~~}~~

Con formato: Español (España, internacional)

Con formato: Español (España, internacional)

~~When a comparison is made with previously calculated values of the hyperfine interaction, our results are quite surprising. Given that all hyperfine interactions had been estimated by calculations, and considering the good general agreement that existed with the known experimental values, until now the values calculated also for the ^{13}C(4a) nucleus were considered plausible. Nevertheless, our results show~~In the table, it can be seen that the hyperfine interaction of ^{13}C(4a) predicted by calculations is significantly overestimated, regardless of whether its isotropic or anisotropic part are compared. This overestimation represents the most significant discrepancy within the flavin isoalloxazine ring in percentage terms. On the other hand, this could be related to the previously mentioned underestimation of the hyperfine splitting in the ^{14}N(5) and ^{14}N(10) nuclei (Weber et al., 2001; García et al., 2002). ~~T~~the isotropic hyperfine constants obtained in the calculations are clearly underestimated for ^{14}N(5) and ^{14}N(10), and severely overestimated for ^{13}C(4a), in which a value more than double the one obtained experimentally is predicted. Furthermore, a similar trend occurs when comparing the calculated and measured data for the anisotropic part of the interactions. The calculations reproduce well the almost axial character of the hyperfine matrices for the three nuclei, but underestimate their magnitude for ^{14}N(5) and ^{14}N(10) (between 10% and 8%), and overestimate it for ^{13}C(4a) (around 35%).

The values of the isotropic hyperfine parameter calculated for all ^{13}C nuclei, except for position 4a, reproduce the real values quite well. The overestimation in the ^{13}C(4a) calculations is quite significant, while in the calculations the interactions with the carbon nuclei in positions 4 and 5a are comparable with those in position 4a (and in previous calculations they were considered much smaller, see Weber et al., 2001), the experimental values reveal that the latter has a value nearly half that of the others. Regarding the anisotropic part, the one of ^{13}C(4a) remains the largest among the carbon nuclei, but its experimental value makes it comparable to that of the ^{14}N(5) and ^{14}N(10) nuclei, despite the fact that the nuclear gyromagnetic factor in these nuclei is quite smaller than that of ^{13}C.

~~The discrepancies between calculations and experimental values could, therefore, indicate that this issue is not specific of type of nucleus for which the interaction is estimated. Instead, it may reflect a broader difficulty in the ability of the of the calculations to predict realistic hyperfine interactions or the potential overlooking of a relevant factor. It is worth noting that reported calculations took both actual flavin structures in the semiquinone state from X-ray diffraction experiments and DFT optimized structures, obtaining very similar results.~~

~~Table 1 shows the comparison between experiments and calculations for the ring nuclei with the highest hyperfine couplings (4, 4a, 5, 5a, 10, 10a), where both the isotropic and the anisotropic parts of the hyperfine principal values are considered. The values of the isotropic hyperfine parameter calculated for all ^{13}C nuclei, except for position 4a, reproduce the real values quite well. Concerning the anisotropic part, it is relatively small in all these nuclei, and has a marked orthorhombic character, which indicates an important mixture of σ orbitals in the SOMO for those positions. Besides, the isotropic hyperfine constants obtained in the calculations are clearly underestimated for ^{14}N(5) and ^{14}N(10), and severely overestimated for ^{13}C(4a), in which a value more than double the one obtained experimentally is predicted. Furthermore, a similar trend occurs when comparing the calculated and measured data for the anisotropic part of the interactions. The calculations reproduce~~

Con formato: Fuente: 10 pto

Con formato: Fuente: 10 pto

Con formato: Inglés (Estados Unidos)

~~well the almost axial character of the hyperfine matrices for the three nuclei, but underestimate their magnitude for ^{14}N(5) and ^{14}N(10) (between 10% and 8%), and overestimate it for ^{13}C(4a) (around 35%). The overestimation in the ^{13}C(4a) calculations is quite significant, while in the calculations the interactions with the carbon nuclei in positions 4 and 5a are comparable with those in position 4a (and in previous calculations they were considered much smaller, see Weber et al., 2001), the experimental values reveal that the latter has a value nearly half that of the others. Regarding the anisotropic part, the one of ^{13}C(4a) remains the largest among the carbon nuclei, but its experimental value makes it comparable to that of the ^{14}N(5) and ^{14}N(10) nuclei, despite the fact that the nuclear gyromagnetic factor in these nuclei is quite smaller than that of ^{13}C.~~

All this shows that the current calculations present an essential difficulty in realistically describing the electronic spin distribution of SOMO, which in turn could indicate the need to also improve other aspects of electronic structure prediction. Our findings suggest that the spin density predicted at position 4a could be actually shifted towards the central positions of pyrazine (5 and 10), which may have important consequences on the understanding of the electron transfer mechanisms that specifically involve these positions (Ghisla & Massey, 1989; Lans et al., 2012; Kaya et al., 2025). The fact that the only significant differences between the hyperfine calculations and the experimental values relate to a shift of the electron density between the two of the most reactive positions of the flavin cofactor is intriguing and it remains to be investigated whether this is a general feature in the semiquinone state of flavoproteins or it can be hypothetically associated to a modulation of the reactivity by the protein environment. For example, more electron density in position C(4a) would favor reoxidation whereas more electron density in N(5) would promote hydride transfer (Schleicher & Weber, 2012; Edwards, 2014; Beaupre & Moran, 2020; Guerriere et al., 2025). Whether this modulation is due to a specific protein-flavin interaction or a structural distortion of the isoalloxacin~~t~~e ring is certainly interesting and remains to be identified in future studies. In addition, accurate values of the largest hyperfine splittings in the isoalloxazine ring are critical to characterize the magnetochemistry involved in the magnetoreception of avian cryptochromes (Hore & Mouritsen~~et al.~~, 2016), so the reported experimental values ~~of~~ should be very useful for modelling this mechanism.

Supplementary Material The supplementary Material is available together with the online version of this article.

Author contributions JIM and MM designed the research. SF prepared the protein samples. IGR carried out the experiments and analyzed the data together with JIM. JIM, IGR and MM wrote the manuscript.

Acknowledgments

Riboflavin analogues were a generous gift from Dr. D. Edmondson. Gunnar Jeschke kindly provided access to the spectrometer park of his laboratory to perform the experiments

Financial support

This work has been funded by the Spanish State Research Agency and by FEDER (MCIN/AEI-FEDER, Grants PID2022-136369NB-I00, PID2022-140923NB-C21 and PID2021-127287NB-I00) and the Regional Government of Aragón-FEDER (grants E35_23R and E09-23R), as well as MCIN with resources from European Union NextGenerationEU (PRTR-C17.I1) promoted by the Government of Aragon

Competing interests. The authors declare no conflict of interest.

Raw data and code availability. Raw data and Matlab/Easyspin simulation codes are available at CSIC Digital/ Zenodo.

References

Barquera, B., Morgan, J. E., Lukoyanov, D., Scholes, C. P., Gennis, R. B., Nilges, M. J.:X- and W-band EPR and Q-band ENDOR studies of the flavin radical in the Na+ -translocating NADH:quinone oxidoreductase from Vibrio cholerae, J. Am. Chem. Soc. 125, 265-275, doi: 10.1021/ja0207201, 2003.

Beaupre, B. A., and Moran, G. R., :N5 Is the New C4a: Biochemical Functionalization of Reduced Flavins at the N5 Position. Front. Mol. Biosci., 7, 598912, doi: 10.3389/fmolb.2020.598912, 2020.

Bestsova, Y. V., Kulik, L. V., Mamedov, M. D., Baykov, A. A:, Bogachev, A. V., Flavodoxin with and air-stable flavin semiquinone in a green sulfur bacterium, Photosynth. Res. 142, 127-136, doi: 10.1007/s11120-019-00658-1, 2019.

Brosi, R., Bittl, R., and Engelhard, C.:, EPR of flavoproteins, Methods in Mol. Biol. 1146,341-360, doi: 10.1007/978-1-4939-0452-5_13, 2014

Buey, R.M., Fernández-Justel, D., González-Holgado, G., Martínez-Júlvez, M., González-López, A., Velázquez-Campoy, A., Medina, M., Buchanan, B.B., Balsera, M., Unexpected diversity of ferredoxin-dependent thioredoxin reductases in cyanobacteria. Plant Physiol. 27;186(1):285-296. doi: 10.1093/plphys/kiab072, 2021.

Calloni, G., and Vabulas, R. M.: The structural and functional roles of the flavin cofactor FAD in mammalian cryptochromes, Front. Mol. Biosci., 9, 2022, doi: 10.3389/fmolb.2022.1081661, 2023.

Curtabbi, A., Guarás, A., Cabrera-Alarcón, J.L., Rivero, M., Calvo, E., Rosa-Moreno, M., Vázquez, J., Medina, M., Enríquez, J.A. Regulation of respiratory complex I assembly by FMN cofactor targeting, Redox Biol., 69:103001. doi: 10.1016/j.redox.2023.103001, 2023.

~~Dikanov, S. A., and Bowman, M. K.: Cross-Peak Lineshape of Two-Dimensional ESEEM Spectra in Disordered S = 1/2, I = 1/2 Spin Systems, J. Magn. Reson. A 116, 125-128, doi: 10.1006/jmra.1995.1199, 1995.~~

Domratcheva, T., Udvarhelyi, A., and Shahi, A. R. M.:, Computational Spectroscopy, Dynamics, and Photochemistry of Photosensory Flavoproteins, in Flavins and Flavoproteins: methods and protocols, Methods in Mol. Biol., 1146, ed. Weber, S., and Schleicher, Humana Press: New York, 191-228, doi: 10.1007/978-1-4939-0452-5_10, 2014.

Con formato: Fuente: Sin Negrita

Con formato: Fuente: Sin Negrita

Con formato: Inglés (Estados Unidos)

Con formato: Inglés (Estados Unidos)

Con formato: Inglés (Reino Unido)

Edwards, A. M., Structure and General Properties of Flavins, in: Weber, S., Schleicher, E. (eds.), Flavins and Flavoproteins. Methods in Molecular Biology, 1146 (Humana Press, New York), doi: 10.1007/978-1-4939-0452-5, 2014.

Fraaije, M. W., Mattevi, A.: Flavoenzymes: diverse catalysts with recurrent features, Trend. Biochem. Sci., 25, 3, 126-132, doi: 10.1016/S0968-0004(99)01533-9, 2000.

Frago, S., Martínez-Júlvez, M, Serrano, A., Medina, M.: Structural analysis of FAD synthetase from Corynebacterium ammoniagenes, BMC Microbiol. 8:160, doi: 10.1186/1471-2180-8-160, 2008.

Frago, S., Lans, I., Navarro, J. A., Hervás, M., Edmondson, D. E., De la Rosa, M. A., Gómez-Moreno, C., Mayhew, S.G., and Medina, M.: Dual role of FMN in flavodoxin function: Electron transfer cofactor and modulation of the protein–protein interaction surface, Biochim. Biophys. Acta 1797(2), 262-271, doi: 10.1016/j.bbabio.2009.10.012, 2010.

Fuchs, M. R., Schleicher, E., Schnegg, A., Kay, C. W. M., Törring, J. T., Bittl, R., Bacher, A., Richter, G., Möbius, K., Weber, S.: g-Tensor of the Neutral Flavin Radical Cofactor of DNA Photolyase Revealed by 360-GHz Electron Paramagnetic Resonance Spectroscopy, J. Phys. Chem. B 106, 8885-8890, doi: 10.1021/jp0259869, 2002.

García, J. I., Medina, M., Sancho, J., Gomez-Moreno, C., Mayoral, J. A., Martinez, J. I.: Theoretical Analysis of the Electron Spin Density Distribution of the Flavin Semiquinone Isoalloxazine Ring within Model Protein Environments, J. Phys.Chem. A 106, 4729-4735, doi: 10.1021/jp014696+, 2002.

Gemperle, C., Aebli, G., Schweiger, A., Ernst, R. R.: Phase cycling in pulse EPR, J. Magn. Reson. 88(2), 241-256, doi: 10.1016/0022-2364(90)90181-8, 1990.

Genzor, C. G., Beldarraín, A., Gómez-Moreno, C., López-Lacomba, J. L., Cortijo, M., & Sancho, J.: Conformational stability of apoflavodoxin Protein Sci. 5, 1373−1388, doi: 10.1002/pro.5560050716, 1996.

Ghisla, S. and Massey, V.: Mechanisms of flavoprotein-catalyzed reactions, Eur. J. Biochem. 181, 1, 1-17, doi: 10.1111/j.1432-1033.1989.tb14688.x, 1989.

González-Viegas, M., Kar, R. K., Miller, A. F., Mroginski, M. A., Noncovalent interactions that tune the reactivities of the flavins in bifurcating electron transferring flavoprotein, J. Biol. Chem., 299, 104762, doi: 101016/j.jbc.2023.104762, 2023.

Gromov, I., Shane, J., Forrer, J., Rakhmatoullin, R., Rozentzwaig, Y., and Schweiger, A.: A Q-band pulse EPR/ENDOR spectrometer and the implementation of advanced one- and two-dimensional pulse EPR methodology, J. Magn. Reson., 149, 196–203, doi: 10.1006/jmre.2001.2298 , 2001.

Guerriere, T. B., Vancheri, A., Ricotti, I., Serapian, S. A., Eggerichs, D., Tischler, D., Colombo, G., Mascotti, M. L., Fraaije, M. W., Mattevi, A., Dehydrogenase versus oxidase function: the interplay between substrate binding and flavin microenvironment, ACS Catal. 15, 1046-1060, doi: 10.1021/acscatal.4c05944, 2025.

Tsukuno, H., Ozeki, K., Kobayashi, I., Hisatomi, O., Mino, H., Flavin-Radical Formation in the Light-Oxygen-Voltage-Sensing Domain of the Photozipper Blue-light Sensor Protein, J. Phys. Chem. B., 122(38), 8819-8823, doi: 10.1021/acs.jpcb.8b05808, 2018.

Höfer, P. J.: Distortion-Free Electron-Spin-Echo Envelope-Modulation Spectra of Disordered Solids Obtained from Two-Dimensional and Three-Dimensional HYSCORE Experiments, J. Magn. Reson., Ser. A, 111, 77-86, doi: 10.1006/jmra.1994.1228, 1994.

Con formato: Fuente: Sin Negrita, Inglés (Estados Unidos)

Con formato: Fuente de párrafo predeter.

Con formato: Inglés (Estados Unidos)

Con formato: Sin subrayado, Color de fuente: Automático, Inglés (Estados Unidos)

Con formato: Inglés (Estados Unidos)

Hore, P. J. and Mouritsen, H.: The Radical-Pair Mechanism of Magnetoreception, Annu. Rev. Biophys. 45, 299-344, doi: 10.1146/annurev-biophys-032116-094545, 2016.

Iyanagi, T., Structure and function of NADPH-cytochrome P450 reductase and nitric oxide synthase reductase domain, Biochem. Biophys. Res. Commun. 338, 520-528, doi: 10.1016/j.abb.2005.05.027, 2005.

Kay, C. W. M., Bittl, R., Bacher, A., Richter, G., Weber, S., Unambiguous Determination of the g-Matrix Orientation in a Neutral Flavin Radical by Pulsed Electron-Nuclear Double Resonance at 94 GHz, J. Am. Chem. Soc., 127, 10780-10781, doi: 10.1021/ja051572s, 2005.

Kaya, S. G., Hovan, A., Fraaije, M. W.: Engineering of LOV-domains for their use as protein tags, Arch. Biochem. Biophys., 763, 110228, doi: 10.1016/j.abb.2024.110228, 2025.

Lans, I., Frago, S., Medina, M.: Understanding the FMN cofactor chemistry within the Anabaena Flavodoxin environment, Biochim. Biophys. Acta, 1817(12):2118-27, doi: 10.1016/j.bbabio.2012.08.008, 2012.

Leys, D., Scrutton, N. S.: Sweating the assets of flavin cofactors: new insight of chemical versatility from knowledge of structure and mechanism, Curr. Opin. Struct. Biol. 41, 19-26, doi: 10.1016/j.sbi.2016.05.014, 2016.

Martínez, J. I., Alonso, P. J., Góomez-Moreno, C., Medina, M.: One- and two-dimensional ESEEM spectroscopy of flavoproteins, Biochemistry 36, 15526-15537, doi: 10.1021/bi971495g, 1997.

Martínez, J. I. , Alonso, P. J., Medina, M.:The elctronic structure of the neutral isoalloxazine semiquinine within Anabaena flavodoxin: New insights from HYSCORE experiments, J. Magn. Res. 218, 153-162, doi:10.1016/j.jmr.2012.02.011, 2012.

-Martínez, J. I., Alonso, P. J., Garcia-Rubio, I., Medina, M.: Methyl rotors in flavoproteins, Phys. Chem. Chem. Phys. 16, 26203-26212, doi: 10.1039/C4CP03115F,2014.

Martínez, J. I., Frago, S., Lans, I., Alonso, P. J., Garcia-Rubio, I., Medina, M.: Spin Densities in Flavin Analogs within a Flavoprotein, Biophys. J. 110, 561-571, doi: 10.1016/j.bpj.2015.11.3525, 2016.

~~Matysik, J., Gerhards, L., Theiss, T., Timmermann, L., Kurle-Tucholski, P., Musabirova, G., Qin, R., Ortmann, F., Solov'yov, I. A. and Gulder, T.: Spin Dynamics of Flavoproteins, Int. J. Mol. Sci., 24(9), 8218, doi: 10.3390/ijms24098218, 20213.~~

Medina, M., Lostao, A., Sancho, J., Cammack, R., Alonso, P. J., Martinez, J., I.: Electron-Nuclear Double Resonance and Hyperfine Sublevel Correlation Spectroscopic Studies of Flavodoxin Mutants from Anabaena sp. PCC 7119, Biophys. J. 77, 1712-1720, doi: 10.1016/S0006-3495(99)77017-7, 1999.

Mayhew, S. G., Ludwig, M. L., Flavodoxins and electron-transferring flavoproteins, Enzymes, 12, 57-118, doi: 10.1016/S1874-6047(08)60225-5, 1975.

Minjárez-Sáenz, M., ,Martínez-Júlvez, M., Yruela, I., Medina, M.: Mining the Flavoproteome of Brucella ovis, the Brucellosis Causing Agent in Ovis aries, Microbiol Spectr. 27, 10(2):e0229421, doi: 10.1128/spectrum.02294-21, 2022.

Minjárez-Sáenz, M., Rivero, M., Correa-Pérez, V., Boneta, S., Suárez, P., Polo, V., Sadeghi, S.J., Yruela, I., Martínez-Júlvez, M., Medina, M., Structural and functional insights into UDP-N-acetylglucosamine-enolpyruvate reductase (MurB) from Brucella ovis, Arch Biochem Biophys, 765:110288. doi: 10.1016/j.abb.2025.110288, 2025.

Con formato: Inglés (Estados Unidos)

Con formato: Normal, Espacio Después: 0 pto, Interlineado: sencillo

Con formato: Fuente de párrafo predeter., Inglés (Reino Unido)

Mohamed-Raseek, N., Miller, A. F., Contrasting roles for two conserved arginines: Stabilizinf flavin semiquinone of quaternary structure, in bifurcatind electron transfer flavoproteins, J. Biol. Chem. 298, 101733, doi: 10.1016/j.jbc.2022.101733, 2022.

~~Moreno, A. Quereda-Moraleda, I., Lozano-Vallhonrat, C., Buñuel-Escudero, M., Botha, S., Kupitz, C., Lisova, S., Sierra, R., Mariani, V., Schleissner, P., Gee, L. B., Dörner, K., Schmidt, C., Han, H., Kloos, M., Smyth, P., Valerio, J., Schulz, J., de Wijn, R., Melo, D.V.M., Round, A., Trost, F., Sobolev, E., Juncheng, E., Sikorski, M., Bean, R., Martínez-Júlvez, M., Martin-Garcia, J. M., Medina, M.: New insights into the function and molecular mechanisms of Ferredoxin-NADP+ reductase from Brucella ovis, Arch. Bioch. Biophys. 762, 110204, doi: 10.1016/j.abb.2024.110204, 2024.~~

Nohr, D, Weber, S., and Schleicher, E., EPR spectroscopy on flavin radicals in flavoproteins, Methods in Enzimol., 620, 251-275, doi: 10.1016/bs.mie.2019.03.013, 2019.

Okafuji, A., Schnegg, A., Schleicher, E., Mobius, K., Weber, S., G-Tensors of the Flavin Adenine Dinucleotide Radicals in Glucose Oxidase: A Comparative Multifrequency Electron Paramagnetic Resonance and Electron-Nuclear Double Resonance Study, J. Phys. Chem. B, 112, 3568-3574, doi: 10.1021/jp077170j, 2008.

Paulus, B., Illarionov, B., Nohr, D., Roellinger, G., Kacprzak, S., Fischer, M., Weber, S., Bacher, A., Schleicher, E., One Protein, Two Chromophores: Comparative Spectroscopic Characterization of 6,7-Dimethyl-8-ribityllumazine and Riboflavin Bound to Lumazine Protein, J. Phys. Chem. B, 118, 13092−13105, doi: ~~/~~10.1021/jp507618f, 2014.

Pompe, N., Illarionov, B., Fischer, M., Bacher, A., Weber, S., Completing the Picture: Determination of ^{13}C Hyperfine Coupling Constants of Flavin Semiquinone Radicals by Photochemically Induced Dynamic Nuclear Polarization Spectroscopy, J. Phys. Chem. Lett., 13, 5160-5167, doi: 10.1021/acs.jpclett.2c00919, 2022.

Rostas, A., Einholz, C., Illarionov, B., Heidinger, L., Al Said, T., Bauss, A., Fischer, M., Bacher, A., Weber, S., Schleicher, E., Long-Lived Hydrated FMN Radicals: EPR Characterization and Implications for Catalitic Variability in Flavoproteins, J. Am. Chem. Soc. 140, 16521-16527, doi: 10.1021/jacs.8b07544, 2018.

Saleem-Batcha, R., Stull, F., Sanders, J. N., ~~Moore~~Moore, B. S., ~~Palfey~~Palfey, B. A., ~~Houk~~Houk, K. N., Teufel, R., : Enzymatic control of dioxygen binding and functionalization of the flavin cofactor, Proc. Nat. Am. Sci. 115 (19) 4909-4914, doi: 10.1073/pnas.1801189115, 2018.

Schleicher, E., Bittl, R., Weber, S.: New roles of flavoproteins in molecular cell biology: Blue-light active flavoproteins studied by electron paramagnetic resonance FEBS J. 276, 4290-4303, doi: 10.1111/j.1742-4658.2009.07141.x , 2009.

Schleicher, E., Weber, S., Radicals in Flavoproteins, Top. Curr. Chem. 321, 41-66, doi: 10.1007/128_2011_301, 2012.

Schleicher, E., Rein, S., Illarionov, B., Lehmann, A., Al Said, T., Kacprzak, S., Bittl, R., Bacher, A., Fischer, M. and Weber, S.: Selective 13C labelling reveals the electronic structure of flavocoenzyme radicals Sci. Reports 11, 18234, doi: 10.1038/s41598-021-97588-7, 2021.

Schosseler, P., Wacker, T., Schweiger, A.: Pulsed ELDOR detected NMR, Chem. Phys. Lett.,224, 319-324, doi:10.1016/0009-2614(94)00548-6, 1994.

Schweiger, A. and Jeschke, G.: Principles of Pulse Electron Paramagnetic Resonance; Oxford University Press: Oxford, p 289, 2001.

Stoll, S., and Schweiger, A.: EasySpin, a comprehensive software package for spectral simulation and analysis in EPR, J. Magn. Reson. 178(1), 42-55, doi: 10.1016/j.jmr.2005.08.013, 2006.

Con formato: Normal

Con formato: Fuente de párrafo predeter., Inglés (Reino Unido)

Con formato: Inglés (Estados Unidos)

Con formato: Inglés (Estados Unidos)

Con formato: Fuente de párrafo predeter.

Con formato: Fuente de párrafo predeter.

Con formato: Fuente de párrafo predeter.

Con formato: Fuente de párrafo predeter.

Con formato: Fuente de párrafo predeter.

Con formato: Fuente de párrafo predeter.

Sucharitakul. J., Wongnate, T., Chaiyen, P.: Hydrogen Peroxide Elimination from C4a-hydroperoxyflavin in a Flavoprotein Oxidase Occurs through a Single Proton Transfer from Flavin N5 to a Peroxide Leaving Group. J. Biol. Chem. 286, 19, 16900-16909, doi: 10.1074/jbc.M111.222976, 2011.

Tschaggelar, R., Kasumaj, B., Santangelo, M.G., Forrer, J., Leger, P., Dube, H., Diederich, F., Harmer, J., Schuhmann, R., Garcia-Rubio, I., and Jeschke, G.: Cryogenic 35GHz pulse ENDOR probehead accommodating large sample sizes: performance and applications, J. Magn. Reson., 200, 81–87, doi: 10.1016/j.jmr.2009.06.007, 2009.

Villanueva, R., Romero-Tamayo, S., Laplaza, R., Martínez-Olivan, J., Velázquez-Campoy, A., Sancho, J., Ferreira, P., Medina, M., Redox- and Ligand Binding-Dependent Conformational Ensembles in the Human Apoptosis-Inducing Factor Regulate Its Pro-Life and Cell Death Functions, Antioxid Redox Signal. 20;30(18):2013-2029. doi: 10.1089/ars.2018.7658, 2019.

Visitsatthawong, S., Chenprakhon, P., Chaiyen, P. and Surawatanawong, P.: Mechanism of Oxygen Activation in a Flavin-Dependent Monooxygenase: A Nearly Barrierless Formation of C4a-Hydroperoxyflavin via Proton-Coupled Electron Transfer, J. Am. Chem. Soc. 137(29), doi: 10.1021/jacs.5b04328, 2015.

Walsh, C. T. and Wencewicz, A. W.: Flavoenzymes: Versatile catalysts in biosynthetic pathways, Nat. Prod. Rep. 30, 1, 175-200, doi: 10.1039/c2np20069d, 2013.

Weber, S., Möbius, K. Richter, G., Kay, C. W. M.: The Electronic Structure of the Flavin Cofactor in DNA Photolyase, J. Am. Chem. Soc. 123, 3790-3798, doi: 10.1021/ja003426m, 2001.

Weber, S., Kay, C. W. M.. Bacher, A., Richter, G., Bittl, R.: Probing the N(5)-H Bond of the Isoalloxazine Moiety of Flavin Radicals by X- and W-Band Pulsed Electron–Nuclear Double Resonance, ChemPhysChem 6, 292-299, doi: 10.1002/cphc.200400377, 2005.

Young, D. N., Hakopian, S., Tam, T. K., Yu, X., Hille, R., Blaha, G. M.: Crystallographic and kinetic analyses of the FdsBG subcomplex of the cytosolic formate dehydrogenase FdsABG from Cupriavidus necator, J. Biol. Chem. 295, 19, 6570-6585, doi: 10.1074/jbc.RA120.013264, 2020.

Con formato: Sin subrayado, Color de fuente: Automático, Inglés (Estados Unidos)