1   **3D-printed microcell for protein NMR at high ionic strengths**

2   **and small sample volumes**

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19   **Abstract.** Standard solution NMR measurements use 5-mm outer diameter (OD) sample

20   tubes that require *ca* 0.5 mL of solvent to minimize "end effects" on magnetic field

21   homogeneity in the active volume of the sample. Shigemi cells reduce the solvent

22   requirement to *ca* 0.29 mL. At high ionic strength, or at ultrahigh magnetic fields, smaller

23   OD samples are needed to study samples in conductive, radiofrequency absorbing solvents

24   such as water. We demonstrate an effective and inexpensive alternative for reducing the

25   active sample volume to 0.13 mL by 3D printing of ellipsoidal shaped cells that are inserted

26   into 5-mm OD NMR tubes. Static magnetic susceptibility, χ, of printer resin was measured

27   using a simple slice-selection pulse sequence. We found that the χ of water increases

28   linearly with NaCl concentration, from -9.05 ppm to -8.65 ppm for 0 to 2 M NaCl. The χ

29   of D_2O was measured to be -9.01 ppm. The susceptibility difference between the resin ($\chi=$

30   -9.40 ppm) and water can be minimized by paramagnetic doping of the resin. Such doping

31   was found unnecessary for obtaining high quality protein NMR spectra when using

32   ellipsoidal shaped cells that are insensitive to susceptibility mismatching. The microcells

33   offer outstanding RF and good B_o homogeneities. Integrated 600-MHz HSQC signal

34   intensities for the microcell sample in PBS buffer were 6.5±4% lower than for 0.5 mL of

35   the same protein solution in a regular 5-mm sample tube. The cell is demonstrated for N-

36   acetylated α-synuclein in PBS buffer, and for observing tetramerization of melittin at 2 M

37   NaCl.

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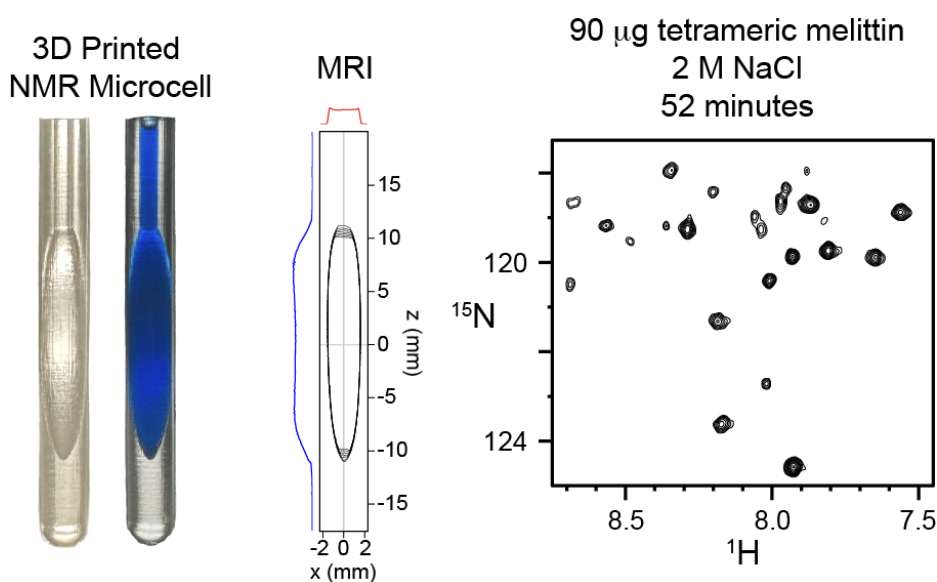
1 Introduction

Nearly all solution NMR measurements are carried out using standard 5-mm outer diameter (OD) sample tubes that require *ca* 0.5 mL of solvent to minimize "end effects" on magnetic field homogeneity in the active volume of the sample. Use of a Shigemi cell, which consist of glass with a magnetic susceptibility that is close to that of the selected solvent, can reduce the amount of sample required and thereby limit the cost of expensive protein preparations. However, the tubes are expensive and need to be matched to the magnetic susceptibility of the NMR solvent. The latter can be challenging considering that the susceptibility of water is somewhat temperature-dependent (Schenck, 1996) and increases substantially (becomes less negative) upon the addition of NaCl, whereas it increases by *ca* 0.04 ppm in D_2O versus H_2O (see below). The sample volume also can be reduced by inserting plugs, of a magnetic susceptibility close to that of the solvent, above and below the active volume of the sample (Barbara, 2009).

Magnetic susceptibility measurements of ionic solutions are often carried out using a magnetic field that oscillates at frequencies ranging from ~50 Hz to low MHz (Tsukada et al., 2006;Gutiérrez-Mejía and Ruiz-Suárez, 2012). However, for magnetic resonance purposes it is the magnetic susceptibility measured in a static magnetic field that is relevant to distortions in field homogeneity (Sangal et al., 2023) and to our efforts to develop a small volume sample cell for protein NMR.

We describe a simple method for measuring magnetic susceptibility of solid material in a high-resolution NMR spectrometer, and the development of a 3D printed microcell that can be inserted into a regular 5-mm NMR tube. The resin that we used for 3D printing at high resolution (25 μm) has a reported magnetic susceptibility of $\chi= -9.34$ ppm (Sangal et al., 2023), which is well below that of water. However, by printing the cell with a spherical (Hizawa et al., 2017) or ellipsoidal geometry, magnetic field homogeneity within the cell becomes insensitive to the susceptibility mismatch between solvent and the printer resin (Schenck, 1996;VanderHart, 1996). The shape of our cell is perturbed by a narrow diameter, cylindrical access port that is needed to fill it with the NMR sample. Magnetic susceptibility mismatching effects, resulting from the deviation of a perfect ellipsoidal shape caused by this access port and by the finite resolution of the printer, can be minimized by paramagnetic doping of the printer resin with organic paramagnetic salt (Evans, 1959)

72 but is found unnecessary for routine applications. The cells are reusable, but the cost of

73 printing such cells is minimal and recycling them therefore may not be necessary.

74 We note that susceptibility mismatching for non-ellipsoidal shapes can also be obtained by

75 introducing nearby suitably shaped small "compensation structures" of different magnetic

76 susceptibility, that effectively shim the sample to homogeneity for microfluidic

77 applications (Ryan et al., 2014). Alternatively, the magnetic susceptibility of an aqueous

78 solution can be increased by addition of Eu^{3+}-complexed diethyl-triamine pentaacetate

79 (DTPA), which due to the short Eu^{3+} electron T_1 has minimal broadening effects on other

80 solutes (Hale et al., 2018) but is restricted to cases where the solvent susceptibility is more

81 negative than the surrounding material, which does not apply for 3D printer resin.

82 The microcell introduced by us for biological NMR spectroscopy purposes is particularly

83 useful when the available sample quantity is limited, or when high ionic strength is

84 required. High ionic strength lowers the quality factor (Q) of radiofrequency (RF) coils,

85 and thereby negatively impacts NMR sensitivity, an effect that scales steeply with

86 frequency (Ugurbil, 2018). RF penetration of water has been studied extensively for

87 applications to magnetic resonance imaging, where it impedes the observation of tissue far

88 from the body surface (Roschmann, 1987). RF absorption at high and ultrahigh magnetic

89 fields also challenges solution ^{1}H NMR spectroscopy, where the use of pulses that are

90 compensated for both offset and RF inhomogeneity (Freeman et al., 1980;Xia et al., 2017)

91 becomes essential, in particular for the vast majority of advanced experiments that include

92 multiple 180° pulses (Manu et al., 2023). Use of the 3D printed microcell greatly reduces

93 problems with probe detuning, lowering of Q, and RF absorption. Consequently, use of

94 the microcell results in short ^{1}Hpulse widths with superior RF homogeneity, even at

95 elevated ionic strength.

96 Magnetic susceptibilities of materials with application to magnetic resonance imaging

97 (MRI) as well as microfluidic NMR technology commonly have employed MRI scanner

98 equipment, and elegant methods applied to a wide range of materials were presented by

99 Wapler et al. (Wapler et al., 2014). The same approach was recently used to measure

100 magnetic susceptibility of 3D printed materials (Sangal et al., 2023), we demonstrate

101 simple methods for deriving magnetic susceptibility differences between ionic solutions

102 and printer material in a high-field solution NMR magnet. We also demonstrate that while

103   using only 130 μL of solvent, the sensitivity in common multi-dimensional NMR

104   experiments, such as the gradient-enhanced HSQC (Kay et al., 1992), is comparable to

105   what is obtained on a regular 500-μL sample. In another application, we show that high-

106   quality spectra can be obtained for 90 μg of recombinantly expressed and chemically

107   amidated melittin at 2 M NaCl concentration. Milligram-scale expression and purification

108   of this uniformly ^{15}N-enriched peptide in its post-translationally modified state, which

109   tetramerizes in a salt-dependent manner, is expensive and very labor-intensive (Gelenter

110   and Bax, 2023).

111

112   **2 Results and Discussion**

113

114   2.1 Measurement of magnetic susceptibility

115   For all materials pertinent to high-resolution solution NMR, the induced magnetization **M**

116   depends linearly on the applied magnetic field **H$_o$**:

117    $\mathbf{M} = \chi \mathbf{H_o}$ (1)

118   where χ is the volume magnetic susceptibility, often expressed in units of parts per million

119   (ppm), i.e. χ is negative for diamagnetic media and positive for paramagnetic substances.

120   The magnetic flux density, **B$_o$**, is directly proportional to **H$_o$**:

121    $\mathbf{B_o} = \mu \mathbf{H_o}$ (2)

122   where the magnetic permeability μ corresponds to $\mu = (1+\chi)\mu_0$, and $\mu_0 = 4\pi\ 10^{-7}$ H/m is

123   the permeability of vacuum. In an NMR sample, the magnetic flux density corresponds to

124   the sum of the applied magnetic field **H$_o$** and the integral of the magnetic field contributions

125   from the induced magnetization over all sample volume elements at locations *r* relative to

126   the point of interest (Barbara, 1994). For a cylindrical sample of infinite length, with its

127   axis parallel to a homogeneous applied magnetic field, the integral over all space within

128   the cylinder is uniform across all locations, resulting in

129    $\mathbf{B_o} = \mu_0(1+\chi)\ \mathbf{H_o}$ (3)

130   Magnetic flux is conserved at any interface orthogonal to **H$_o$** between two media with

131   magnetic susceptibilities χ_1 and χ_2. For the example of a Shigemi sample cell, where the

132   bottom segment of the cylindrical tube consists of glass with susceptibility χ_g, and χ_s is the

133   solvent susceptibility above it (Figure 1a), the flux density at the interface is given by

134      $\mathbf{B_o} = \mu_0[1 + (\chi_g + \chi_s)/2]\,\mathbf{H_o}$ (4)

135   Away from the interface, the integrated contributions to $\mathbf{B_o}$ from volume elements below

136   and above the interface depend on the height above the interface as well as the transverse

137   location, with values converging to $\mathbf{B_o} = \mu_0(1 + \chi_s)\mathbf{H_o}$ at distances above the interface that

138   are large relative to the tube inner diameter, D. Therefore, when selectively observing

139   solvent signal with a chemical shift of δ_o from a very thin slice perpendicular to the sample

140   axis at height h above the interface (Figure 1a), a narrow signal at a frequency $\delta(h) = [1 +$

141      $(\chi_g + \chi_s)/2]\delta_o$ is observed for $h \ll D$, with $\delta(h)$ approaching $(1 + \chi_s)\delta_o$ for $h \gg D$, and a

142   complex line shape for slices taken at intermediate values (Barbara, 1994).

143   Hence, for a solution above the solid glass of a Shigemi cell with a total solvent height that

144   is large relative to both D and the height of the receiver coil, the line shape corresponds to

145      $\delta(h)$ contributions ranging from $[1 + (\chi_g + \chi_s)/2]\delta_o$ to $(1 + \chi_s)\delta_o$. In other words, the total

146   width of the line shape at its base corresponds to half the difference between χ_g and χ_s in

147   units of ppm. The above analysis does not take into account the B_0 gradient inside the NMR

148   tube that is caused by the large magnetic susceptibility mismatch at the bottom of the tube.

149   However, when assuming that the magnetic susceptibility of the Shigemi glass is much

150   closer to that of water than air, shimming on a water-filled sample tube inserted to the same

151   depth into the probe can be used to remove this B_0 gradient, to first order.
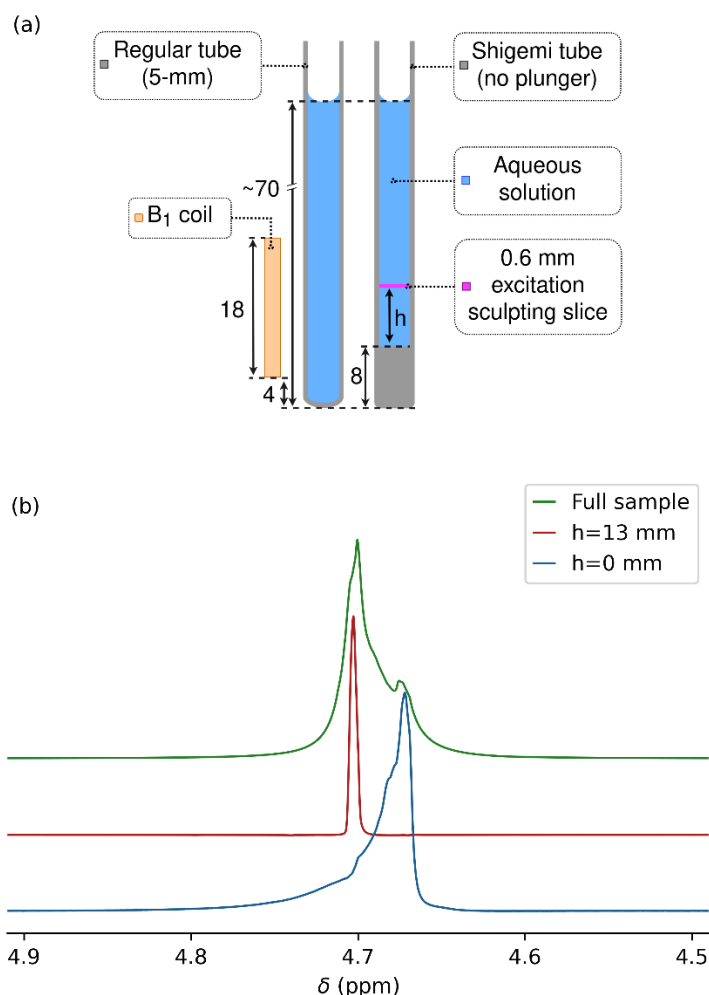
Figure 1. Measurement of magnetic susceptibility in a Shigemi sample tube. (a) Prior to measurements on this sample, the magnetic field homogeneity was optimized for a regular 5-mm NMR sample tube, filled to the same height of 70 mm above the bottom of the tube, and inserted to the same depth into the probehead. Dimensions in mm. A thin slice (pink) through the sample at height h above its bottom was selected by excitation sculpting. (b) HDO resonances for the Shigemi sample tube containing 0.3% H_2O/99.7% D_2O (no plunger). The glass/liquid interface of the Shigemi cell was positioned 5 mm below the center of the receiver coil, using the shim settings of a regular 5-mm sample tube filled to the same total height with the same solution, inserted to the same depth into the NMR probe (see left half of panel a). Overlayed are spectra recorded for the entire sample (green) and for 0.6-mm thickness horizontal slices through the Shigemi cell, centered at $h = 0$ mm (blue), and at $h = 13$ mm (red) by using excitation sculpting while applying z gradients. Spectra were scaled to show the same intensity.

2.2 Measurement of magnetic susceptibility in a high-resolution NMR magnet

167   Careful measurements of solvent susceptibility in a high-resolution magnet that used the

168   gas phase of TMS as an internal reference have been reported by Hoffman (Hoffman,

169   2020;Hoffman, 2022). Here, we describe a different approach that also permits

170   susceptibility measurements of solid objects. First, we demonstrate the method for

171   measurement of the susceptibility of the glass bottom section of a Shigemi cell.

172   Bruker's topshim program was used to minimize magnetic field inhomogeneity of a regular

173   5-mm sample that contained a 1-mL solution of a variable NaCl concentration in 97%

174   D_2O/3% H_2O, that was inserted into the probe to have its bottom ~13 mm below the center

175   of the receiver coil. Then, without changing the shim settings, the HDO line shape on the

176   same solvent composition was observed for a Shigemi sample tube, inserted to the same

177   depth into the probehead. Because the Shigemi tube had an 8-mm bottom segment of solid

178   glass, the flat solvent-glass interface then is located 5 mm below the center of the receiver

179   coil (Figure 1a). If χ_g were identical to χ_s, the same perfect line shape would be expected,

180   but with approximately 15% lower intensity because the bottom ~15% of the receiver coil

181   now was filled with solid glass from the Shigemi tube.

182   For the 97% D_2O/3% H_2O Shigemi tube sample, a pronounced upfield shoulder was

183   observed (Figure 1b, green), indicative of susceptibility mismatching. Excitation sculpting

184   (Stott et al., 1995) (Appendix A1) while applying a pulsed z-gradient was then used to

185   select a slice of 0.6-mm thickness, centered at the glass-solvent interface, i.e. selecting a

186   ~0.3-mm solvent layer just above the interface. A resonance for this layer was observed

187   that was 0.032 ppm upfield from the most intense segment of the solvent obtained with a

188   30° pulse, without slice selection (Figure 1b). When selecting a slice at a height of 13 mm

189   above the interface, the signal (Figure 1b, red) coincided with the maximum of the

190   resonance obtained without slice selection. Therefore, the difference in frequency between

191   the red and blue resonances provides a good measure for the difference between $(\chi_g + \chi_s)/2$

192   and χ_s. The precision at which the frequency at the glass-water interface can be measured

193   is impacted by the slice thickness of the selected solvent layer, which is subject to lateral

194   gradients that increase with slice layer thickness and towards the edges of the slice. By

195   varying the position of the center of the slice from ~0.2 mm below the interface to ~0.2

196   mm above the interface, increased total intensity with a strong downfield shoulder is

197   observed. The upfield edge of this line shape (Figure 1b, blue) remains invariant to the

198   precise position of the selected slice and represents the true $(\chi_g + \chi_s)/2$ value, which can be

199   determined at a precision of ~0.02 ppm.

200   Repeating the same measurement but using 97% H_2O/3% D_2O and strongly mistuning the

201   probehead to reduce radiation damping, showed a shoulder that was 0.0183 ppm closer to

202   the frequency observed 8 mm above the center of the coil (i.e. $h = 13$ mm; Appendix A2).

203   Accounting for the solutions not being fully deuterated or protonated, their difference in

204   static magnetic susceptibility then equals $\chi_{H2O}-\chi_{D2O} = 2\times0.0183\times(100/94) = 0.04$ ppm. The

205   widely used literature value for χ_{H2O} is -9.05 ppm (Sangal et al., 2023), yielding $\chi_{D2O} = $ -

206   9.01 ppm, in fair agreement with Hoffman's measurements (Hoffman, 2022). Using χ_{D2O}

207   = -9.01 ppm as a reference, the susceptibility of the Shigemi glass used in our
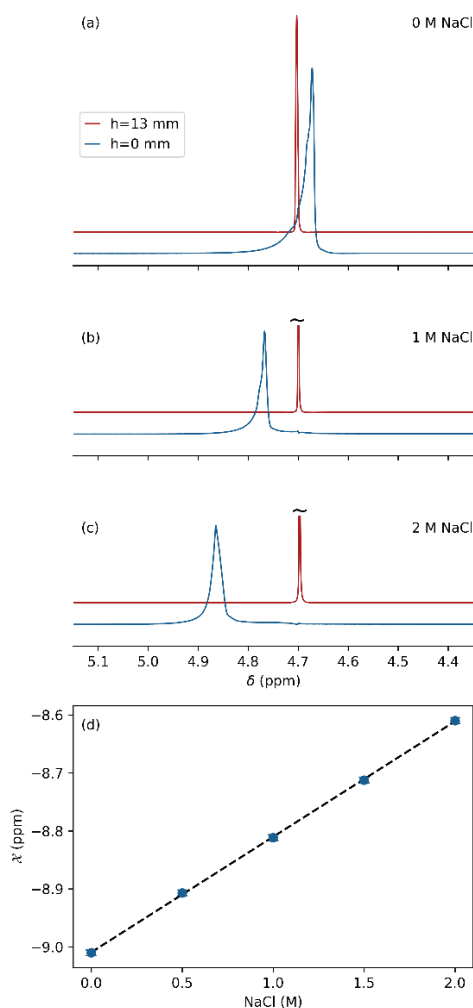
208   measurements is $\chi_g = $ -9.08 ppm.

Figure 2. Effect of salt on magnetic susceptibility of 97% D_2O. Resonances shown correspond to a *ca* 0.6-mm thick slice through a Shigemi tube, centered at h = 0 (blue) and h= 13 mm (red) above the glass bottom of the Shigemi sample cell, using the same protocol as for Figure 1b at (a) 0 mM NaCl; (b) 1.0 M NaCl; (c) 2.0 M NaCl. (d) Magnetic susceptibility of D_2O as a function of NaCl concentration, using χ_{D2O} = -9.01 ppm as a reference. The intensities of the slices at h =0 are upscaled *ca* 16-fold to account for the 50% smaller aqueous volume and the strong magnetic field inhomogeneity in these lower slices.

2.3 Salt dependence of water magnetic susceptibility

The effect of dissolved NaCl on magnetic susceptibility of water is important to protein NMR. We therefore repeated the above measurements of the 97% D_2O/3% H_2O Shigemi tube sample after addition of 0.5, 1, 1.5, and 2 M analytical grade NaCl (Sigma-Aldrich) to the solvent (Figure 2a-c), showing a linear increase in solvent susceptibility with salt

222   concentration over this range: $\chi_{D2O+NaCl} = ([NaCl] \times 0.2 - 9.01)$ ppm, where [NaCl] denotes

223   the concentration molarity in units of moles per liter (Figure 2d).

224   2.4 Susceptibility and paramagnetic doping of 3D printer resin

225   Clear V4 resin was used because it enabled the highest precision (25-μm resolution) of

226   printing on a Formlabs Form3+ 3D printer available in our laboratory, and its optical

227   transparency facilitated sample handling for 3D printed microcells. The susceptibility of

228   the printed Clear V4 resin was measured in the same manner as described above for the

229   Shigemi tube. A cylindrical plug of 2 cm length was printed and pushed to the bottom of

230   a standard 5-mm NMR sample tube that was prefilled with 0.7 mL 1% H_2O/99% D_2O,

231   such that the top of the plug was again 5 mm below the center of the receiver coil once

232   inserted into the magnet. Shimming of the magnetic field was carried out on a sample

233   without the plug, filled to the same level by using ~1 mL of the same solvent, inserted into

234   the probehead at the same depth, i.e. with the bottom of the sample tube at 25 mm below

235   the center of the RF coil.

236   The frequency difference observed at the D_2O/resin interface versus the top of the coil was

237   -0.195 ppm (Figure 3a), corresponding $\chi_{ClearV4} = -9.40$ ppm, which is close to the value

238   of -9.33 ppm measured for this resin by magnetic resonance imaging (Sangal et al., 2023).

239   Considering that a mismatch in magnetic susceptibility between the sample cell and the

240   solution impacts the achievable B_0 homogeneity, and that $\chi_{ClearV4} < \chi_{H2O}, \chi_{D2O}$, increasing

241   the value of $\chi_{ClearV4}$ by paramagnetic doping of the resin in principle allows elimination of

242   this difference. Finding a paramagnetic doping substance that is miscible with the printer

243   resin and does not impact the performance of the 3D printer proved challenging. For

244   example, the use of concentrated $CuCl_2$ in methanol strongly impacted the polymerization

245   kinetics The same problem was encountered for a range of strong paramagnetic chelated

246   substances, such as gadodiamide (Omniscan), which is commonly used in protein

247   paramagnetic relaxation enhancement measurements and as a contrast agent in magnetic

248   resonance imaging. However, the hydrophobic cobalt(II) complex, cobalt(II) bis(2-

249   ethylhexanoate), available from Sigma-Aldrich as a 65 wt. % solution in mineral spirits

250   (product number: 444545), proved miscible with the Clear V4 printer resin without major

251   adverse impact on print quality.

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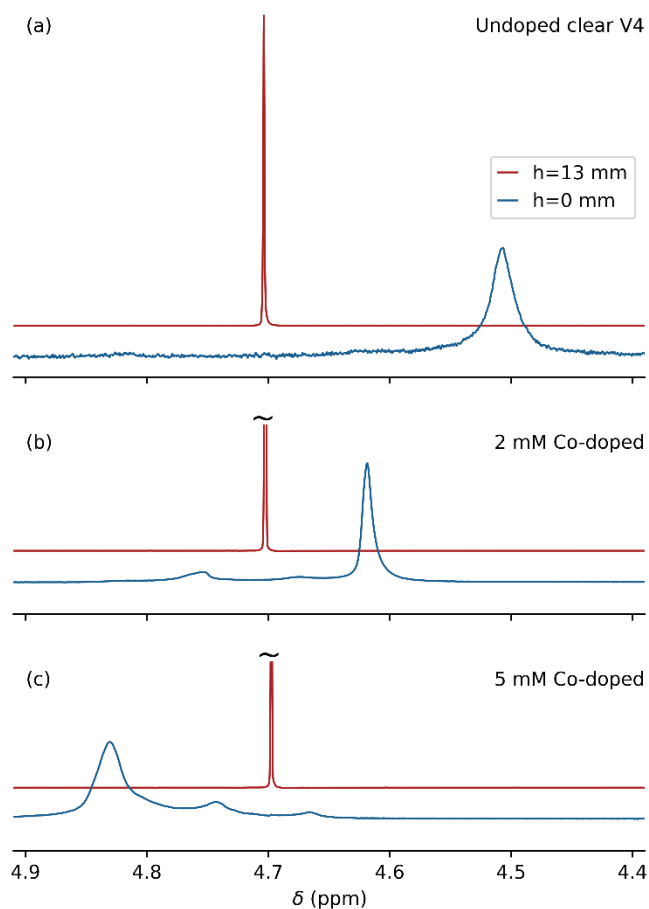
Figure 3. Effect of cobalt(II)2-ethylhexanoate doping on the magnetic susceptibility of printed
Formlabs Clear V4 resin. A solid plug of printed resin that is 2 cm in length and has a 4.0-mm outer
diameter (OD) was inserted into a regular 5mm OD NMR sample tube (Wilmad-507; ID 4.2 mm),
prefilled with 0.7 mL 99%D_2O/1%H_2O. Overlaid spectra are shown from 0.6-mm thickness slices
centered at the top of a Clear V4 plug (blue), and 13 mm above the plug (red). (a) No doping; (b)
2 mM cobalt(II) doping; (c) 5 mM cobalt(II) doping.

259   Comparison of the difference in HDO resonance frequencies obtained for slices at 13 mm

260   above the interface between solvent and printed plug and at the interface for three different

261   levels of the cobalt(II) doping, 0 mM (Figure 3a); 2 mM (Figure 3b) and 5 mM (Figure

262   3c), shows a doping-dependent decrease from +117 Hz to -80 Hz for the NaCl-free 99%

263   D_2O sample. This result indicates that it is possible to match the susceptibility of the solvent

264   to that of the printed resin. However, that would require a large number of printed cells to

265   cover the ionic strength range from 0 to 2 M salt, while also accounting for the difference

266   between D_2O and H_2O samples.

267   In practice, printing sample cells with different levels of cobalt(II) doping is labor-intensive

268   because it requires thorough cleaning of the printing vat used by the Formlabs 3D laser

269   printer. Printing with doped resin requires mixing of the viscous resin at the molecular level

270   with the doping agent and keeping it homogenous during printing. The latter also required

271   some modification of the printer to prevent refreshing of the printer resin with undoped

272   resin from a sealed cassette during printing. We therefore resorted to printing the sample

273   cells with an ellipsoidal shape, that to a good approximation were insensitive to the

274   susceptibility mismatch between solvent and printed resin (Schenck, 1996;VanderHart,

275   1996).

276   2.4 Performance of a Clear V4 ellipsoidal microcell

277   For applications to proteins, we settled on an ellipsoidal microcell design with a volume of

278   130 µL (Figure 4a). A printed access channel of 1.6 mm diameter (measured 1.3 mm) and

279   8.5-mm length and a volume of ~11 µL was used for cleaning of the sample after the initial

280   print and prior to further hardening of the resin in a Formlabs light chamber (see Methods).

281   A subsequent overnight rinse with Milli-Q H_2O at 60 °C was used to remove small water-

282   soluble contaminants. Comparison of the NMR spectrum of these impurities with those in

283   a sample obtained by briefly vortexing a mixture of unpolymerized resin and water

284   followed by entering the aqueous phase into an NMR sample tube (Figure A3) suggests

285   that these impurities consist of residual, methacrylate-based small oligomeric species that

286   apparently can diffuse from the cell walls into the aqueous contents of the cell. Leaving

287   the sample cells filled with H_2O for a week prior to a final rinse reduces the impurity levels

288   further, but in our experience is not necessary considering we have not observed any

289   interaction between these very low impurity concentrations and isotopically enriched

290   proteins. For applications to samples in D_2O, leaving the microcell filled with D_2O for 24

291   h, prior to using it, reduces the intensity of a weak, very broad (~1 kHz) signal at ~3.8 ppm,

292   that results from H_2O diffusing into the resin.

293 Because the digital printer increases the size of printed parts by a small amount due to

294 partial polymerization adjacent to the laser-selected spots, the printed walls of the chamber

295 are actually slightly thicker than designed, and the total volume of the cell including its

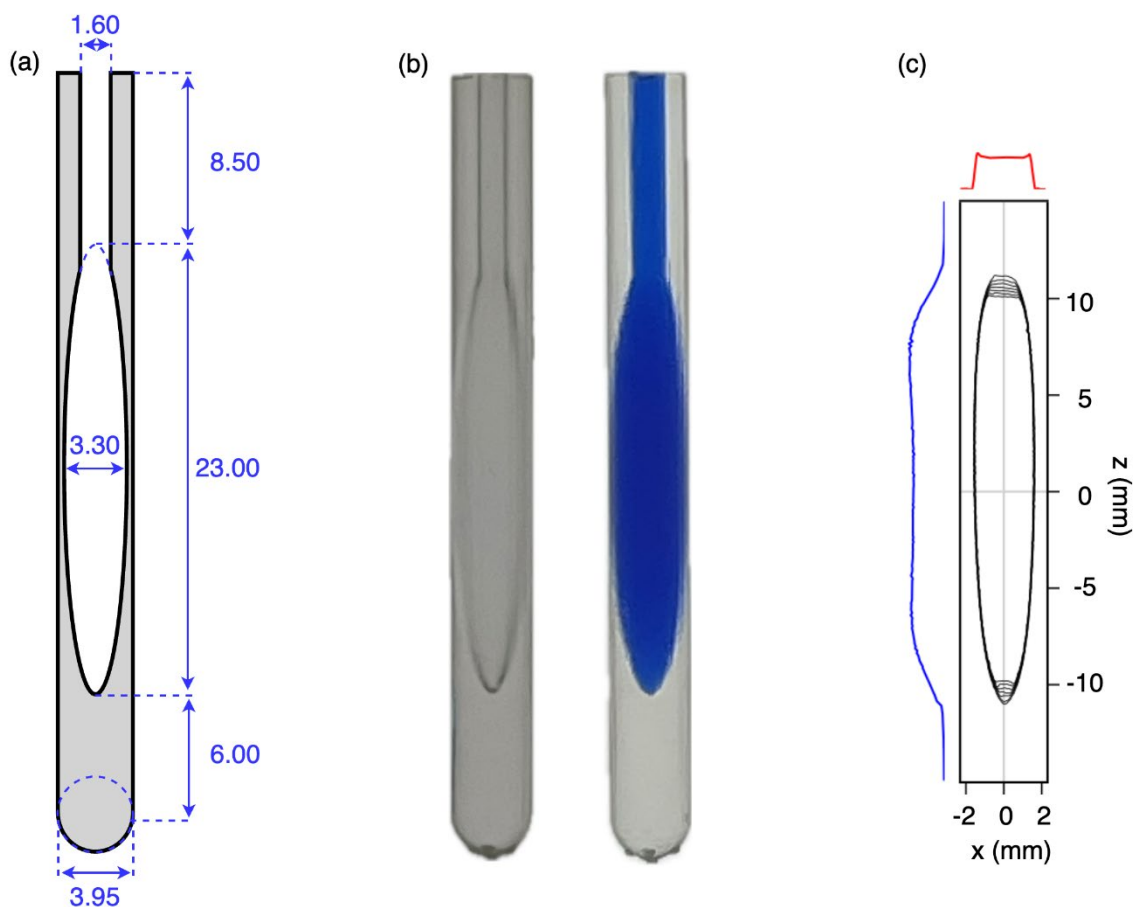296 access channel was measured gravimetrically to be 130 µL (Figure 4b).

297
298
299 **Figure 4.** Images of the printed microcell. (a) Technical drawing; note that the actual dimensions
300 of printed material are slightly larger due to polymerization of a thin (~0.15 mm) sticky surface
301 layer that remains on the printed cell prior to subsequent UV curing. (b) Photographs of the
302 printed cells (left) prior to and (right) after filling with a blue dye solution; (c) Contour plot of a
303 sagittal *xz* cross-section through a 3D image of a 97% D_2O, 3% H_2O sample containing 1.8 mM
304 $CuCl_2$ to shorten the $^1H\,T_1$ value to ~0.7 s. The image was recorded in absorption mode on an 800-
305 MHz NMR spectrometer equipped with a 3-axis pulsed field gradient probehead. The total
306 measurement time was 9 minutes.

307 A sagittal (xz) cross-section through the absorption mode 3D image of the cell, recorded

308 on a Bruker Neo 800-MHz instrument equipped with a 3-axis pulsed field gradient

309 probehead, yielded a shape that matched the ellipsoidal design (Figure 4c) but that did not

310   include the access channel because the solvent in that channel falls outside the RF coil.

311   Small distortions near the bottom of the image correspond to the drop off in RF coil

312   receptivity. Distortions at the top of the sample originate from the access channel which

313   also distorts the ellipse. However, cross-sections taken through the 3D image along the x

314   and z axis through the center of the sample, plotted along the sides of the 3D image, show

315   the expected nearly rectangular shape, indicative of linear imaging gradients.

316   [1]H non-spinning linewidths of ~1 Hz at half height (600 MHz) obtained with the cell

317   (expanded anomeric doublet in Fig.5) were slightly larger than those obtained for a regular

318   5-mm NMR sample in the same probehead. With a width of only ~13 Hz at 0.55% of the

319   HDO peak height the lineshape is also very good, which is most important for protein NMR

320   studies, where protein ^{1}H line widths at half height commonly exceed 10-20 Hz due to fast

321   transverse relaxation and the absence of a "hump" in the water line shape is critical for
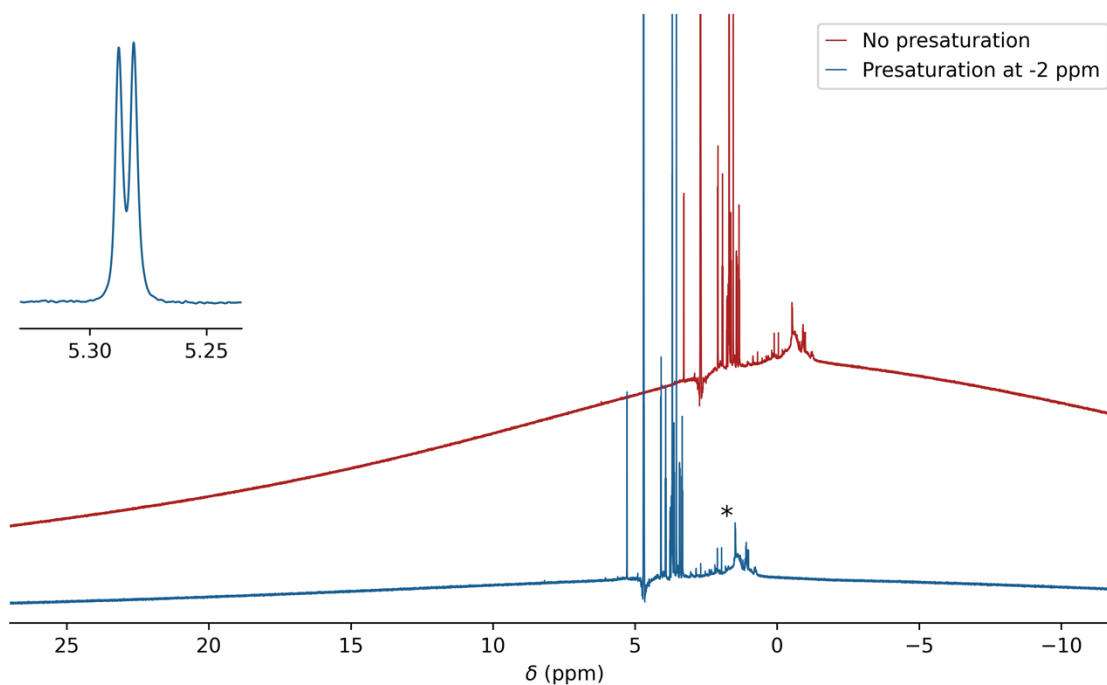
322   good solvent suppression.

323
324   **Figure 5.** [1]H background of printed microcell. The regular ^{1}H NMR spectrum of a 1 mg/mL solution
325   of sucrose in 99% D_2O, recorded without presaturation (red) and with ^{1}H presaturation at -2 ppm
326   (blue), using a 100 Hz RF field strength to suppress the ^{1}H background of the solid resin.
327   Resonances from small impurities released from the printed cell when it has only briefly (~ca 2 h)
328   been rinsed with H_2O are marked with an asterisk. However, a subset of these resonances very
329   slowly reappear over a period of multiple days, at concentrations much lower than seen in the

330   figure (see Figure A3). For display purposes, the not-presaturated spectrum has been shifted
331   upfield by 2 ppm and offset vertically. The inset shows the splitting for the sucrose anomeric
332   doublet at 5.39 ppm.

333

334   We note that sometimes microscopic air bubbles can form inside the cell after filling it with

335   the NMR sample solution. When this happens, it has a very strong adverse impact on both

336   lineshape and linewidth. To eliminate the potential presence of such air bubbles, we briefly

337   (~20 sec) spin the filled sample cell in a speedvac, operating at a pressure of *ca* 150 Pa,

338   and replenish any lost volume by adding a few μL of the protein solution to the access

339   channel.

340   For observation of simple one-dimensional ^{1}H spectra, without echo delays, the protons of

341   the resin yield a strong background signal that is broad due to the rapid transverse

342   relaxation of this solid material. This background signal is readily suppressed by spin echo

343   delays prior to the start of signal acquisition, as are already present in nearly all protein

344   NMR experiments. It can also be effectively reduced by presaturating this broad

345   background with a weak RF field outside the spectral region of interest. Saturation with a

346   100-Hz RF field, applied at -2 ppm in the ^{1}H spectrum, attenuates the background signal

347   by about six-fold (Figure 5).

348   The ^{1}H RF field homogeneity was compared for the microcell and for a Shigemi sample

349   (straight wall) containing 280 μL of phosphate buffered saline (PBS) solution in 99% D_2O.

350   Due to the substantial detuning of the 600 MHz cryoprobe used for this work by the ionic

351   solution in the Shigemi sample, the RF power for this sample was increased by 2.7 dB over

352   the power used for the microcell to yield the same 90° pulse width of ~17 μs. Comparison

353   of the decay of the signal when the excitation pulse is increased from 0.2 to 497.7 μs using

354   the Bruker "paropt" module (Figure 6) shows slightly better RF field homogeneity for the

355   smaller 3D printed microcell than for the Shigemi sample. Notably, the intensity of the

356   signal after a 90° pulse was only ~10% lower for the printed microcell than for the Shigemi

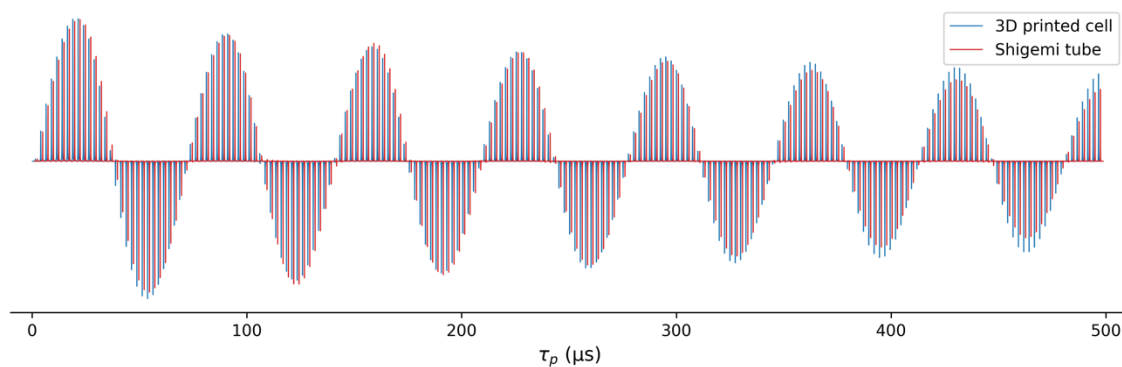357   tube that contained more than double the volume of the same solution.

Figure 6. Comparison of ^{1}H RF field homogeneity in a 280-µL Shigemi sample cell (red) and in the 3D printed 130-µL 23×3.3-mm ellipsoidal microcell (blue). Signal intensity is shown as a function of ^{1}H pulse duration, ranging from 0.2 to 497.7 µs, generated by the Bruker macro 'paropt'. Both samples contained the same solution of PBS buffer in 99% D_2O. The RF power for the microcell sample was adjusted 2.7 dB lower than for the Shigemi sample to equalize the 90° pulse lengths.

2.6 Observation of high-resolution protein NMR spectra

The high resolution and sensitivity obtained with the microcell are illustrated for two proteins, N-acetylated α-synuclein and native C-terminal amidated melittin in its monomeric and tetrameric forms. Complete or nearly complete N-acetylation of α-synuclein is invariably present in mammalian cells (Bartels et al., 2011) and strongly impacts its interaction with phospholipids (Kang et al., 2012;Maltsev et al., 2012). By simultaneously including a plasmid for expressing the NatB complex, needed for acetylation of α-synuclein, together with a plasmid for α-synuclein, fully N-acetylated protein can also be obtained from bacterial expression systems (Johnson et al., 2010). Although this combined expression reduced protein yields in our hands, it enabled the pivotal incorporation of stable isotopes, such as ^{15}N, in the biologically relevant state of the protein.

As can be seen, in *ca* 20 min, a high-quality gradient-enhanced ^{1}H-^{15}N HSQC spectrum was obtained for 130 µg of N-acetylated α-synuclein (14.5 kDa) in PBS buffer, pH 6.5, when using the microcell, which approaches the sensitivity and resolution that was

380 obtained for 500 µg protein in a regular 5-mm sample cell, using a 0.5 mL sample volume

381 (Figure 7).
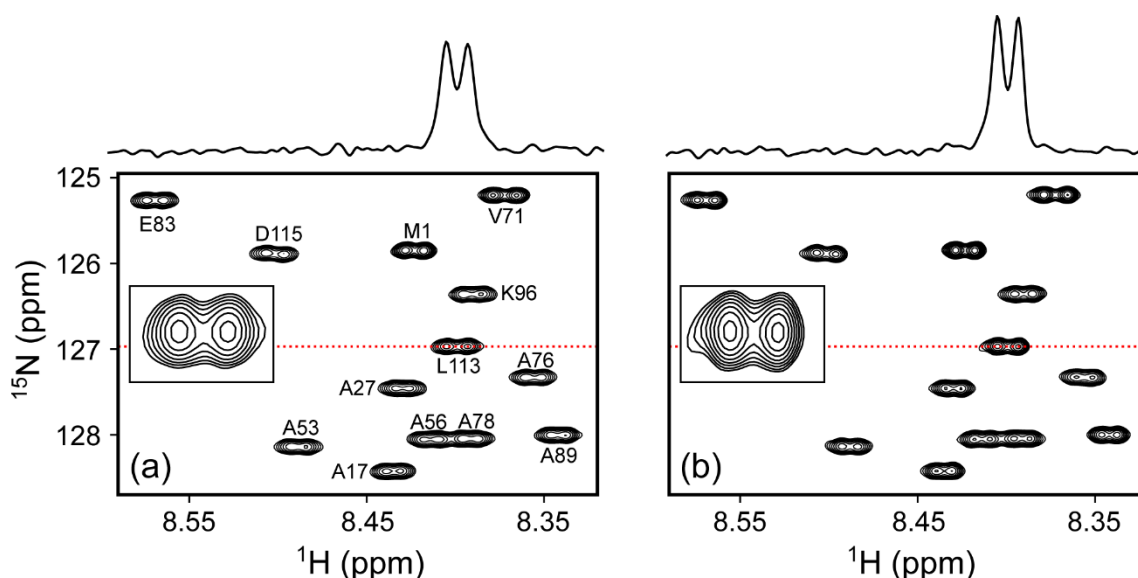
382

383 **Figure 7.** Comparison of small regions of the 600-MHz ^{1}H-^{15}N HSQC spectra of 70 µM N-acetylated
384 α-synuclein at 20 °C in PBS-buffer, pH 6.5. Each spectrum results from 175* × 4000* data points
385 with two transients per FID, for total measuring times of 21 min each. (a) 130 µL in the 3D printed
386 microcell; (b) 500 µL in a standard 5-mm NMR tube. Lowest contours are drawn at the same level
387 above the respective RMS noise. Insets show the expansion of the L113 cross peak, with cross-
388 sections through L113 above the panels at locations marked by the red dotted lines. The
389 comparison shows slightly lower resolution of the ^{1}H^N-^{1}H^α doublets for the microcell, reflected in
390 12% lower S/N versus 6.5% lower peak integrals relative to RMS noise.

391 The utility of the microcell becomes even more compelling for the study of the

392 tetramerization of native melittin, which requires C-terminal amidation. The latter involves

393 a large number of chemical derivatization and purification steps (Gelenter and Bax, 2023),

394 making it very challenging to generate adequate quantities of peptide for driving it to its

395 tetrameric state within a standard 5-mm NMR tube. Tetramerization is promoted by

396 increasing the NaCl concentration, but the tuning of the RF circuitry in cryoprobes often

397 limits salt concentrations to be below *ca* 0.5 M. Here, we demonstrate that the microcell

398 enables observation of the monomer-tetramer equilibrium at salt concentrations of 2 M,

399 and that indeed even at a moderate peptide concentration of 250 µM the peptide is fully

400 tetrameric in the presence of 2 M NaCl (Figure 8). Interestingly, residues L9-R24, close

401 to the center of the peptide, are attenuated by an exchange process that is much less

402 prevalent at low ionic strength and high peptide concentration (Gelenter et al., 2024). We

403 speculate that the line-broadening associated with these weaker resonances arises from the

404 exchange process between the asymmetric arrangement of the two dimers seen in its X-ray

405 tetrameric structure (Terwilliger and Eisenberg, 1982) switching from fast exchange at

406 lower ionic strength to intermediate exchange at 2 M NaCl.

407 Notably, the same microcell was used for the two melittin spectra. After recording of the

408 low-ionic strength spectrum, contents of the microcell was removed and used to dissolve

409 15.2 mg NaCl, prior to insertion of the high ionic strength sample into the original cell,
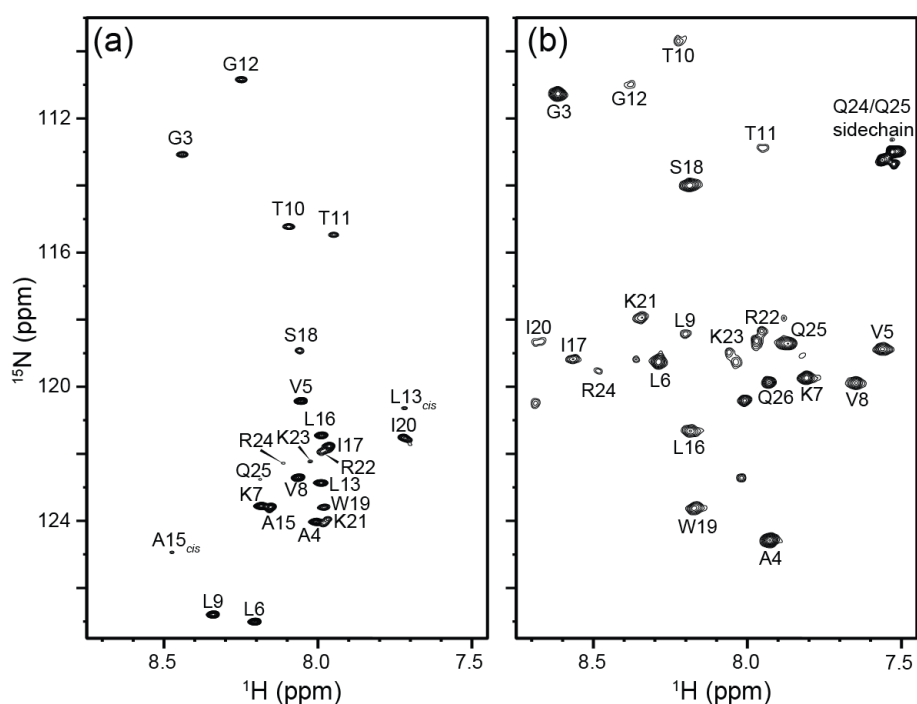
410 thereby demonstrating the recyclability of the tube.

411

Figure 8. 800-MHz ^{1}H-^{15}N HSQC spectra from 90 μg (250 μM) of ^{15}N-labeled native melittin in a 130-μL microcell. Both spectra were collected at 288 K in 10 mM sodium phosphate buffer, pH 7.0, containing 3% D$_2$O. (a) Sample containing no NaCl, where melittin remains an intrinsically disordered monomer. L13$_{cis}$ and A15$_{cis}$ correspond to residues in monomers with P14 in the *cis* conformation. The data was collected with 2 transients per FID, using 200* points in the indirect dimension corresponding to an evolution of 103 ms and a total measurement time of 27 min. (b) 15.2 mg of NaCl was added to the sample from (a) to reach a final concentration of 2 M NaCl. Under these conditions melittin adopts an α-helical tetrameric conformation. The data was collected with 8 transients, using 100* points in the indirect dimension ($t_{1,max}$ = 51 ms) and a total experimental time of 52 min.

3 Concluding remarks

3D printing enables the efficient and relatively inexpensive creation of complex, customized products with minimal waste. It is extensively used for prototyping new designs of items with intricate or complex geometries and has broad impact in science and engineering. 3D printing also enabled the design and construction of high performance solid-state NMR probes, offering similar or improved filling factors due to the coil being in close proximity to the sample, resulting in high RF transmit and receive efficiencies (Long et al., 2021;Pereira et al., 2023).

In solution NMR spectroscopy, 3D-printed bioreactor platforms were introduced that are compatible with low-field NMR spectrometers that accommodate bioengineered 3D cell models (Mangas-Florencio et al., 2025). That work consisted of a bioreactor made of biocompatible materials and included a microfluidic system for optimization of cell culture conditions during the actual NMR data collection process.

The application of 3D printing to high-field solution NMR spectroscopy has remained rather limited, largely due to the requirements of high magnetic field homogeneity and minimal background signals. Our study demonstrates that the homogeneity requirement can be met by printing small sample cells with an ellipsoidal shape. The half-height line widths achievable for our microcell is *ca* 1 Hz and remains limited by the precision at which the cell's surface can be printed. For shimming purposes, we first used Bruker's topshim program to adjust field homogeneity to starting values on a regular solution NMR sample filled to the same height (~40 mm) as the length of the microcell. Subsequently, after entering the microcell into the magnet, we used topshim followed by iterative tuning of z, z^2, x, y, xz and yz gradients. The microcells can easily be recycled as they slide in and out of standard high-quality Wilmad-507, New Era NE-HP5, or Norell Standard Series 5-mm NMR tubes. The access channel of the microcell is sufficiently small that surface tension prevents the aqueous solution from leaving the microcell when the 5-mm NMR tube is fully inverted while the microcell slides out of it.

Although inexpensive, sample cells are easily recycled as highlighted for the melittin sample, where 15.2 mg of salt was added to the initial sample by first removing the sample

451   solution, dissolving the NaCl, and re-injecting the solution into the original cell, all with

452   minimal losses. It is advisable to briefly spin and expose the sample to vacuum to remove

453   dissolved gasses, in particular for lengthy experiments. Formation of even microscopic air

454   bubbles deteriorates homogeneity for the microcell sample more than for the Shigemi or

455   larger conventional NMR sample tubes.

456   The sample cell appears unsuitable for the use of organic solvents which dissolve and

457   release resin components, resulting in strong narrow background signals. Even when using

458   water as the solvent, slowly increasing signals from micromolar quantities of resin-derived

459   small molecules appear in the ^{1}H spectrum over a period of days (marked by an asterisk in

460   Figure 5). However, the standard use of isotope-enriched multi-dimensional multinuclear

461   NMR experiments keeps these resonances well below the signal-to-noise threshold level,

462   and unless such contaminants have a strong interaction with the protein studied they have

463   no effect on the acquired spectra. We note that commercial 3D printing resins are

464   formulated in a way that tries to meet various performance specifications, such as material

465   strength, printing resolution, and printing speed. As such, they may not be optimal for high

466   precision work such as required for protein NMR. Future development of resins is needed

467   to allow high precision printing while minimizing release of small, incompletely

468   polymerized precursors into the aqueous phase.

469   While a small amount of solvent (<0.1%) can diffuse into cavities of the polymerized resin,

470   no detectable loss of solute signal was observed. For example, the intensity of the sucrose

471   NMR resonances remained unchanged, to within ±0.2% over a duration of more than one

472   week.

473   We cleaned the microcells by soaking them overnight at 60 °C in Milli-Q water and

474   subsequent removal of most of the solvent with a standard gel micropipette tip that didn't

475   quite reach the bottom of the cell, followed by upside down centrifugation after insertion

476   into an Eppendorf tube to remove the remainder of the solvent. An additional rinse with

477   130 μL D_2O, followed by centrifugation and vacuum exposure can be used to remove any

478   residual solvent protons if the sample is intended for measurements in highly deuterated

479   D_2O. These are the most labor-intensive steps in preparing the sample cells, but limited

480   quantities of unrinsed sample cells are available upon request.

481

4. Methods

483   4.1 Magnetic susceptibility measurements

484   Selective excitation while applying a 11.5 G/cm (20% on the Bruker Neo-600 instrument)

485   was used for collecting the HDO solvent resonance measurements (Appendix A1) of 0.6-

486   mm thickness slices at various heights, h, above the flat interface between a solid printed

487   plug and the aqueous solvent. The printed plug with outer diameter of 4-mm and a length

488   of 20-mm, including its hemispheric bottom, was inserted and pushed to the bottom of a

489   regular NMR tube, prefilled with 0.7 mL of 97% D_2O or 97% H_2O solution. A similar tube

490   without the plug was filled to the same height with the same solution and used for shimming

491   the magnetic field using topshim prior to inserting the sample with the plug at its bottom,

492   where the same shim settings of the sample without the plug were used. The thickness of

493   the aqueous fraction of the slice collected for h=0 was two-fold smaller than for slices at

494   h>0.3 mm, with a correspondingly lower integrated volume. The frequency of slices

495   collected at heights >~10 mm above the plug became essentially independent of h. The

496   slice collected at h=0 shows extensive line broadening due to the large field gradient at the

497   solvent-plug interface (Figure 2a-c). The difference in ppm frequency was used as a

498   measure for $(\chi_{solvent} - \chi_{resin})/2$.

499   4.2 Printing of the microcell

500   Cells were printed in Clear V4 resin on a Form 3+ printer at 25-micron resolution to achieve

501   a smooth surface finish. The designs were created in OpenSCAD (2021.01) using standard

502   STL export settings, then prepared for printing in PreForm (3.43.2). Models were oriented

503   so that the bottom of each cell faced downward on the build platform, with 0.2 mm

504   touchpoint supports attached only at the bottom of the cells. The cells were washed with

505   15 mL isopropyl alcohol (IPA) at a rate of 5 mL/min through a syringe needle (0.58 mm

506   ID) with a blunt tip, connected to a Fast Protein Liquid Chromatography (FPLC) pump.

507   Cells were kept vertical during printing and subsequent UV-curing using Form Cure for 16

508   hours at 60 °C. Subsequently, cells were filled with Milli-Q water and immersed in a water-

509   filled falcon tube that was heated at 60 °C for 12 hours to remove water-soluble

510   components. After this cleaning, to further minimize the amounts of resin-derived

511   impurities from leaching from the cell wall into the solvent, which occurs at a rate that

512   decreases steadily with time, cells can be stored filled with and immersed in water for a

513   week at room temperature. After removal of the water from the cells by pipetting followed

514   by centrifugation upside down in Eppendorf tubes, they were briefly dried under vacuum.

515   Subsequent use of such a cell showed strongly reduced intensities of these impurity signals

516   (Figure A3).

517   The paramagnetically doped resin was prepared by mixing Formlabs Clear V4 Resin with

518   a cobalt(II) bis(2-ethylhexanoate) solution (65 wt.% in mineral spirits, Sigma-Aldrich,

519   444545) on a shaker at 37 °C and 200 rpm for 30 minutes. The plugs printed from doped

520   resin were washed with IPA and cured at 60 °C for 16 hours using Form Cure.

521

522   4.3 NMR sample preparation

523   The microcells were filled to the top with the sample solution (~130 μL) using a gel-tip

524   pipette and degassed at *ca* 150 Pa pressure for 20-30 seconds using a SpeedVac (Savant,

525   SVC-100-H). After degassing, the cells were topped off with *ca* 2 μL of additional sample

526   solution and inserted into standard Wilmad-507, New Era NE-HP5, or Norell Standard

527   Series 5-mm NMR tubes.

528   4.4 Imaging of the microcell

529   Although MRI normally uses absolute value mode displays, higher resolution absorption

530   mode spectra can also be obtained (Bretthorst, 2008). For generating images of the

531   microcell, we used a very simple one-pulse sequence with variable durations of the x, y,

532   and z gradients for encoding the three spatial dimensions (Appendix A4). The experiment

533   used Rance-Kay quadrature selection (Palmer et al., 1991;Kay et al., 1992) in both the x

534   and y dimensions, by collecting four scans per hypercomplex time domain data point

535   (ns=1). NMRPipe (Delaglio et al., 1995) processing of the 3D time domain matrix was

536   used to generate regular, amplitude-modulated quadrature (States et al., 1982) in both

537   indirect dimensions (see Appendix A5 for NMRPipe processing script). A total of 20*(x)×

538   20*(y) × 512*(z) data points were collected with a total acquisition time of *ca* 9 minutes

539   for a sample that contained 1.8 mM $CuCl_2$ in 99% D_2O.

540

541 **Appendix A: Supplementary data and code**

542

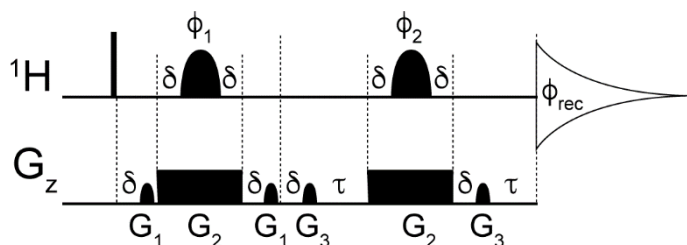543 A1: Pulse diagram used for slice selection

544

545 **Figure A1.** Diagram of the slice selection pulse sequence used to measure magnetic
546 susceptibility. The narrow bar represents the 90 ^{1}H excitation pulse. The shaped pulses ϕ_1 and
547 ϕ_2 have a profile of Reburp (Geen and Freeman, 1991) with a duration of 2 ms and a variable
548 offset frequency, ranging from *ca* +23 to -30 kHz from the water signal resonating at 4.7 ppm.
549 The gradient pulses are either sine-bell shaped or rectangular, with durations of 0.5, 2.1 and
550 0.5 ms and a strength of 2, -11.5 and 3.5 Gauss/cm for G_1, G_2 and G_3, respectively. Delays: δ
551 = 50 μs and τ = 60 μs. The following phase cycling scheme was used: ϕ_1 = y, y, y, y, -x, -x, -x, -x,
552 -y, -y, -y, -y, x, x, x, x; ϕ_2 = x, y, -x, -y; ϕ_{rec} = x, -x, x, -x, -x, x, -x, x. The interscan delay was set to 4
553 s, number of scans to 4, spectral width to 8620 Hz and the acquisition time to 2 s.

554

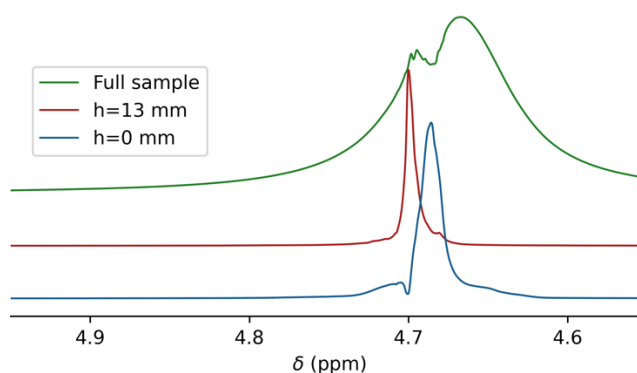555 A2: Spectra used for measuring $\chi_{glass} - \chi_{H2O}$

556
557 **Figure A2.** Same as Figure 1b (main text), but for a solution containing 97% H_2O/3% D_2O. The
558 resonance of the full Shigemi tube sample (green) is broadened by radiation damping and
559 shown together with slices taken at h = 0 mm (blue), and at h=13 mm (red). The center of the
560 green resonance is shifted relative to its regular position due to radiation damping effect in
561 the deliberately mistuned probe (Torchia, 2009).

562

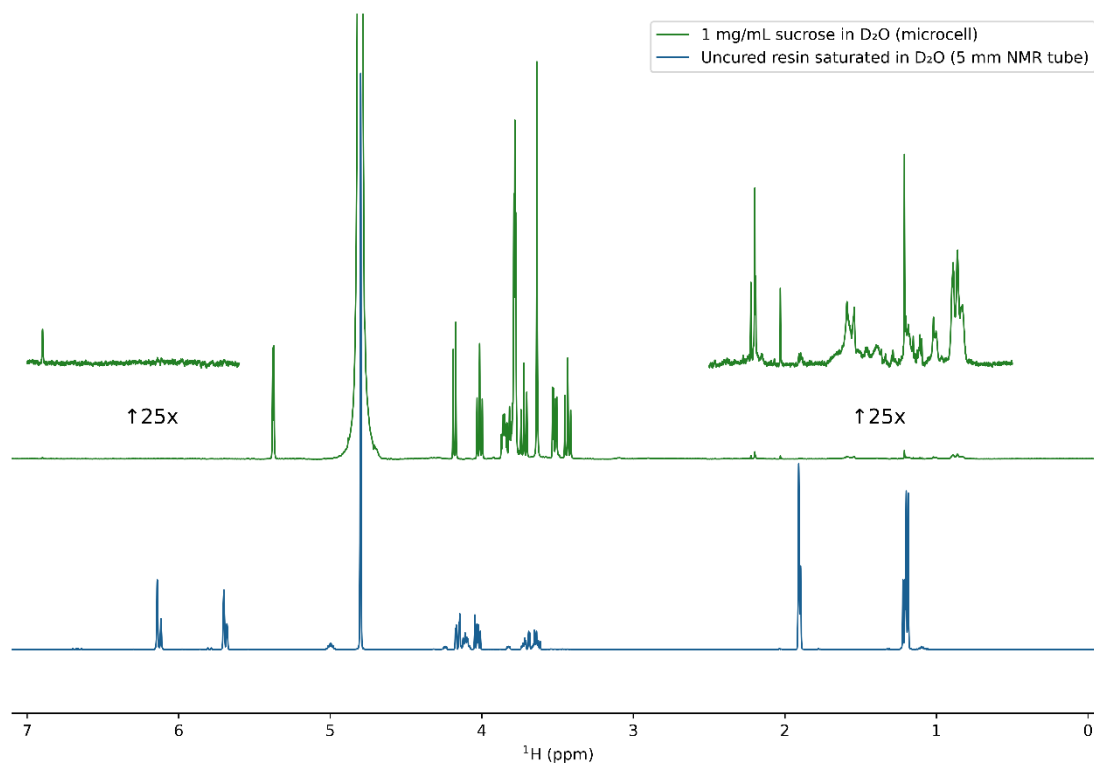## A3: Spectra of polymer precursors and oligomeric impurities

Figure A3. ^{1}H NMR spectra (500 MHz) of liquid Formlabs Clear V4 resin, saturated in D_2O (5 mm tube, 128 scans; blue trace), and 1 mg/mL sucrose in D_2O recorded in a 3D-printed sample cell, 48 h after preparation (512 scans; green trace). The blue spectrum is scaled down 10-fold relative to the green spectrum and corresponds to $ca\,10$ mg/mL precursor concentration. The dissolved oligomer impurity concentration in the 3D-printed microcell sample corresponds to *ca* 24 μg mL^{-1}. Insets correspond to the olefinic and aliphatic regions, upscaled 25-fold. According to the Formlabs Clear V4 datasheet, the monomeric precursors in the blue spectrum correspond to methacrylic acid monoester with propane-1,2-diol and 7,7,9(or 7,9,9)-trimethyl-4,13-dioxo-3,14-dioxa-5,12-diazahexadecane-1,16-diylbismethacrylate (see https://formlabs-media.formlabs.com/datasheets/1801037-SDS-ENEU-0.pdf).

576   A4: Bruker pulse program used for imaging the microcell:

```
577   #include<Avance.incl>
578   #include<Grad.incl>
579   #include<De.incl>
580
581
582   "d11=30m"
583
584   "in0=inf1".   ;125u
585   "in10=inf2"   ;125u
586
587   "d0=0"
588   "d10=0"
589
590   define list<gradient> EA2 = { 1.000 -1.000}
591
592   aqseq 321
593
594
595   1 ze
596     1m
597   2 d11 groff
598     10u BLKGRAD
599     10u pl9:f1                 ;set to 1000dB unless presat is needed
600     10u fq=cnst2(bf ppm):f1   ;chemical shift for the resin to be
601   presat'ed
602     d1 cw:f1
603     10u do:f1
604     10u UNBLKGRAD
605     p20:gp20.     ;3m at 33% of z-grad
606     1m fq=0:f1    ;shift carrier back to be on-resonance with water
607     1m pl1:f1
608    (p1 ph1):f1    ;20 degree flip angle
609     1u gron1*EA   ;x-grad 8%
610     d0
611     1u groff
612
613     1u gron2*EA2  ;y-grad 8%
614     d10
615     1u groff
616     600u          ;delay to dephase a very broad hump
617     1u gron3      ;z-grad -1%
618     go=2 ph31
619     d11 groff mc #0 to 2
620     F1EA(calgrad(EA), caldel(d0))    ;TD1=40
621     F2EA(calgrad(EA2), caldel(d10))  ;TD2=40
622     d11 BLKGRAD
623   exit
624
625
626   ph1=0
627   ph31=0
```

A5: NMRPipe conversion and processing scripts

```
#!/bin/csh

bruk2pipe -verb -in ./ser \
  -bad 0.0 -ext -aswap -DMX -decim 1240 -dspfvs 20 -grpdly 68 -ws 8 -
noi2f  \
  -xN              1024  -yN              40  -zN              40
\
  -xT               512  -yT              20  -zT              20
\
  -xMODE            DQD  -yMODE  Echo-AntiEcho  -zMODE  Echo-AntiEcho
\
  -xSW        16129.032  -ySW        8000.000  -zSW        8000.000
\
  -xOBS         800.134  -yOBS         800.134  -zOBS         800.134
\
  -xCAR           4.821  -yCAR           4.821  -zCAR           4.821
\
  -xLAB              Hz  -yLAB              Hy  -zLAB              Hx
\
  -ndim               3  -aq2D         Complex
\
| pipe2xyz -x -out ./fid/test%03d.fid -ov


xyz2pipe -in fid/test%03d.fid -x                      \
| nmrPipe  -fn SP -off 0.5 -end 0.98 -pow 2 -c 0.5   \
| nmrPipe  -fn ZF -zf 2 -auto                         \
| nmrPipe  -fn FT                                     \
| nmrPipe  -fn PS -p0 149 -p1 38 -di                  \
| nmrPipe  -fn TP                                     \
| nmrPipe  -fn SP -off 0.5 -end 0.98 -pow 1 -c 0.5   \
| nmrPipe  -fn ZF -zf 2 -auto                         \
| nmrPipe  -fn FT                                     \
| nmrPipe  -fn PS -p0 135 -p1 0 -di                   \
| nmrPipe  -fn ZTP                                    \
| nmrPipe  -fn SP -off 0.5 -end 0.98 -pow 1 -c 0.5   \
| nmrPipe  -fn ZF -zf 2 -auto                         \
| nmrPipe  -fn FT                                     \
| nmrPipe  -fn PS -p0 90 -p1 0 -di                    \
| nmrPipe  -fn TP                                     \
| nmrPipe  -fn POLY -ord 4 -nw 300 -nl 301 1740       \
| pipe2xyz -out ft/test%04d.ft3 -x -ov -verb
```

Author contributions

TK: conceptualization (equal), 3D printing and cleaning, figures 1-6, review and editing (equal). MDG: review and editing (equal), data collection (equal), figure 8. JY: review and editing (equal), data collection (equal), figure 7. AB: conceptualization (equal), writing original draft, review and editing (equal), recording data (equal).

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Competing interests

AB is a member of the editorial board of *Magnetic Resonance*. The peer-review process was guided by an independent editor, and the authors also have no other competing interests to declare.

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